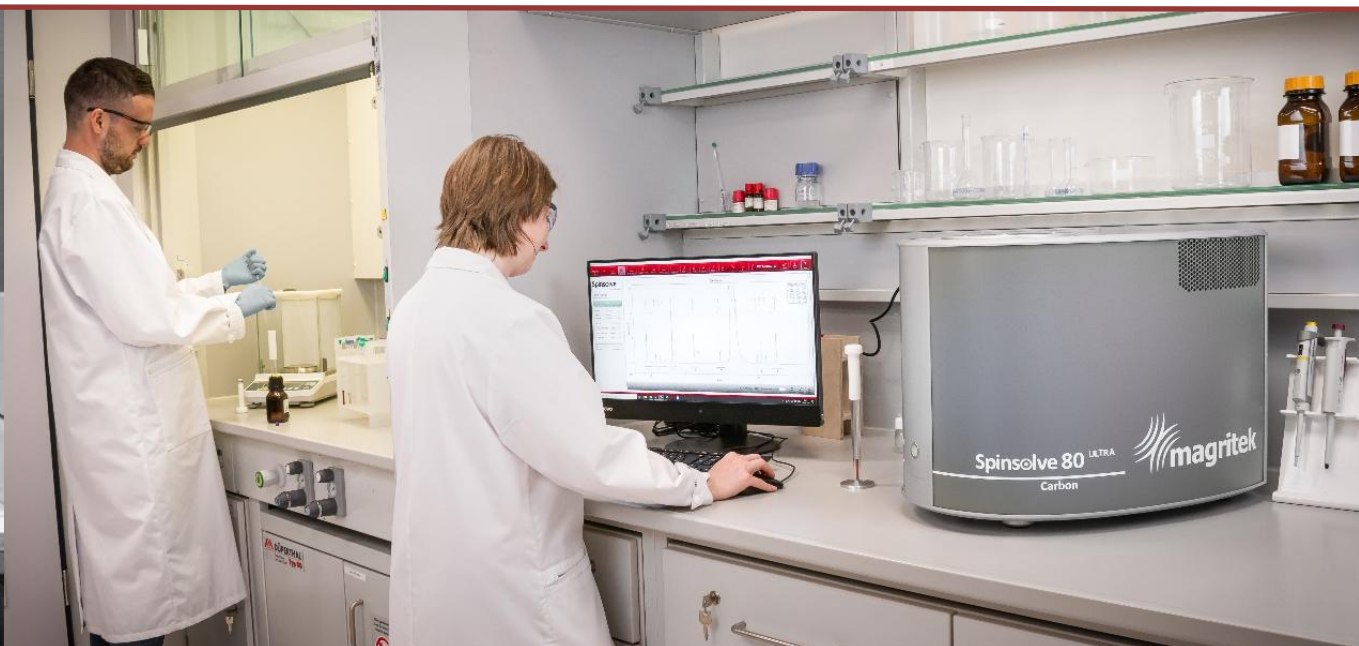


Applications of benchtop NMR: organic chemistry, polymer chemistry, teaching, and many others

Thursday 9th of April 2026, 10h00



Dr. Hélène Freichels

Application scientist– Magritek GmbH

Helene@magritek.com



<https://www.linkedin.com/company/magritek-ltd/>



<https://X.com/magritek>

Magritek - Worldwide leader in Benchtop NMR Spectroscopy

- Founded in 2004 – operating for more than 20 years
- Introduced Spinsolve Benchtop NMR in 2012
- Large base of customers (> 2500), more than 600 publications

Aachen (Cáchy)



Our production site in Aachen, Germany

The Spinsolve Journey

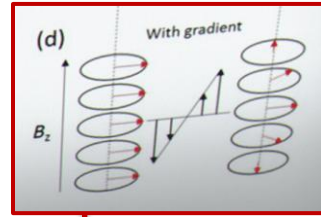
2013
Reaction
monitoring



2014
Spinsolve
Double channel



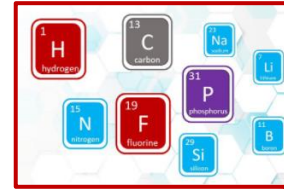
2015
Spinsolve Diffusion



2018
Autosampler



2021
Spinsolve Multi X



2025
Spinsolve Multi Xⁿ



Advanced Autosampler



2026

2012



Spinsolve

2016



Spinsolve™ 60

2017



Spinsolve 80

2021



Spinsolve™ 90

2025





Spinsolve^{STC}





Spinsolve 100

Spinsolve features (for all models)

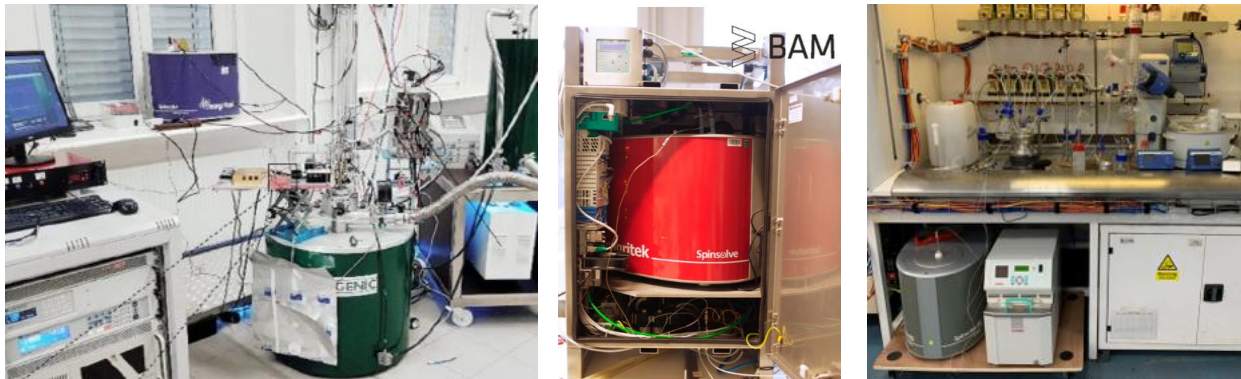
 Benchtop NMR spectrometers designed to work in **ANY** lab: easily moved next to the reactor or under the fumehood

 Room temperature variation robust (18-28°C)
No stray field outside of the box (<2G)

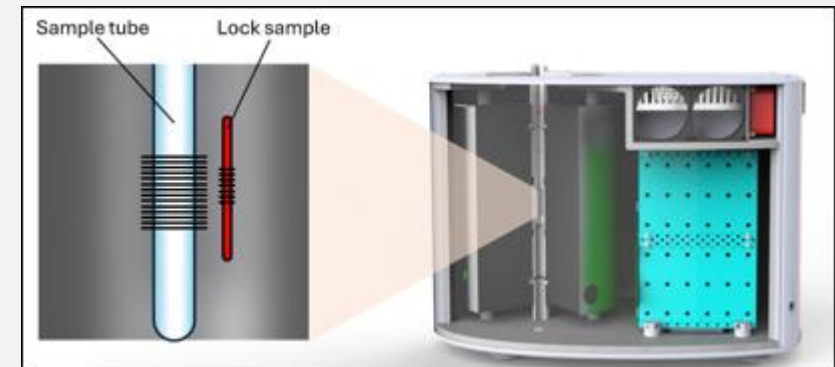
 Intuitive interface. The system cannot be damaged by wrong parameter selection.

 Highly automatable and flow compatible

 3d PFG-gradients for 2D & advanced pulse sequences




Spinsolve integrated in complex/constrained environment




Spinsolve External lock technology

External AND Hardware lock

-  - **Deuterium-free** samples
- System always ready to measure
- Automatic spectrum reference

No setting time

-  - Instant & automatic tuning/matching/locking
- 30s shimming procedure once or twice a day
- Can measure **instantly** after sample insertion

 Ever linear and constant response factor for fast and accurate **quantification**

Bringing NMR to the table



GC-MS



Raman spectroscopy



IR spectroscopy



LC-MS



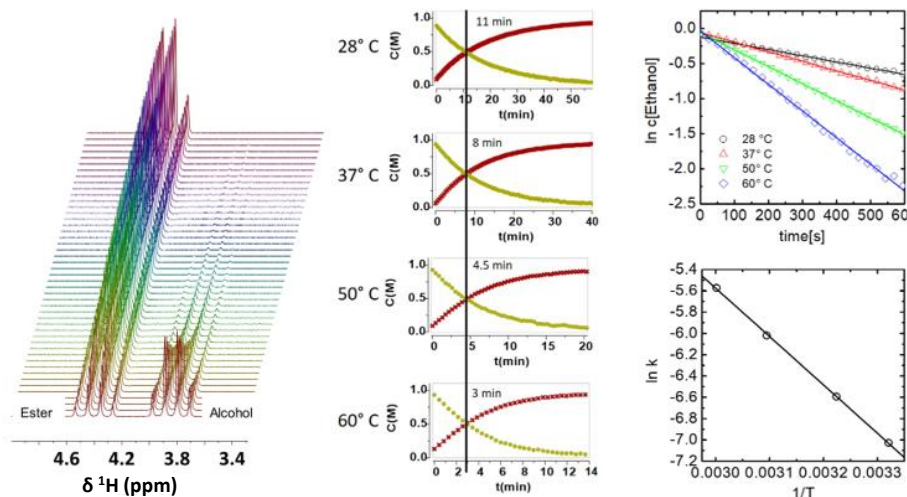
NMR spectroscopy

- Negligible-cost consumable
- No maintenance/long lifespan (> 10y)
- Low solvent volume (< 0.5 mL)
- Non-destructive
- Universal liquid detector (No-D)
- Immediate/real-time analysis
- Flow compatible
- Quantitative

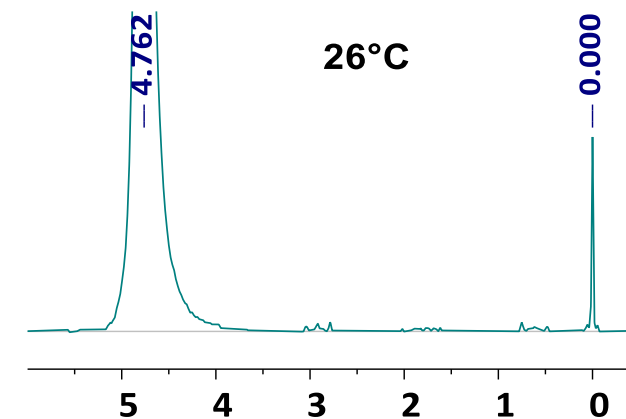
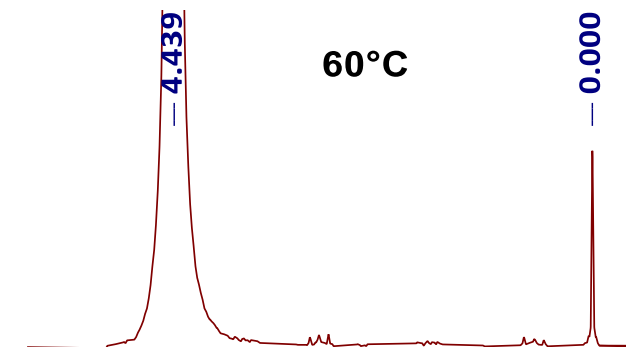


Spinsolve variable temperature option

- No external accessory or gas supply needed
- Change working temperature through software selection
- High stability, can stay weeks/months at higher temperature
- Electrical heating provides high temperature control without temperature gradient
- Compatible with all other options (Multi-X, Ultra...) , without performance loss



Esterification kinetics at different temperature

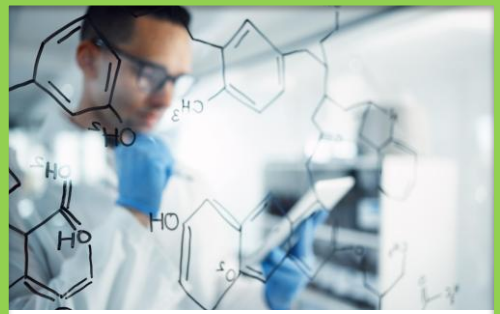


"NMR thermometer"
DSS in H₂O



Ideal with fully regulated temperature line

Benchtop NMR applications



Synthesis



Pharma



Macromolecules



Education



Flow chemistry



Batteries



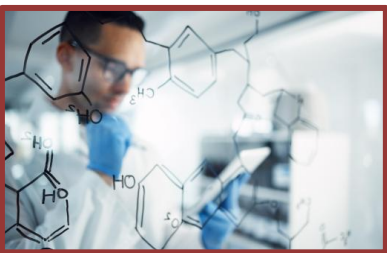
Food & Agriculture



Forensics



Analytics



High quality 1D data for structure confirmation

Gibberellic acid

Solvent: MeOH-d4

Concentration: 250 mM (20 mg)

Spinsolve 80

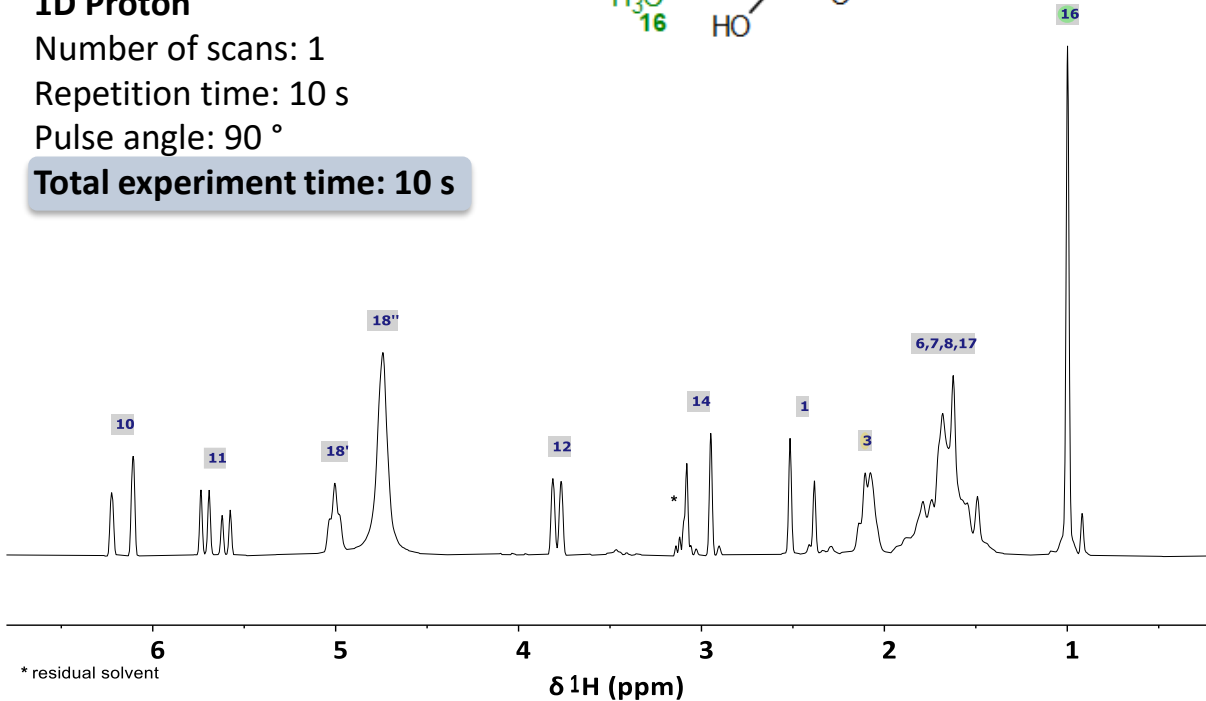
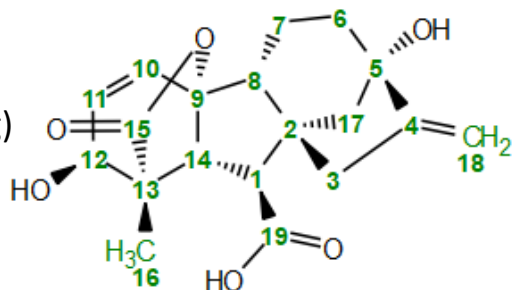
1D Proton

Number of scans: 1

Repetition time: 10 s

Pulse angle: 90 °

Total experiment time: 10 s



Gibberellic acid

Solvent: MeOH-d4

Concentration: 250 mM (20 mg)

Spinsolve 80

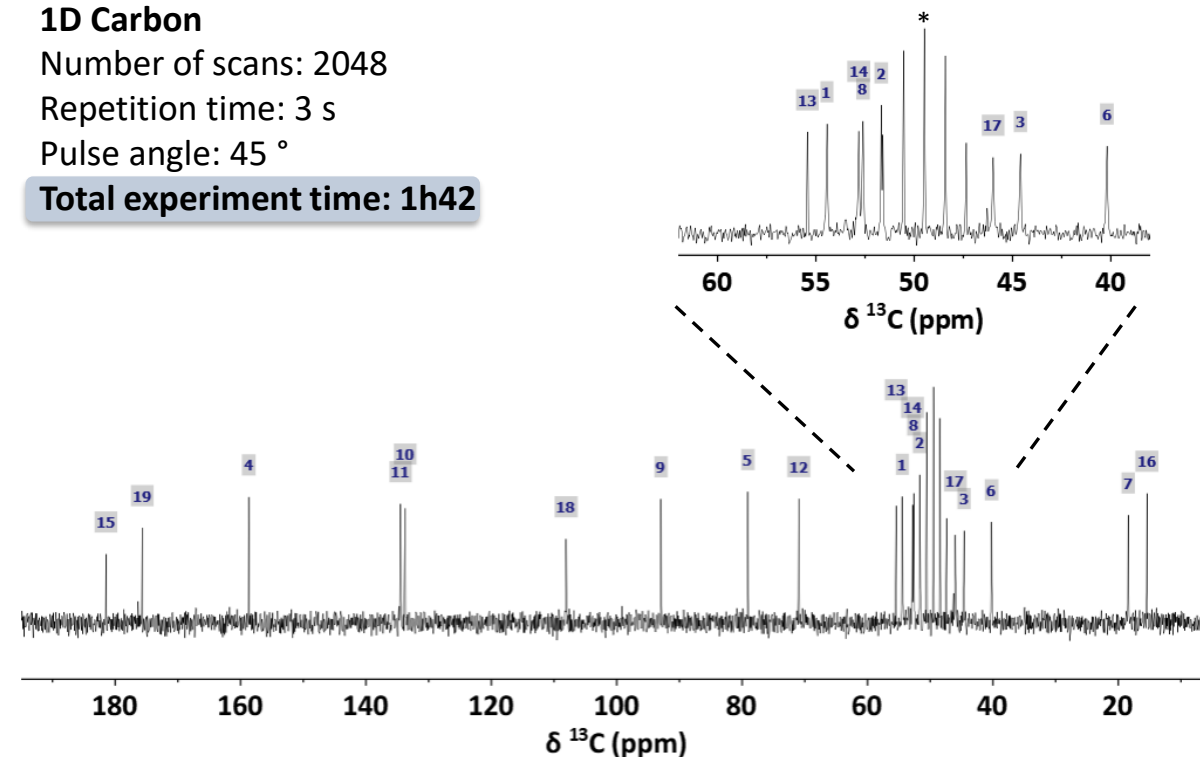
1D Carbon

Number of scans: 2048

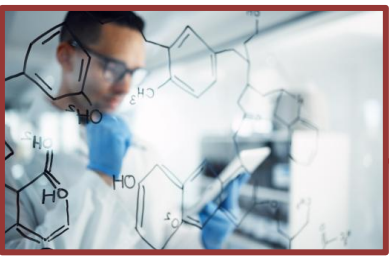
Repetition time: 3 s

Pulse angle: 45 °

Total experiment time: 1h42



Free resources available online: <https://magritek.com/resources/spectra-library/>



High quality 2D data for structure confirmation

● **Sample: Gibberellic acid** / Solvent: MeOH-d₄ / Concentration: 250 mM (20 mg)



Spinsolve 80 ULTRA

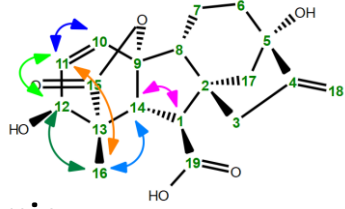
2D ¹H-¹H gsCOSY

Number of scans: 1

Repetition time: 1 s

Number of steps: 512

Total experiment time: 10 min



2D ¹H-¹³C gsHSQC-ME

Number of scans: 2

Repetition time: 1 s

Number of steps: 128

NUS rate: 50%

Total experiment time: 4 min

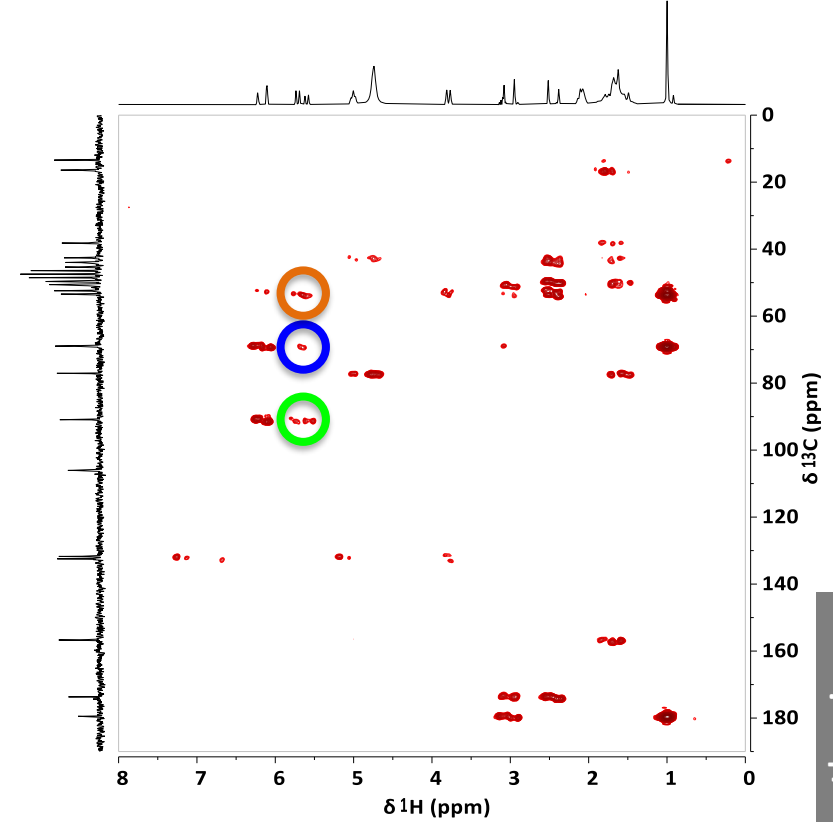
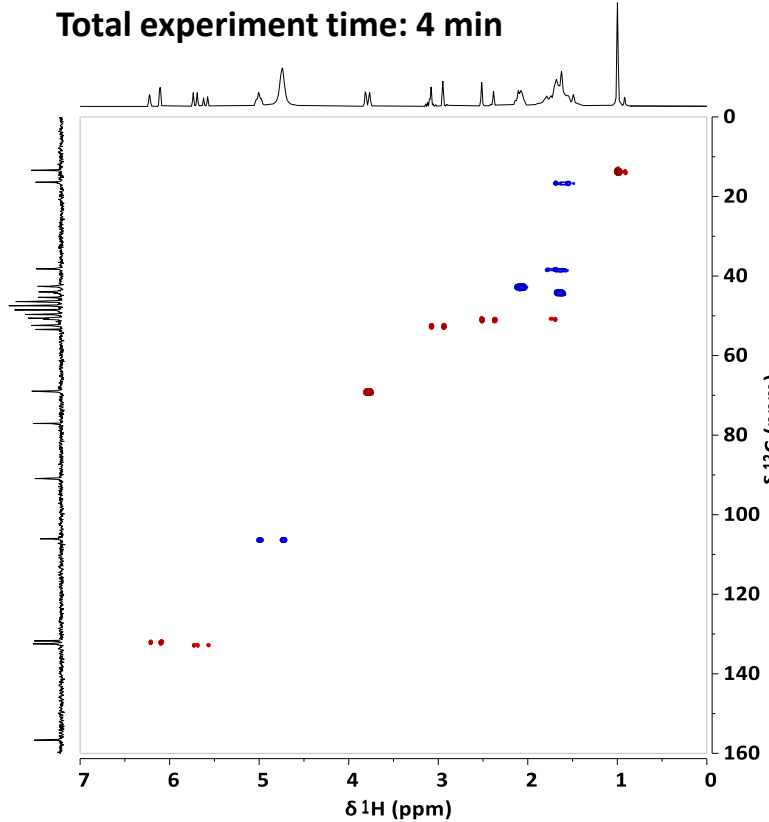
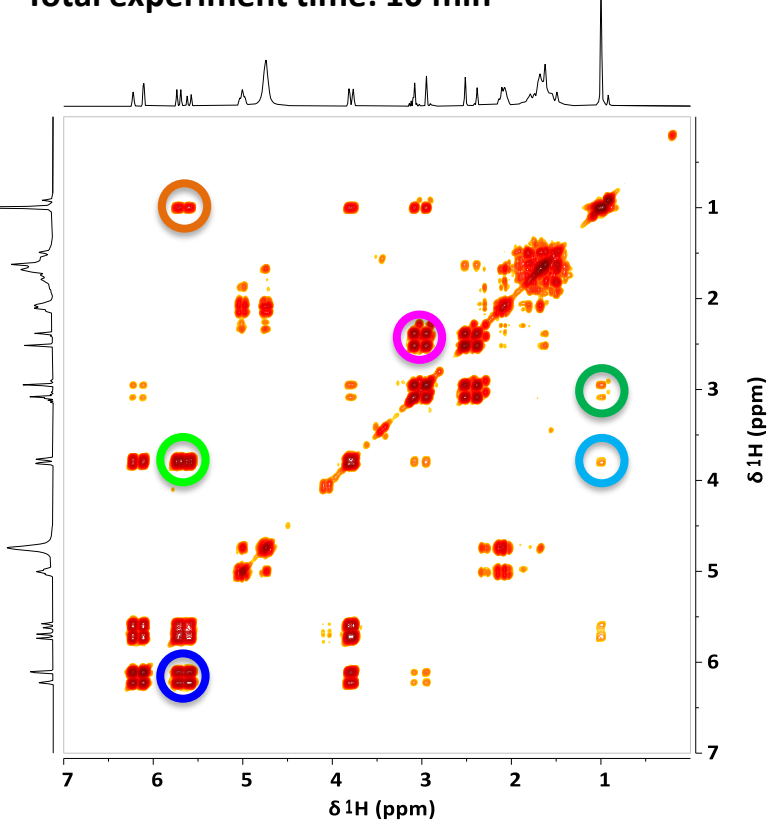
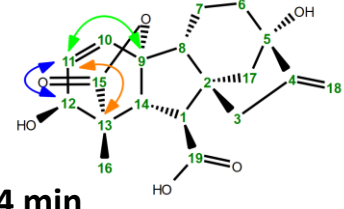
2D ¹H-¹³C gsHMBC

Number of scans: 2

Repetition time: 1 s

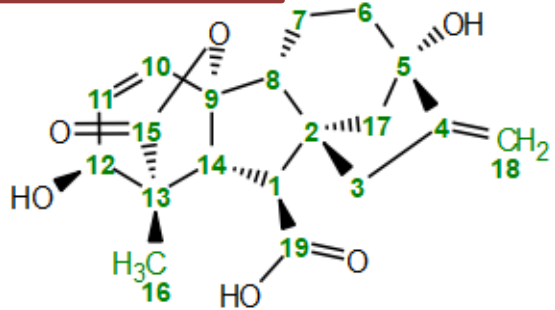
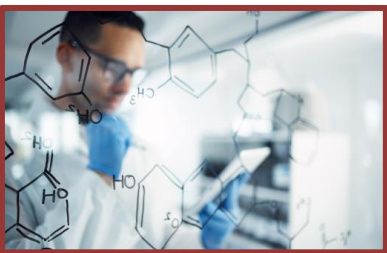
Number of steps: 256

Total experiment time: 34 min



Free resources available online: <https://magritek.com/resources/spectra-library/>

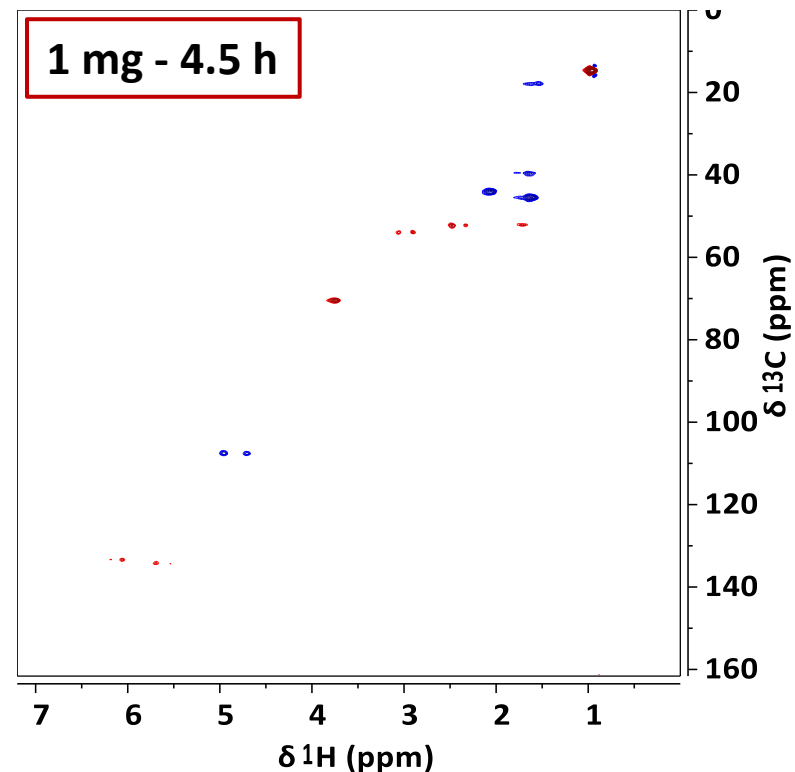
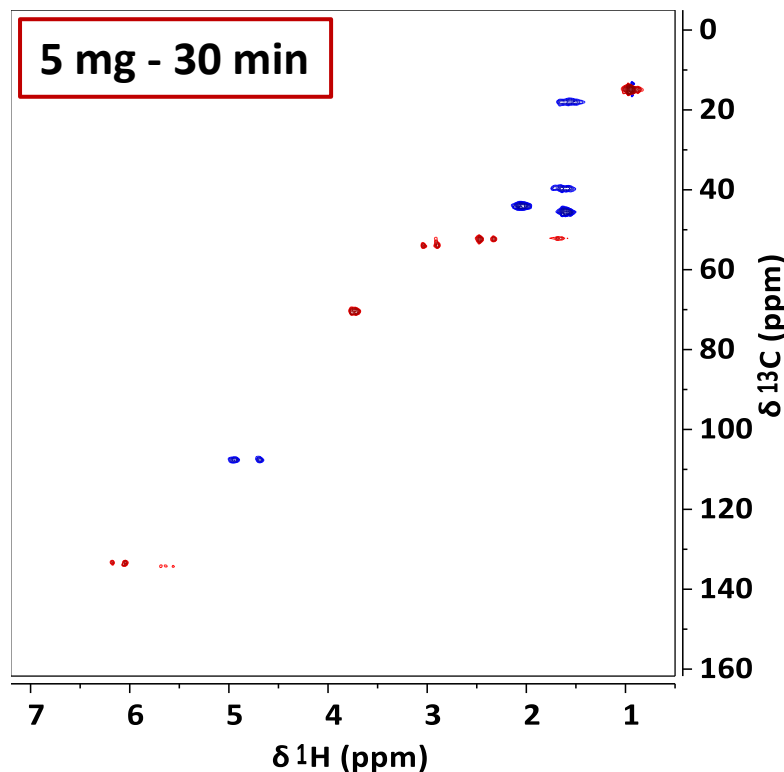
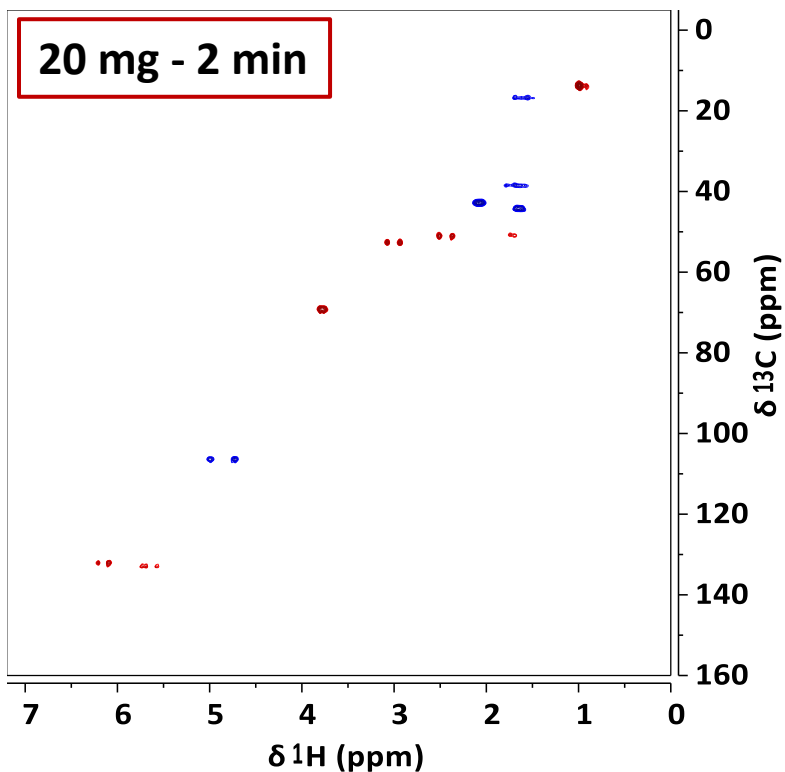
Measurement time & concentration relation



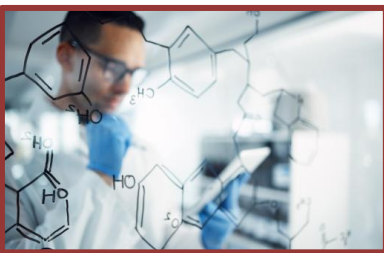
 **Spinsolve 80 ULTRA**
 **Gibberellic acid/Solvent: MeOH-d4**
 **gsHSQC-ME**

Quadratic relation between concentration and measurement time

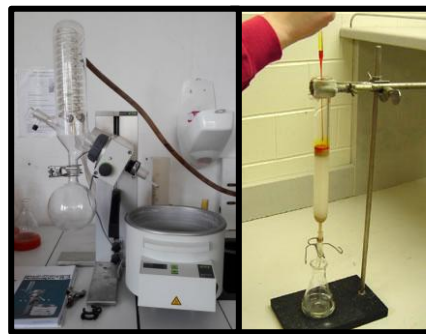
Peak separation is independent from sample concentration



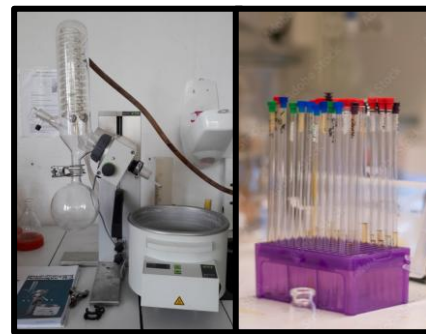
When can I get the information about my synthesis ?



Synthesis



Evaporation + separation



Evaporation + NMR tube
preparation in deuterated solvent



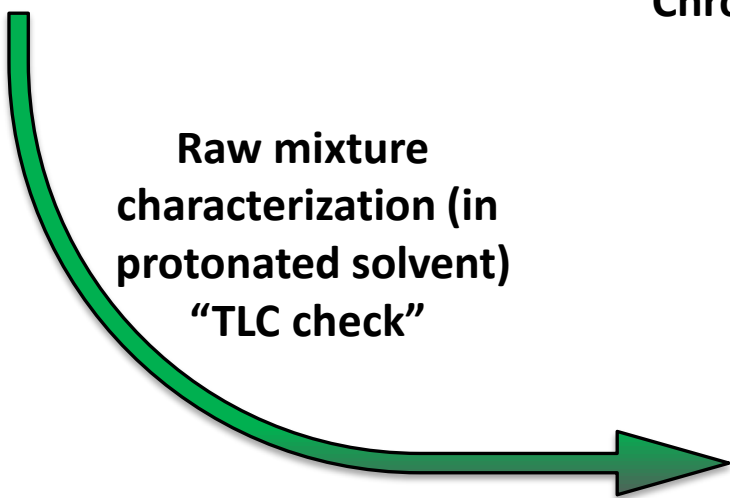
High-field NMR
analysis



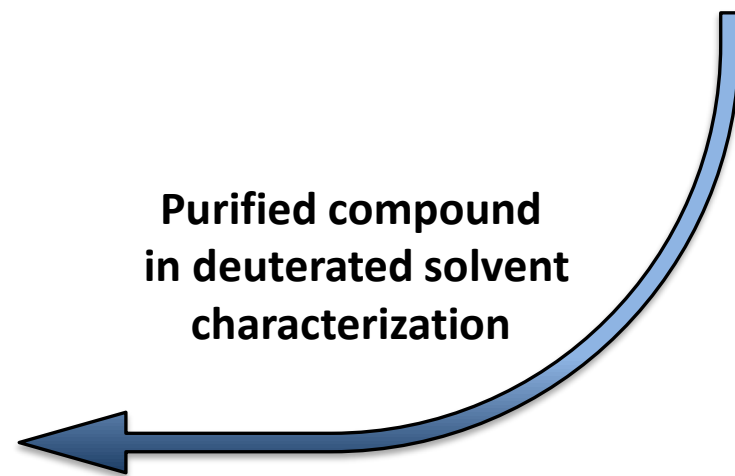
Chromatography fraction
characterization
"TLC check"



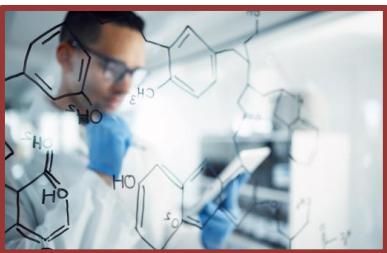
Raw mixture
characterization (in
protonated solvent)
"TLC check"



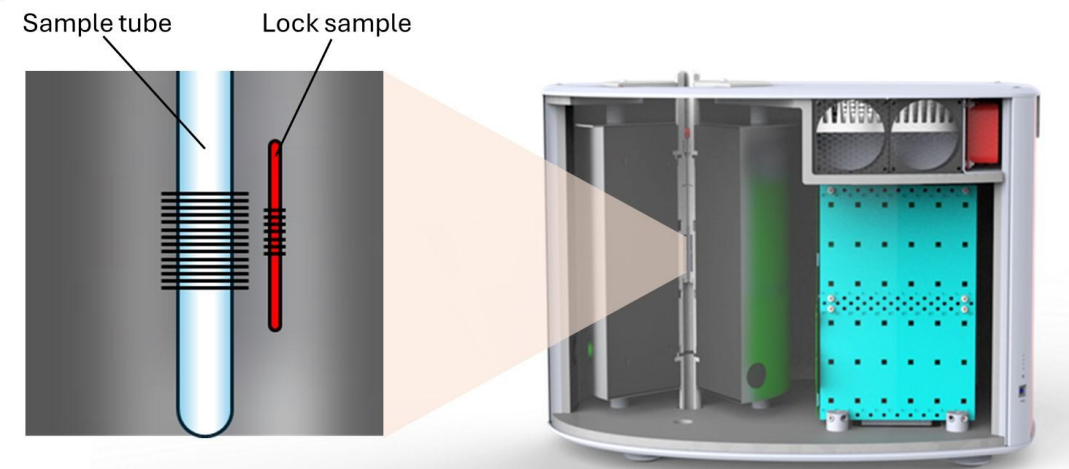
Purified compound
in deuterated solvent
characterization



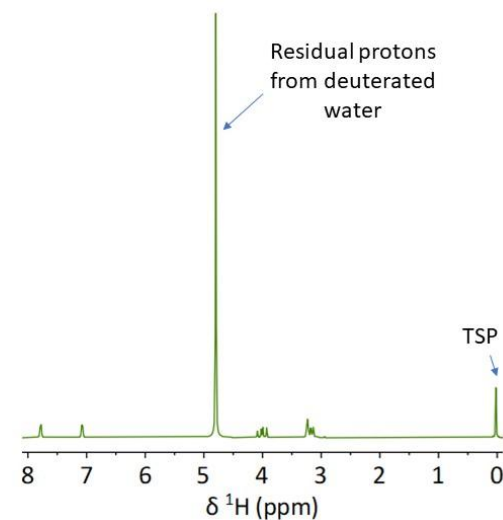
The challenge of protonated solvents – NoD NMR



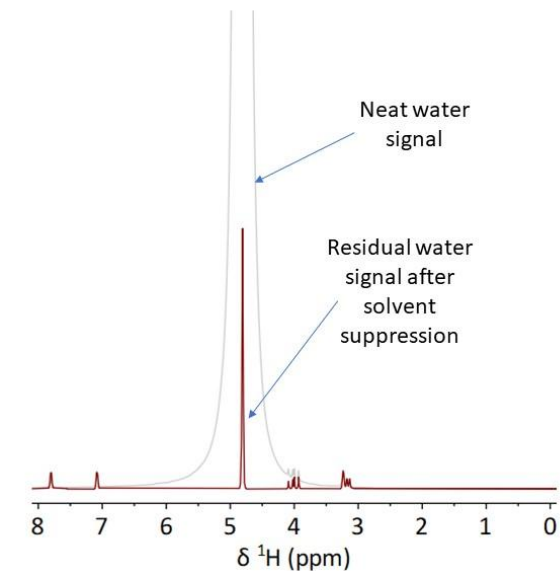
- Each Spinsolve makes use of a Hardware Lock system
- This lock runs in a fully automatic way in the background and the user does not need to take care on it.
- It is always engaged and also works when no sample is in the NMR.
- Consequently, deuterated solvents are not required for the locking of the NMR
- This means that any type of sample can be measured: neat samples, samples in protonated solvents, aqueous solutions ... and of course also samples in deuterated solvents
- Powerful solvent suppression sequences are in place (Wet, Pre-Sat – all perfectly adapted to the Spinsolve hardware)



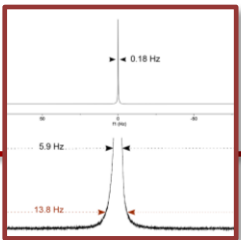
A) 8 mM Histidine in D₂O (99.9%)
1D Proton



B) 8 mM Histidine in H₂O
WET solvent suppression



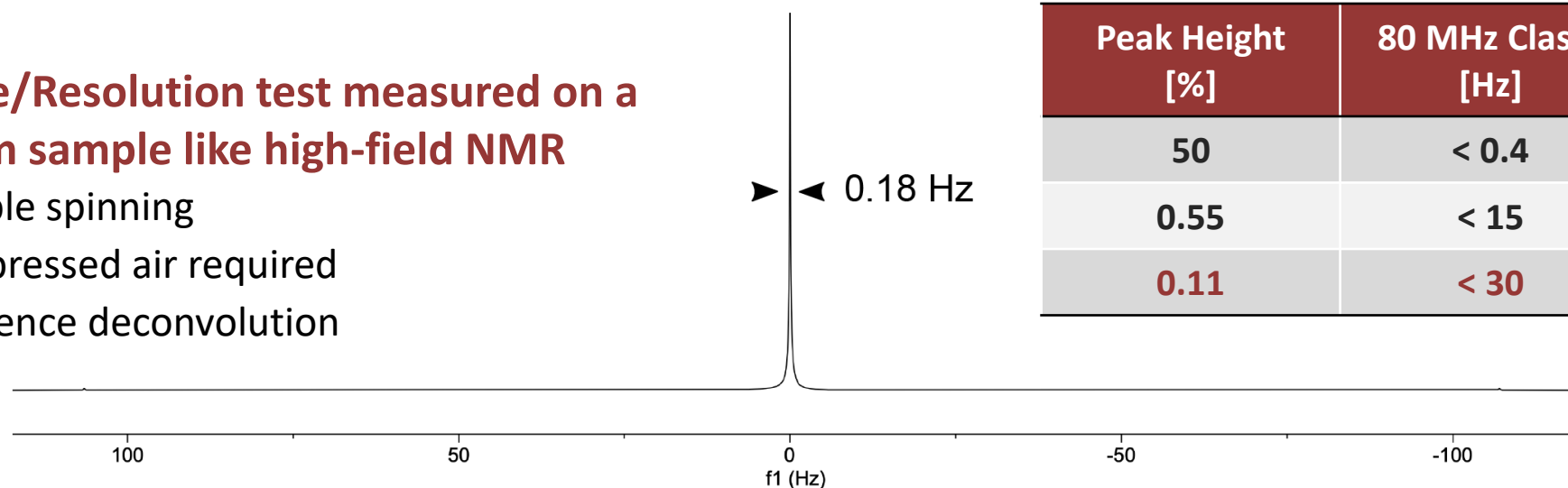
ULTRA Magnet homogeneity



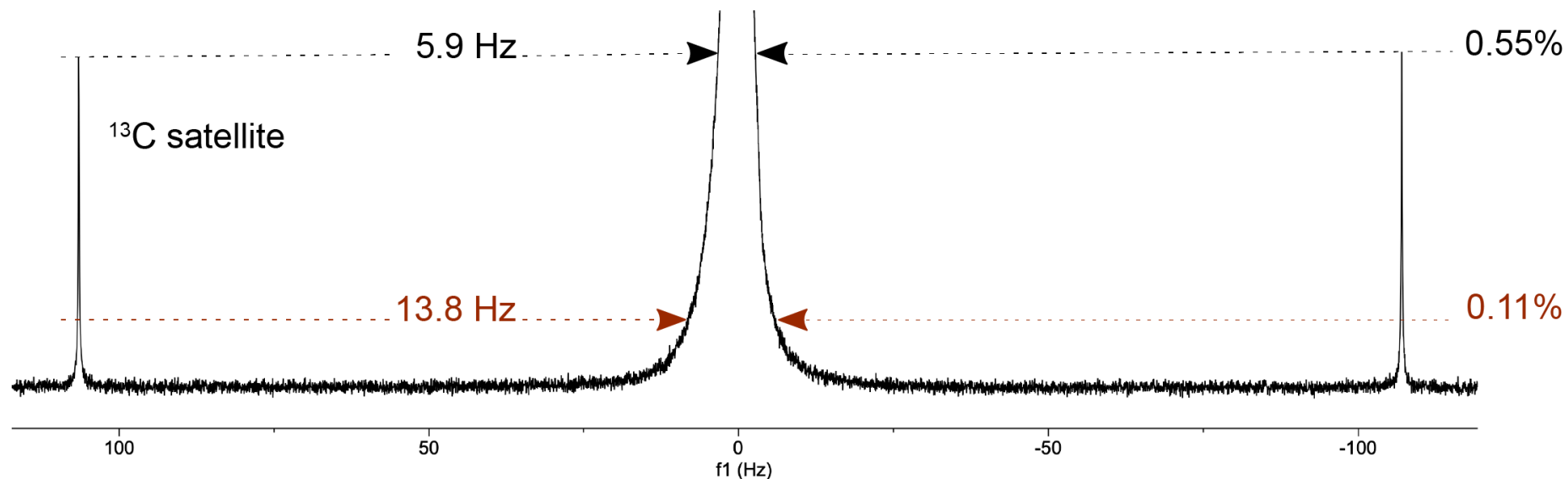
Line Shape/Resolution test measured on a chloroform sample like high-field NMR

- No sample spinning
- No compressed air required
- No reference deconvolution

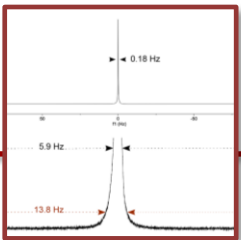
Peak Height [%]	80 MHz Classic [Hz]	80 MHz ULTRA [Hz]
50	< 0.4	< 0.2
0.55	< 15	< 8
0.11	< 30	< 16



Zoom x 200



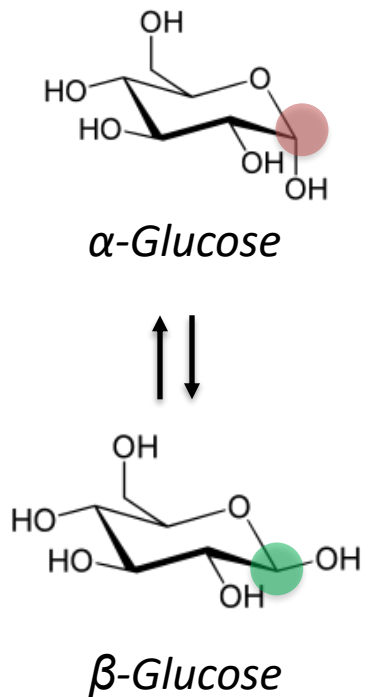
No-D NMR: Remove the interference of protonated solvents



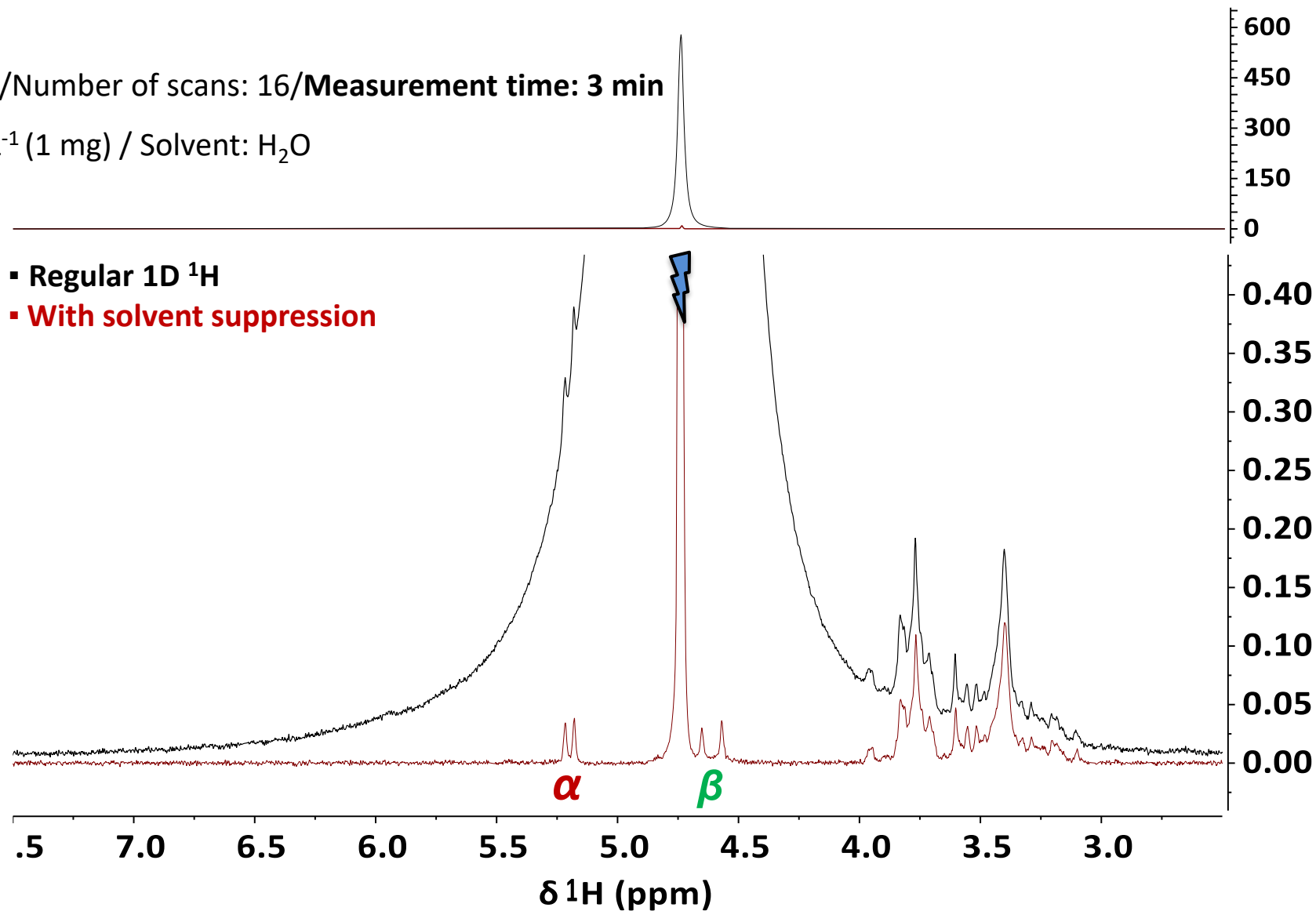
1D ¹H (WET)/Repetition time: 10 s/Number of scans: 16/Measurement time: 3 min

Glucose/Concentration: 20 mmol.L⁻¹ (1 mg) / Solvent: H₂O


Spinsolve 90 ULTRA



- Regular 1D ¹H
- **With solvent suppression**

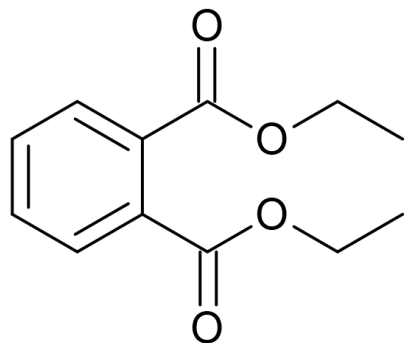


No-D NMR: Remove the interference of protonated solvents

 **1D ^1H (WET)**/Repetition time: 10 s/Number of scans: 8/Measurement time: 2 min

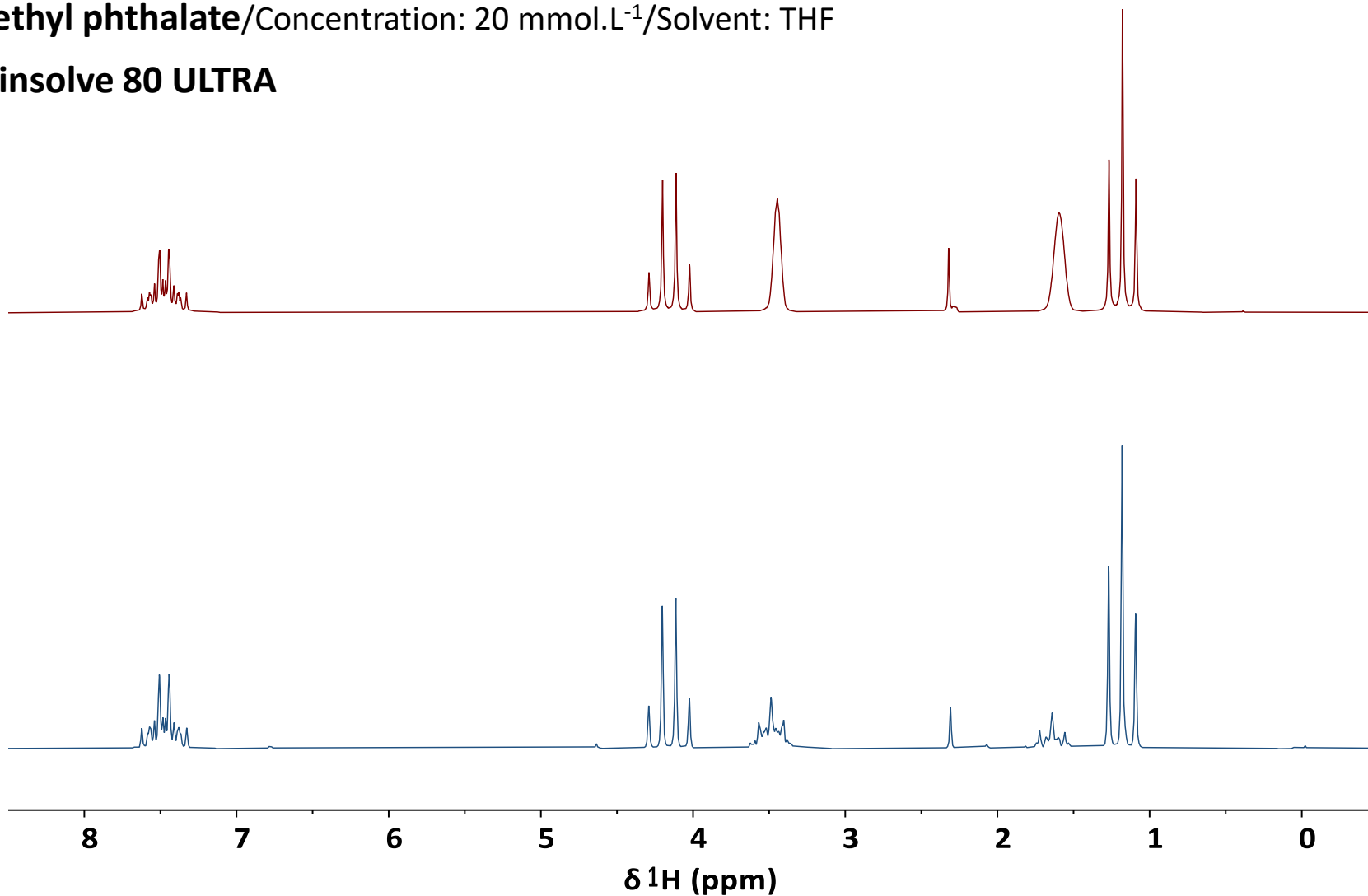
 **Diethyl phthalate**/Concentration: 20 mmol.L⁻¹/Solvent: THF

 **Spinsolve 80 ULTRA**




Diethyl phthalate

2 mg
in
THF-D8
or
THF-H8
?



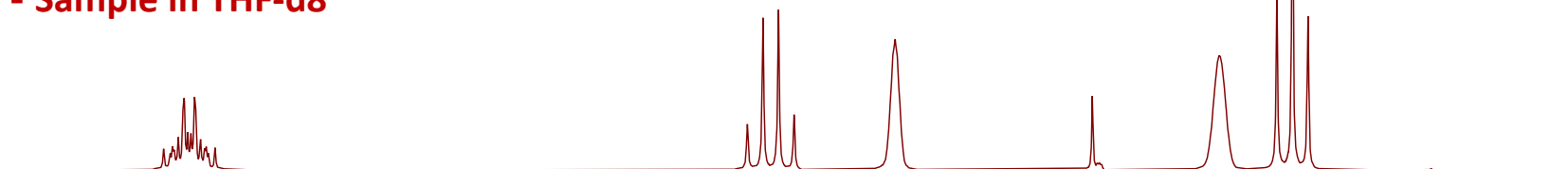
No-D NMR: Remove the interference of protonated solvents

 **1D ¹H (WET)**/Repetition time: 10 s/Number of scans: 8/Measurement time: 2 min

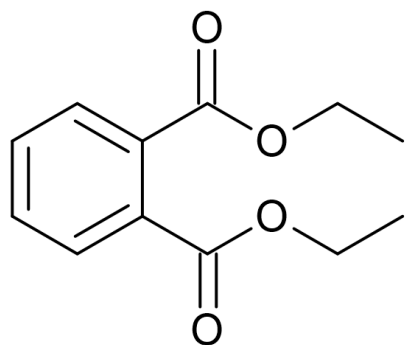
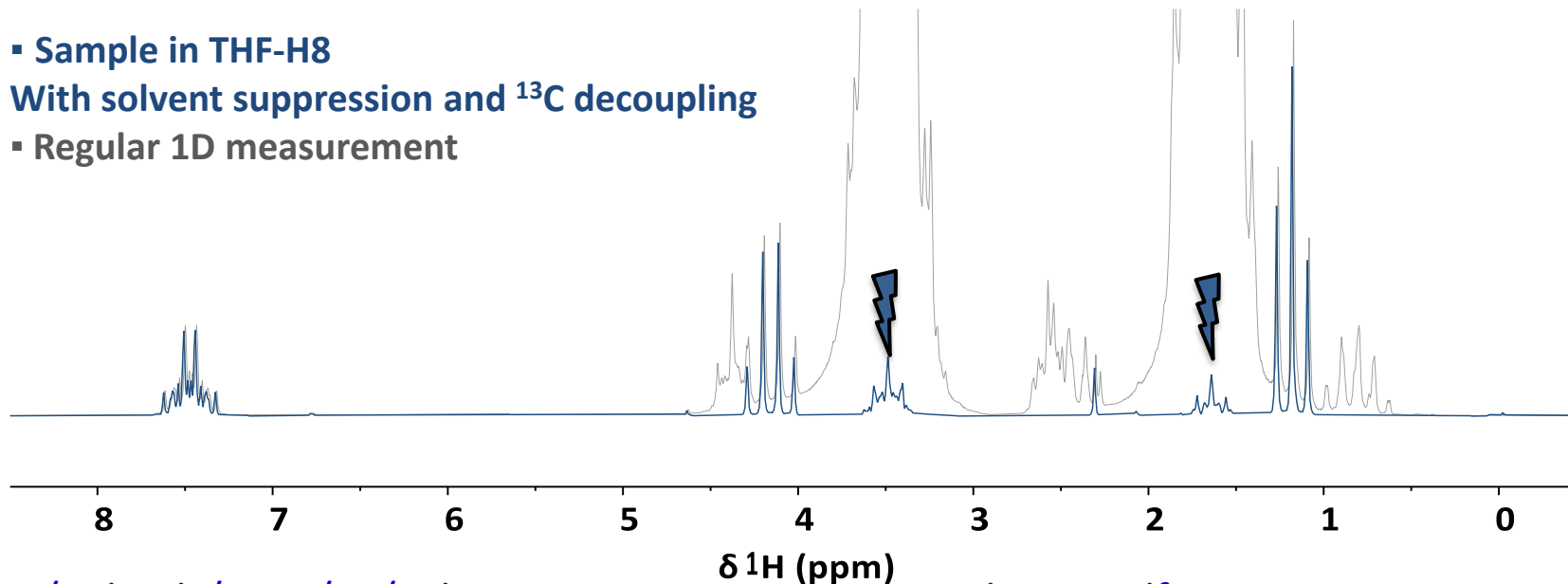
 **Diethyl phthalate**/Concentration: 20 mmol.L⁻¹/Solvent: THF

 **Spinsolve 80 ULTRA**

▪ **Sample in THF-d8**



▪ **Sample in THF-H8**
 With solvent suppression and ¹³C decoupling
 ▪ Regular 1D measurement



Diethyl phthalate

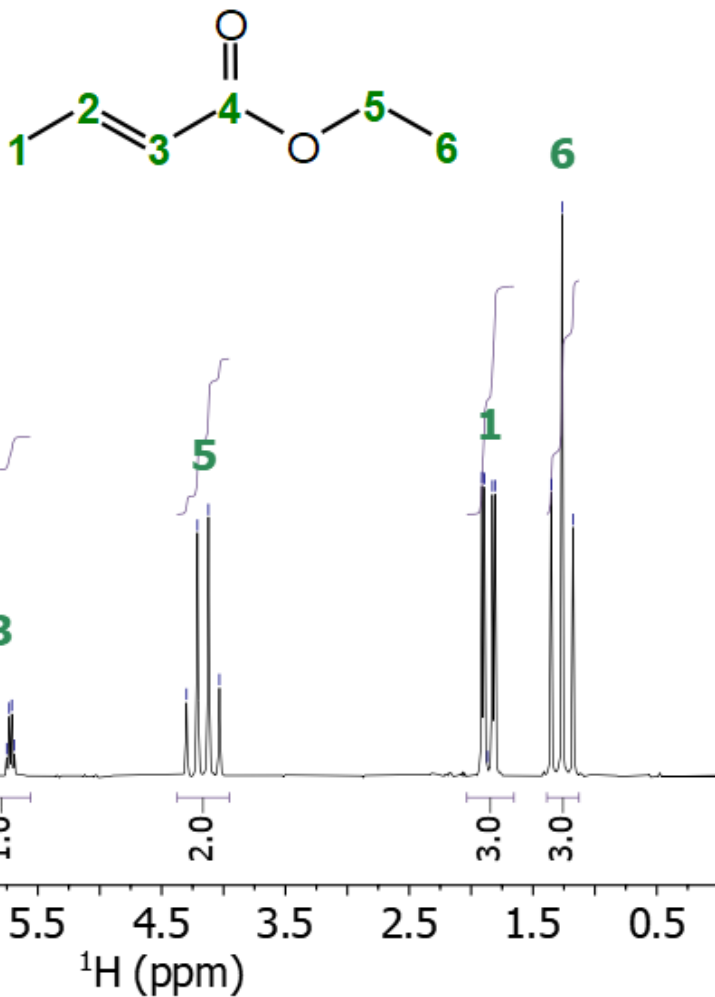
2 mg
 in
 THF-D8
 or
 THF-H8
 ?

Relative quantification with NMR



Ethyl crotonate

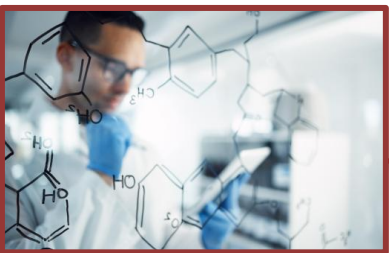
Solvent = CDCl_3
 Concentration = 250 mM
 Number of scans = 1
 Total experimental time = 10 s
 Frequency = 80 MHz



Specificity

- Each chemical group of a compound does give a signal at a distinct position
- The position and the pattern of the signals give you information about the chemical environment
- The ratio of the peaks gives information about how many hydrogen nuclei contribute to a signal
- Hence, a single signal can be used to quantify the compound

Spinsolve quantification in different tubes & different solvents



Common solvents (analytical grade, neat)



1D ^1H



Spinsolve 60

- Unlike high-field systems, Spinsolve systems work with a fixed receiver gain
- Spinsolve NMR has this unique feature that every proton in ~~the~~ same any sample (including different solvents) gives the same "amount" of signal, if relaxation is fulfilled

Acetonitrile

THF

Dichloromethane

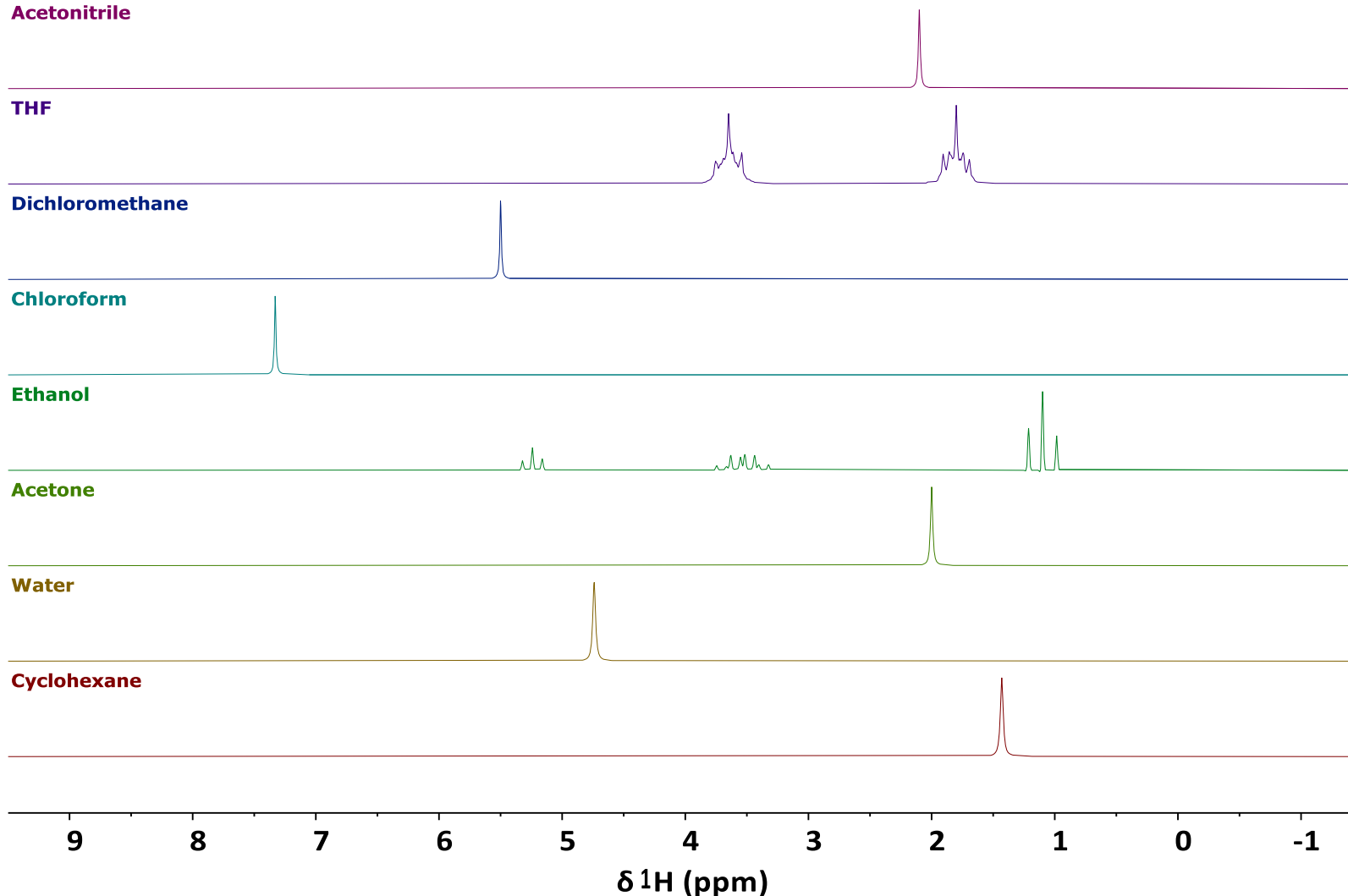
Chloroform

Ethanol


Acetone


Water

Cyclohexane



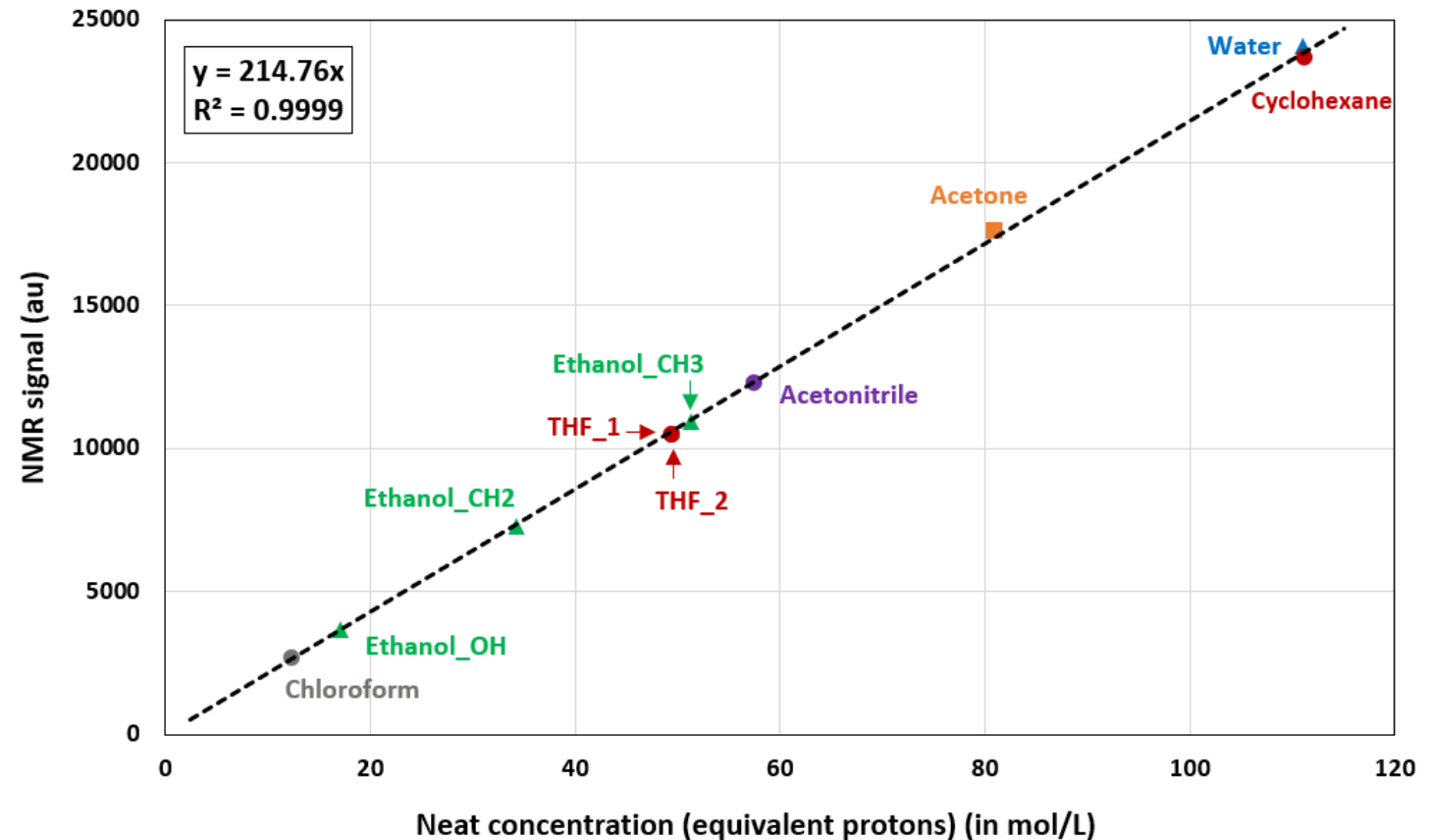
Spinsolve quantification in different samples

 Common solvents (analytical grade, neat)

 1D ^1H (Single scan)

 Spinsolve 60

Solvent	Density (g/L)	M (g/mol)	C _{neat} (mol/L)	Heq	C _{perH} (mol/L)	NMR signal (au)
Water	1000	18.01	55.50	2	111.0	24035
Chloroform	1480	119.3	12.39	1	12.4	2682
Ethanol_CH3	789	46.0	17.12	3	51.4	10917
Ethanol_CH2	789	46.0	17.12	2	34.3	7269
Ethanol_OH	789	46.0	17.12	1	17.1	3637
Acetone	784	58.0	13.49	6	81.0	17589
Acetonitrile	786	41.0	19.14	3	57.4	12299
THF_1	890	72.1	12.34	4	49.4	10524
THF_2	890	72.1	12.34	4	49.4	10517
Cyclohexane	779	84.15	9.25	12	111.1	23696

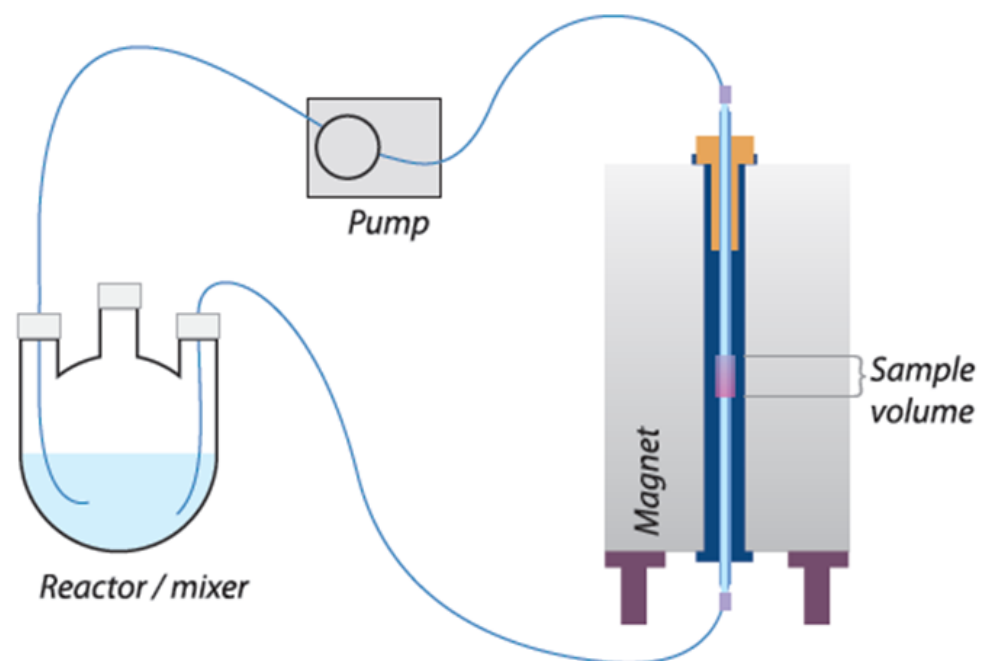


➤ Any peak that can be integrated can be quantified, allowing to convert integrals into yields in raw mixtures



Reaction monitoring kit

Following the progress of a reaction from starting material to end-point

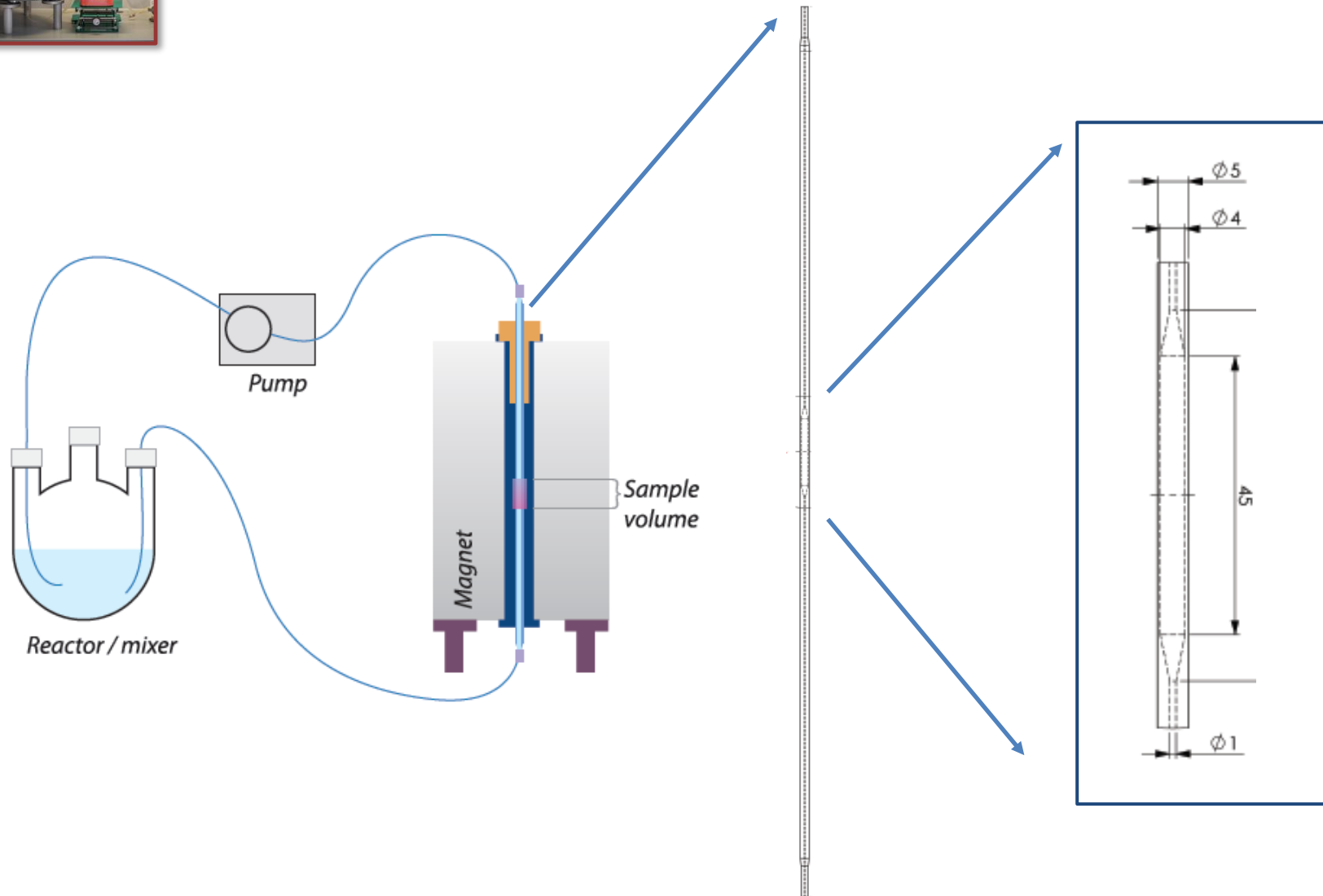


Flow reaction monitoring setup under a fumehood

- Every Spinsolve system has an opening all the way from top to bottom
- So a flow cell can be positioned into the NMR spectrometer
- Take an NMR spectrum all 30 ... 60 seconds
- Run the setup either in continuous or stop-flow mode
- No Need for Deuterated Solvents / External Lock in Operation



Optimized glass flow-cell for maximum SNR



The Spinsolve Glass Flow cell is volume-optimized to achieve the best possible SNR with the minimum dead time / volume.

- Maximum diameter in the center of the cell (rf coil region) to use full sample volume and match the SNR specified for 5 mm NMR tubes
- Total volume of the cell kept to the minimum to reduce the dead volume of the setup and minimize the pumping time from the reactor to the system.

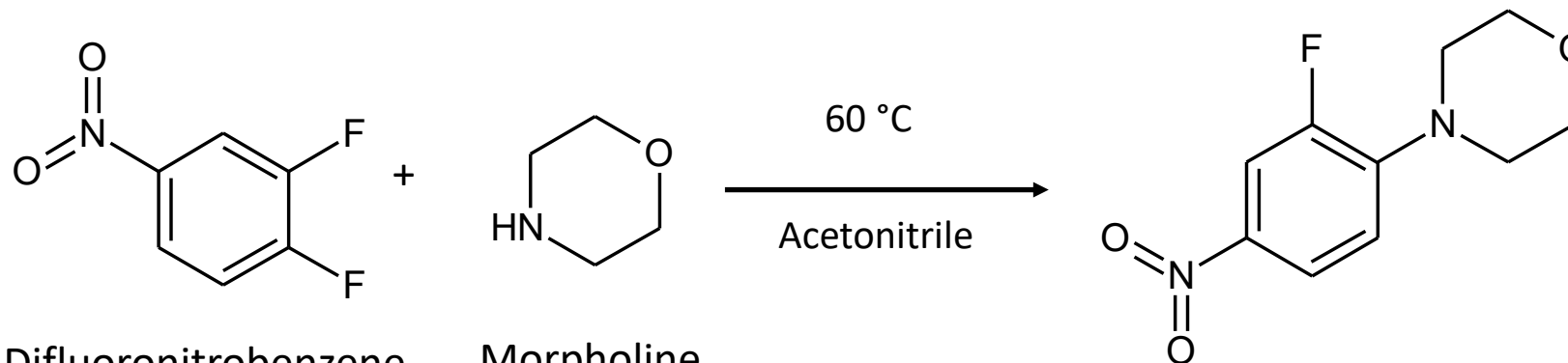


RMX software module for on-line monitoring





Reaction monitoring - S_NAr reaction



3,4-Difluoronitrobenzene
(Dropwise addition)

Morpholine

- Continuous flow with ~0.8 mL/min
- Dropwise addition of 3,4-FNB to solution of Morpholine in ACN
- Heating to 60 °C in waterbath after addition was complete
- Reaction was monitored for ~2 h, after that mixture was stirred over night
- Precipitation of white solid

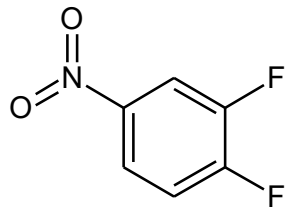
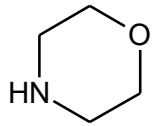
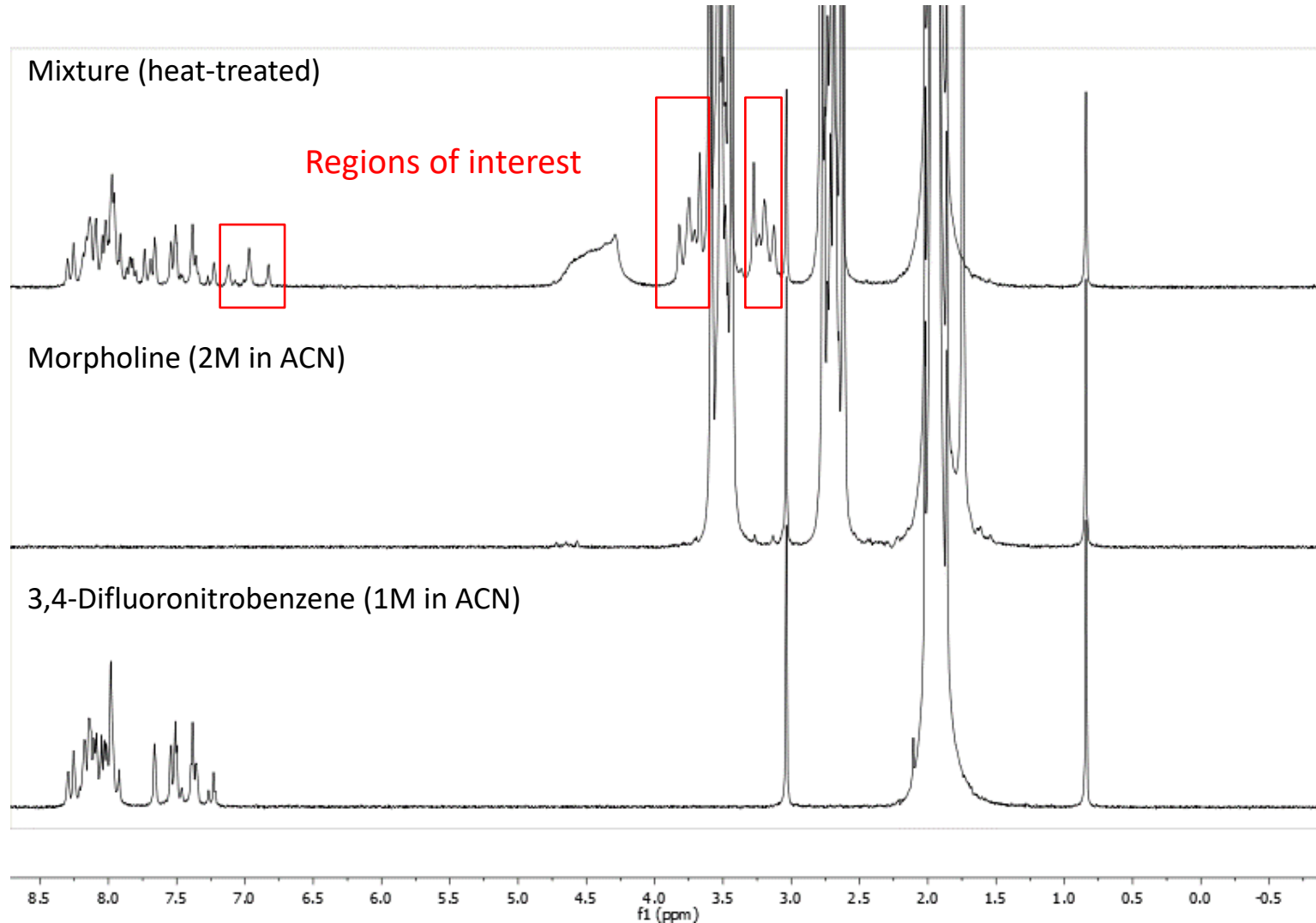




Preliminary experiments in 5 mm NMR tube

1D ¹H/Repetition time: 10 s/Number of scans: 2/Measurement time: 20 s

Spinsolve 60



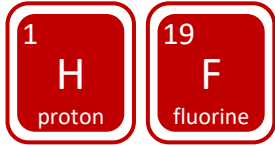


Reaction Monitoring – Stacked plot 1H and 19F NMR

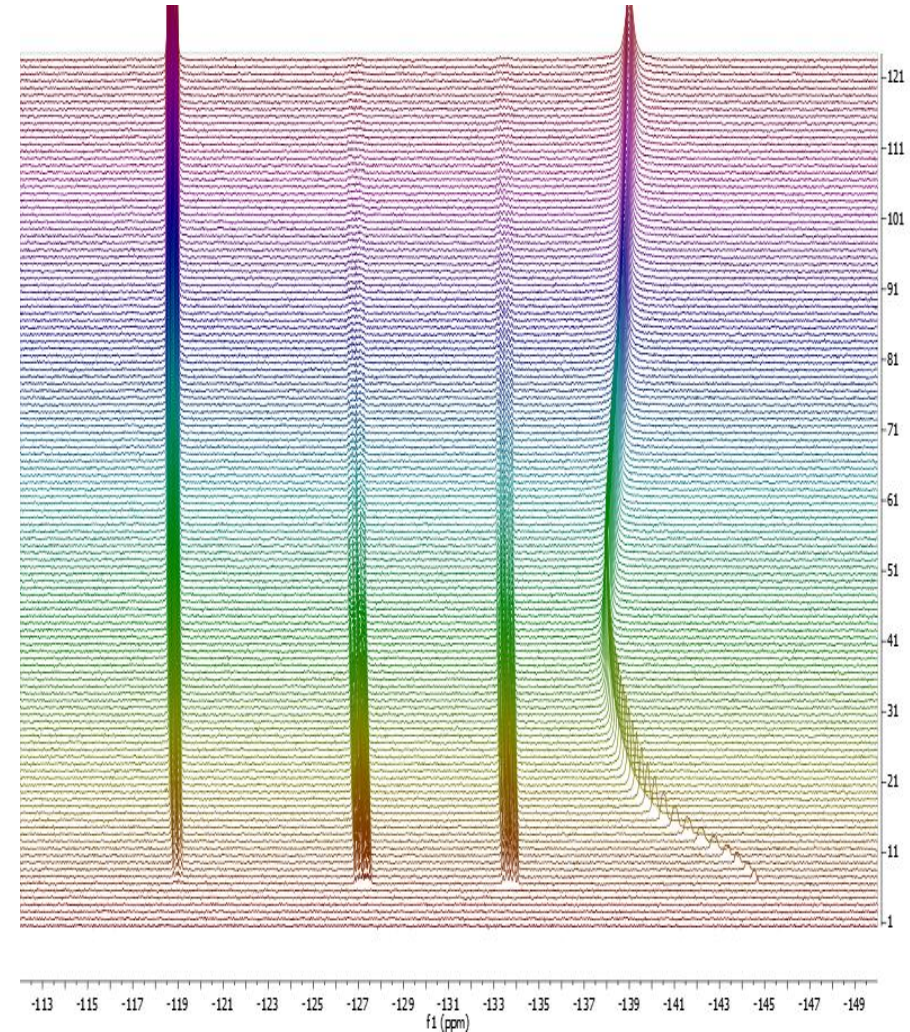
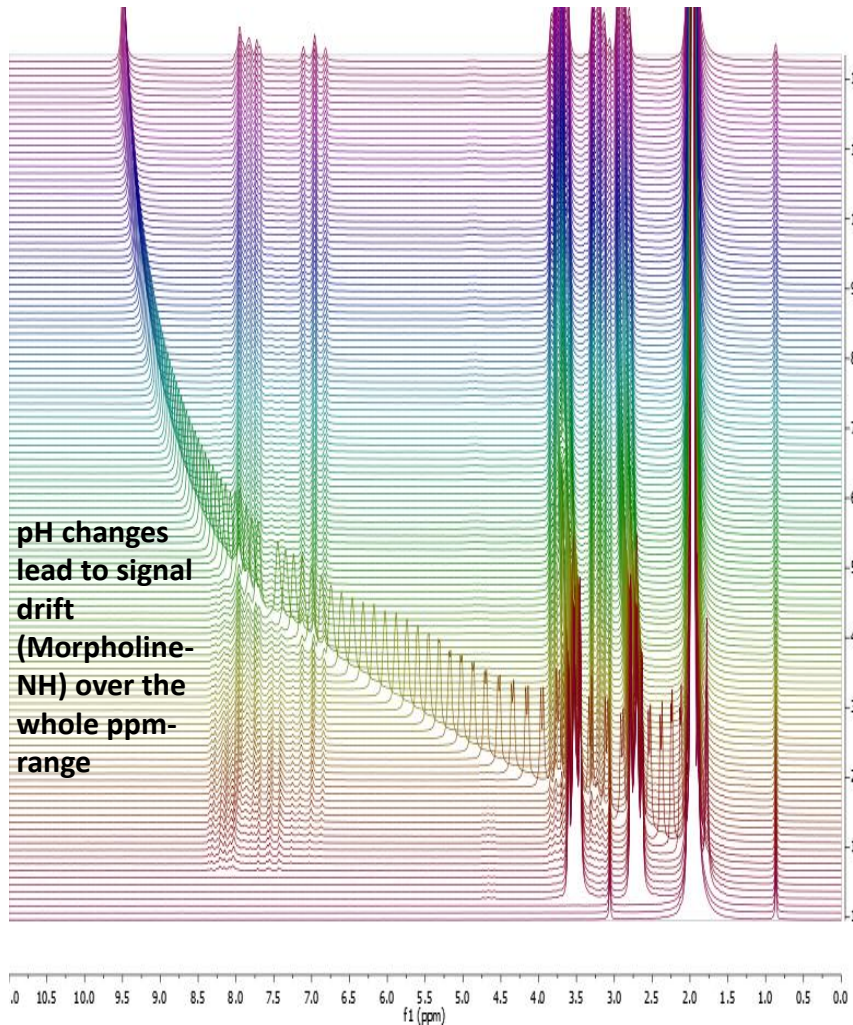
124 spectra were acquired over the time of the study

1D ¹H/Repetition time: 10 s/Number of scans: 2/Measurement time: 20 s

1D ¹⁹F/Repetition time: 4 s/Number of scans: 8/Measurement time: 32 s



Spinsolve 60





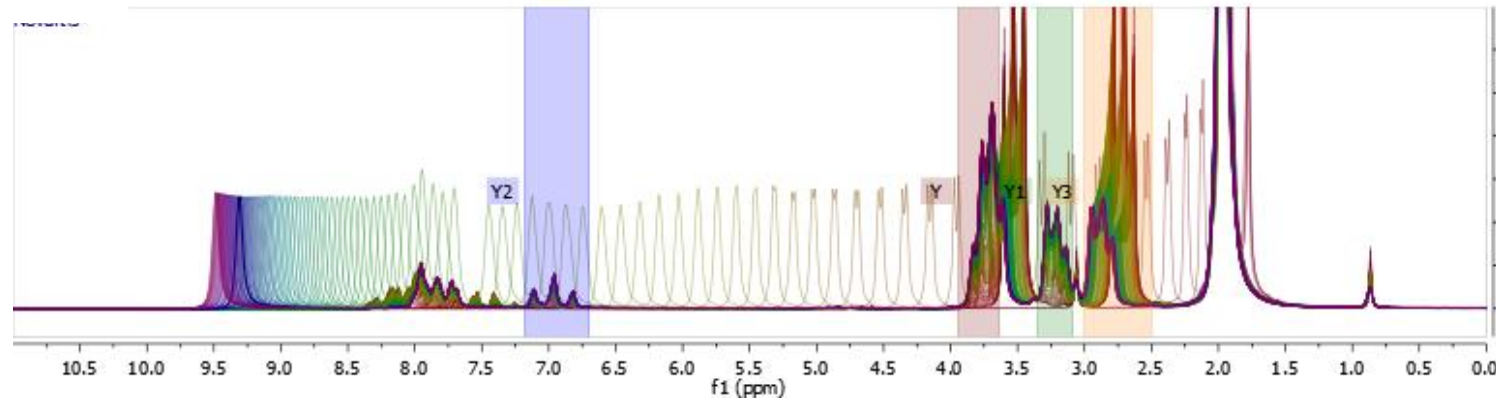
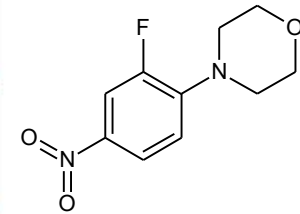
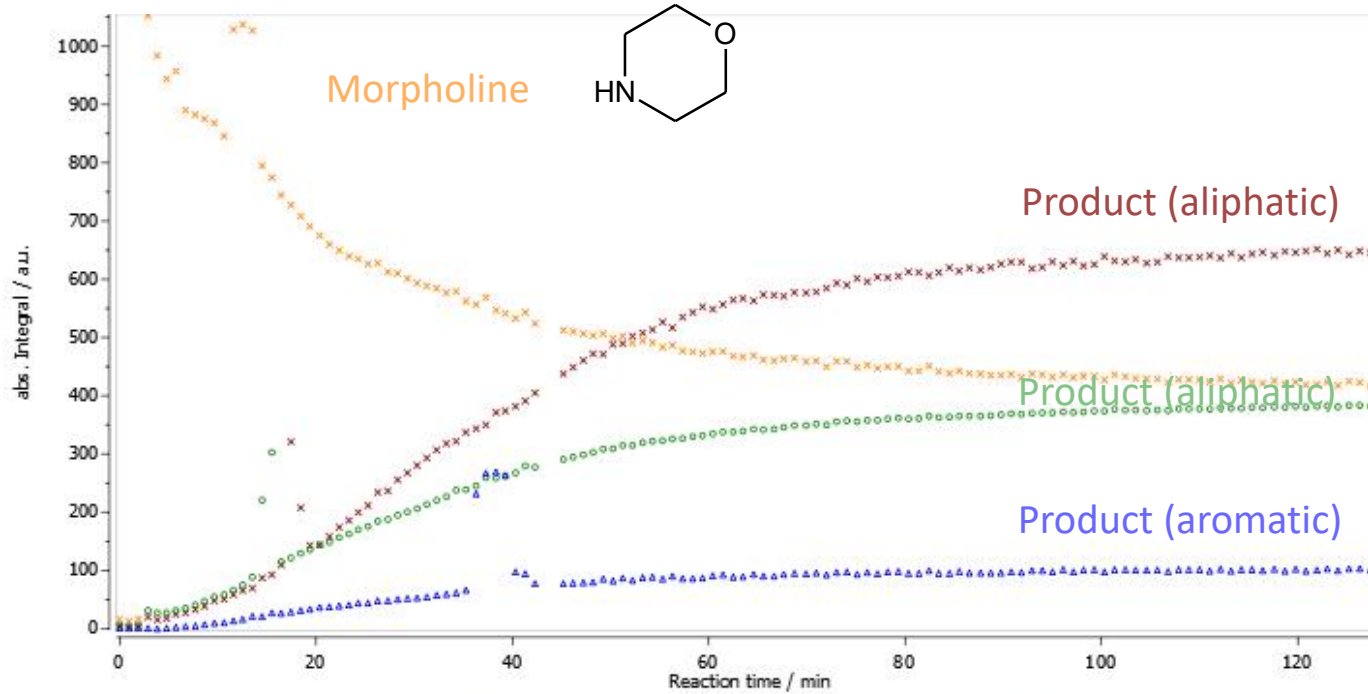
Reaction Monitoring – Regions of Interest 1H NMR

1D ¹H/Repetition time: 10 s/Number of scans: 2/Measurement time: 20 s

Spinsolve 60



Spinsolve 60



Reaction Monitoring – Regions of Interest 19F NMR

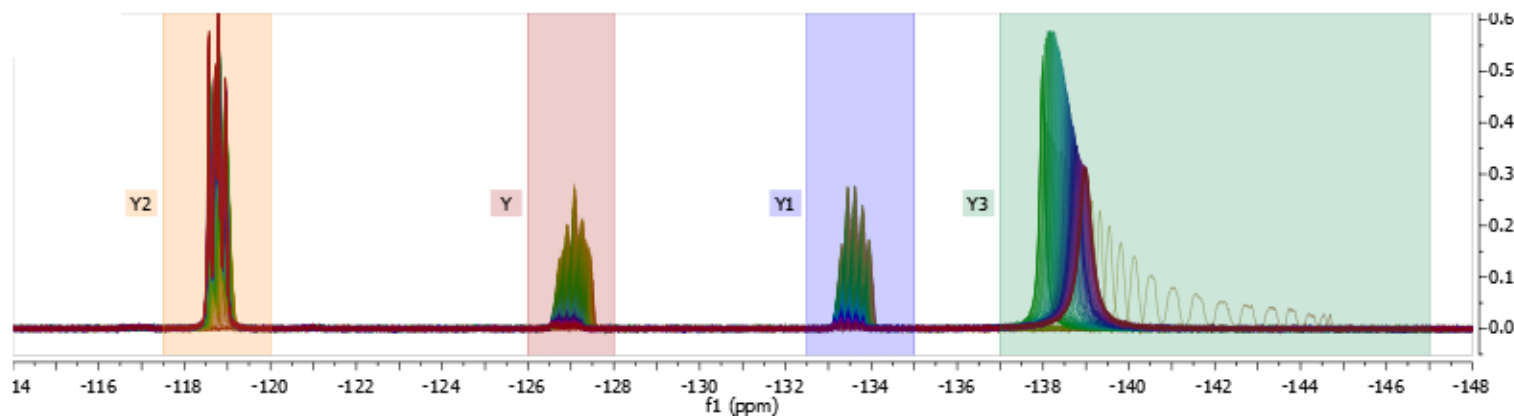
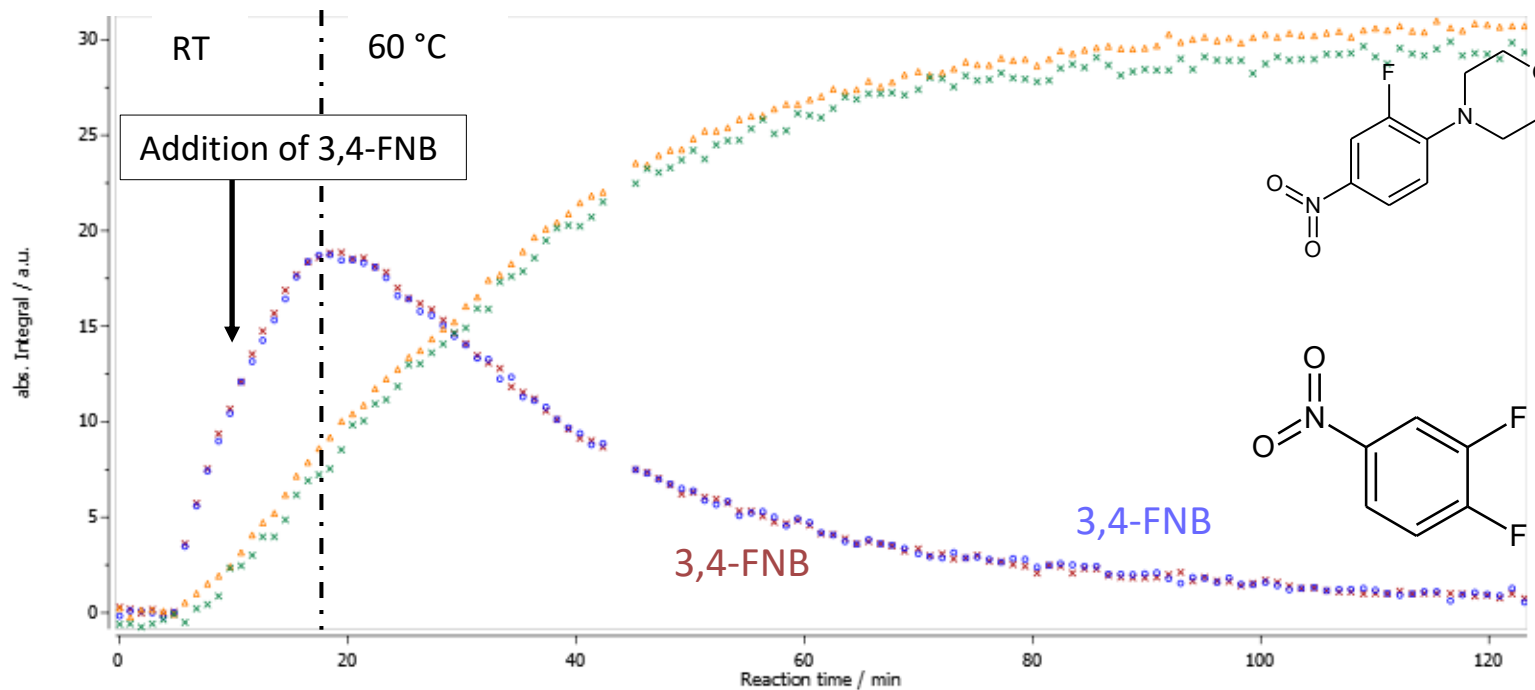


1D ¹⁹F/Repetition time: 4 s/Number of scans: 8/Measurement time: 32 s

Spinsolve 60

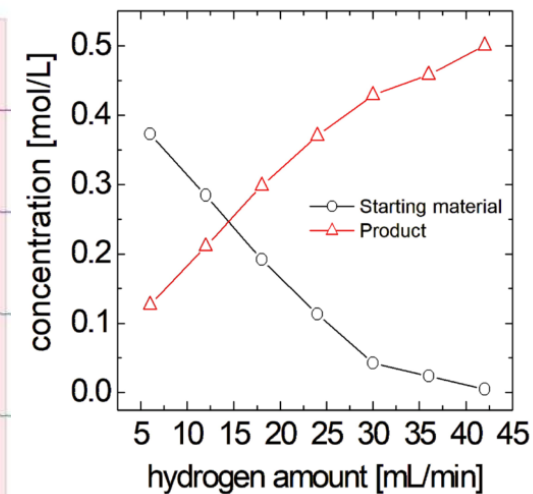
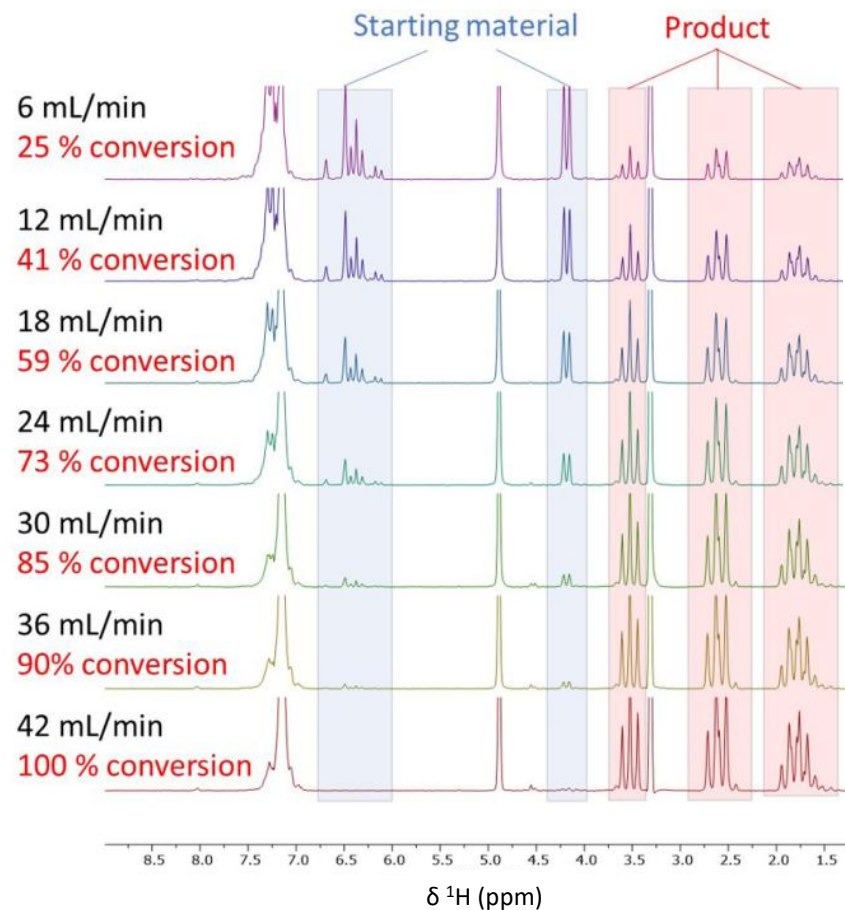
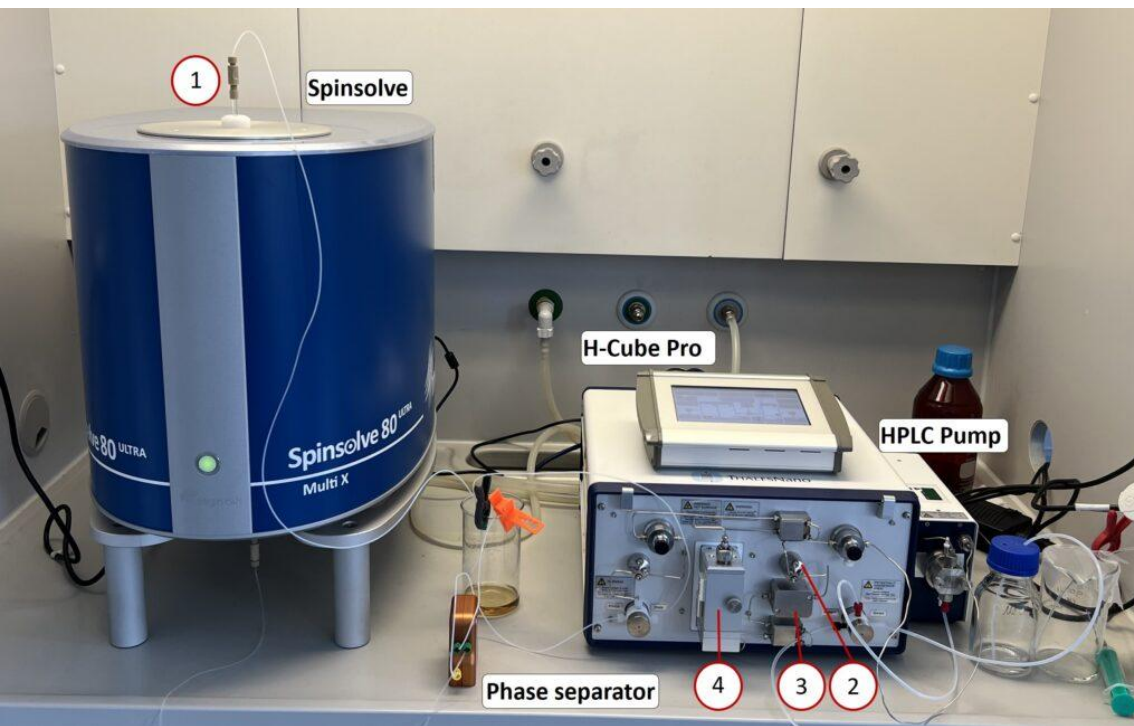
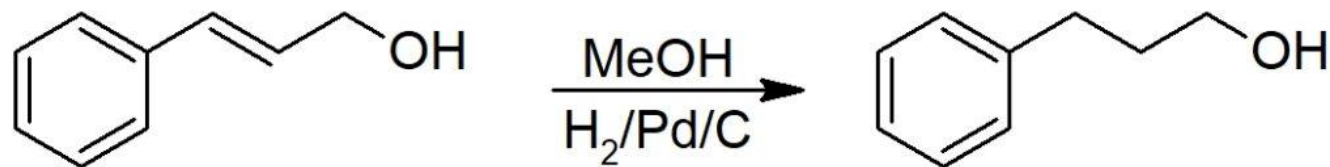
¹⁹F
fluorine

Spinsolve 60





Spinsolve hyphenation with an H-Cube Pro[®] from ThalesNano[®]

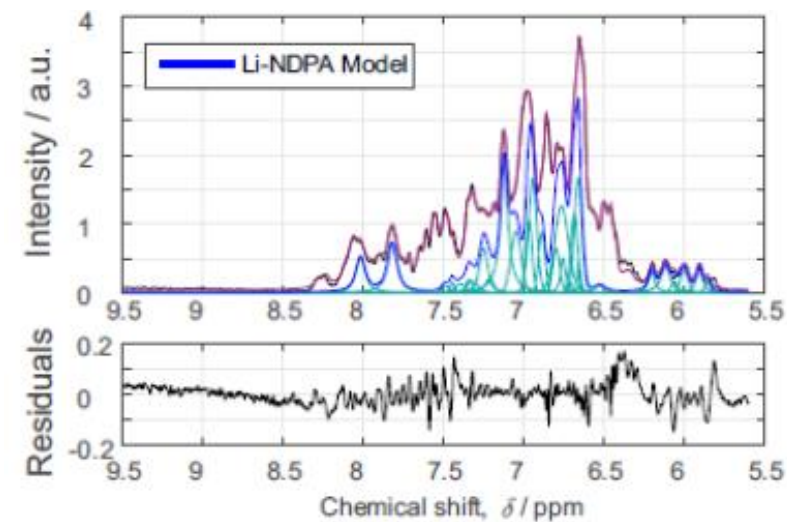
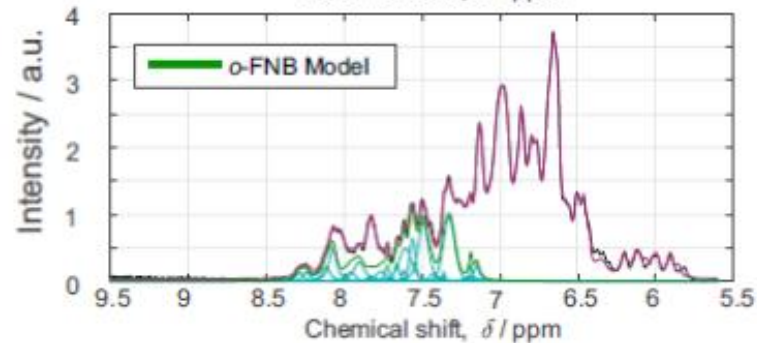
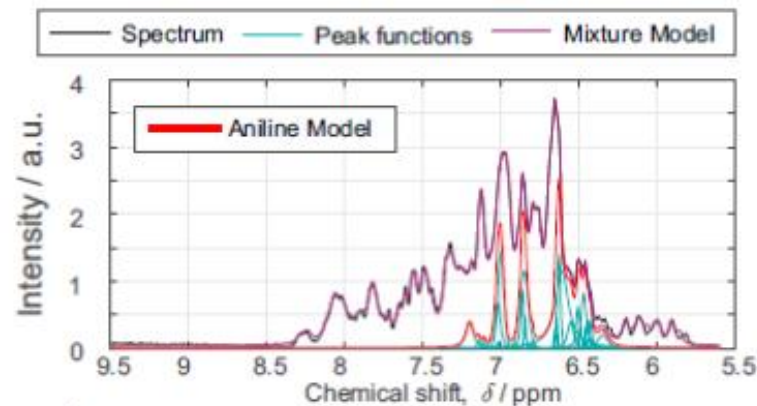
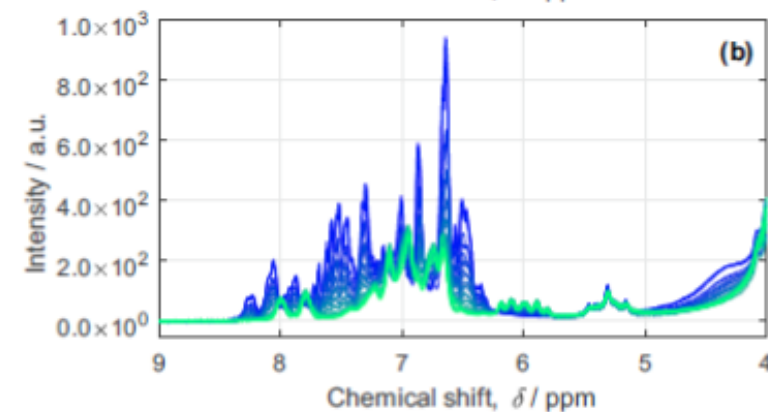
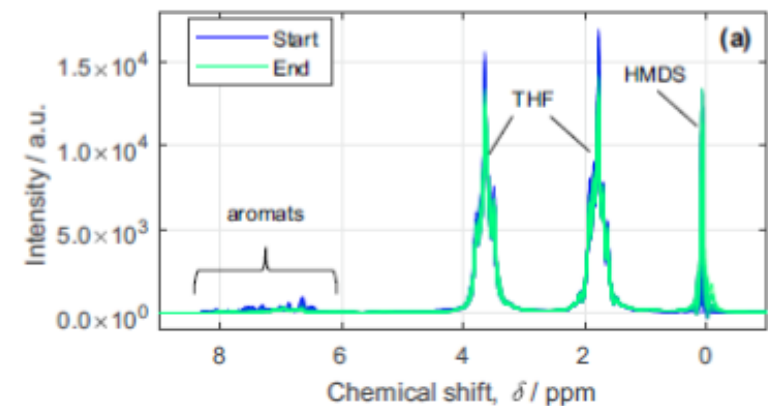


Free resources available online: <https://magritek.com/applications/reaction-monitoring/>



Models to demystify complex mixtures

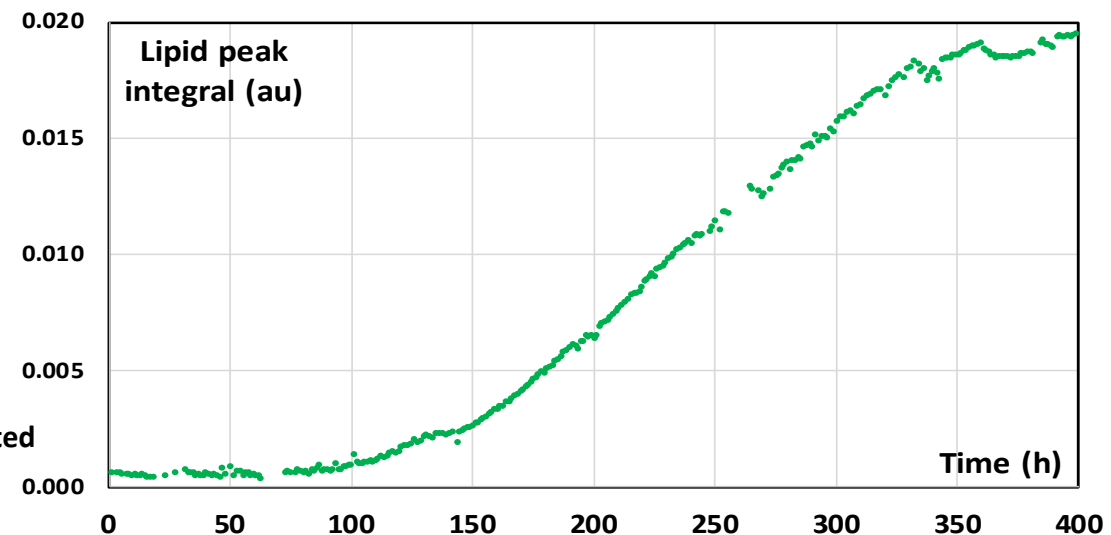
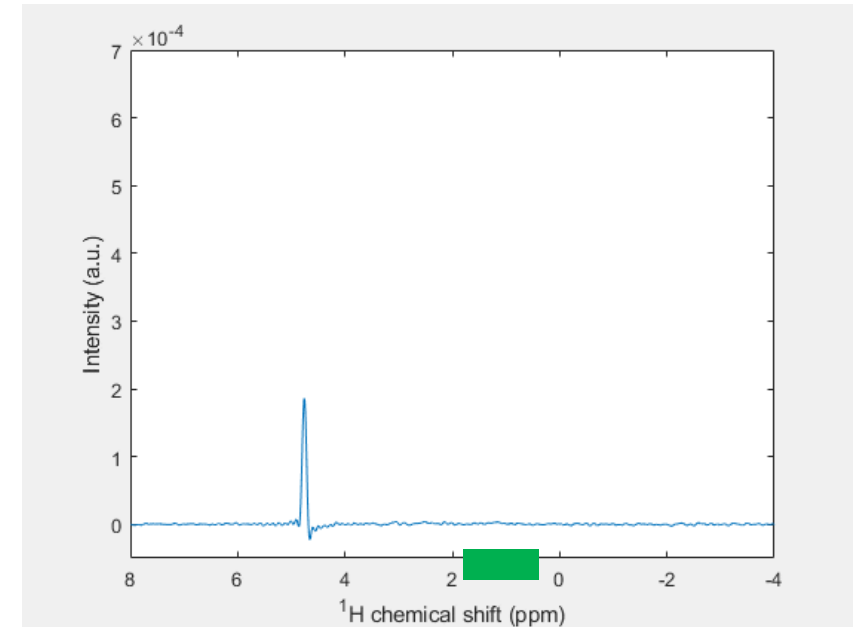
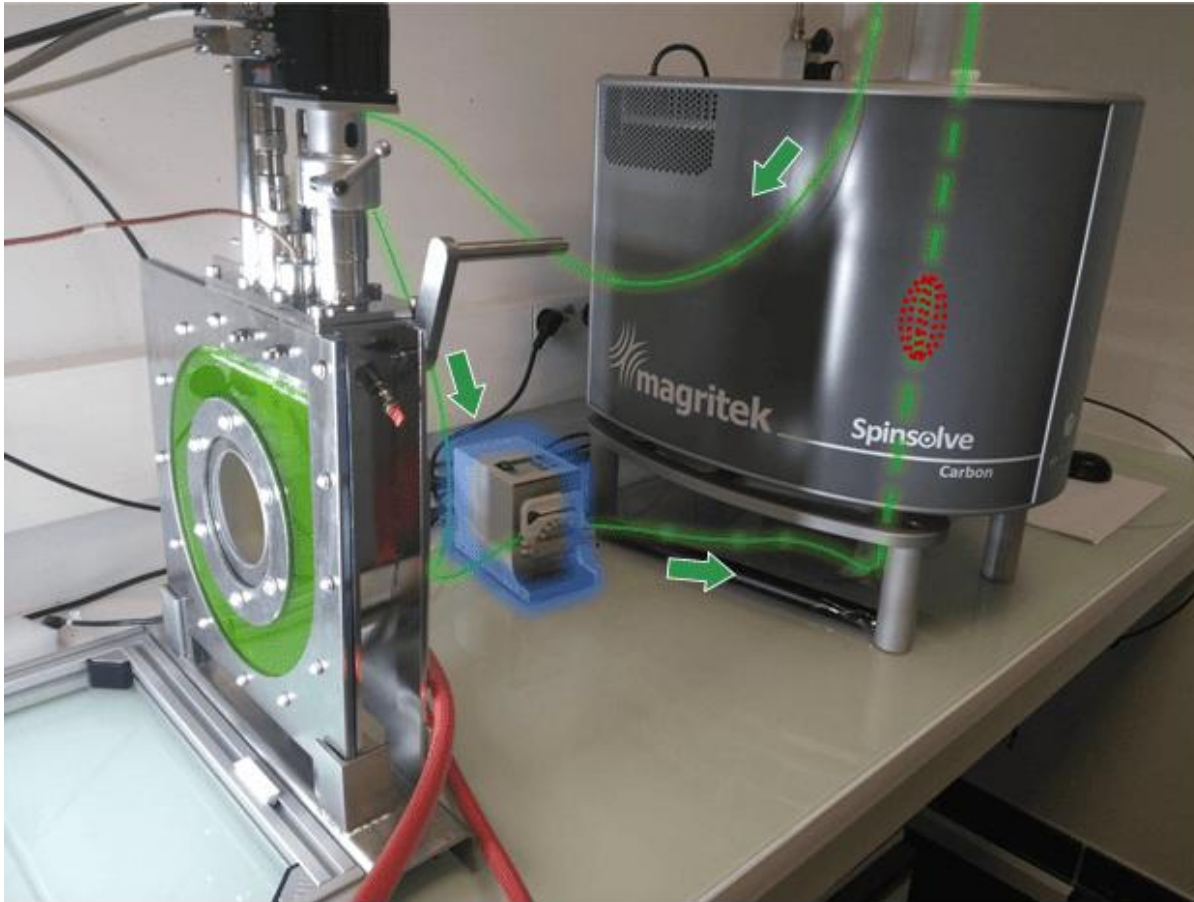
- Indirect Hard Modelling



Online low-field NMR spectroscopy for process control of an industrial lithiation reaction-automated data analysis, S. Kern, K. Meyer, S. Guhl, P. Gräber, A. Paul, R. King, M. Maiwald, *Anal. Bioanal. Chem.*, **2018**, *410*, 3349
[10.1007/s00216-018-1020-z](https://doi.org/10.1007/s00216-018-1020-z)

Monitoring intracellular lipid content

 Spinsolve 43



Using benchtop NMR spectroscopy as an online non-invasive in vivo lipid sensor for microalgae cultivated in photobioreactors, D. Bouillaud, D. Drouin, B. Charrier, C. Jacquemmoz, J. Farjon, P. Giraudeau, O. Gonçalves, *Process Biochemistry*, Volume 93, (2020), <https://doi.org/10.1016/j.procbio.2020.03.016>



Organic chemistry robots



UNIVERSITY OF AMSTERDAM

- Combination of different online detectors
- Feedback loop coupled with algorithm for self-automating chemistry

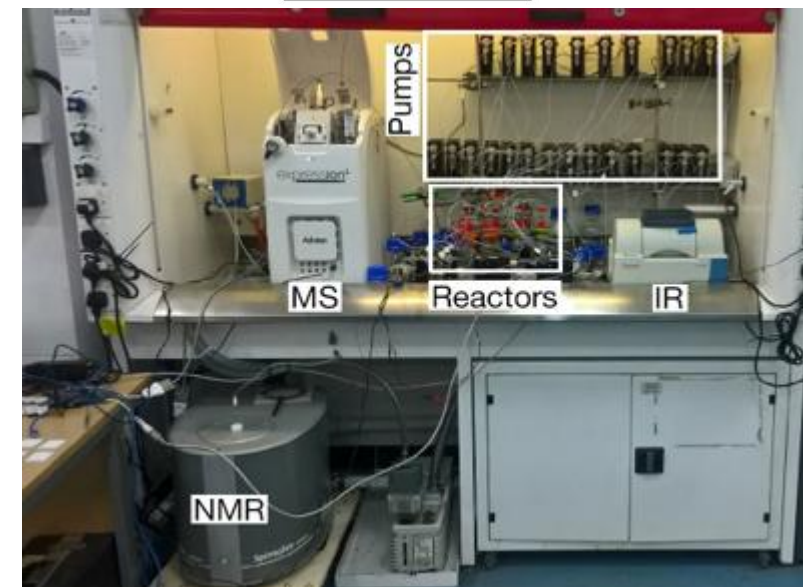
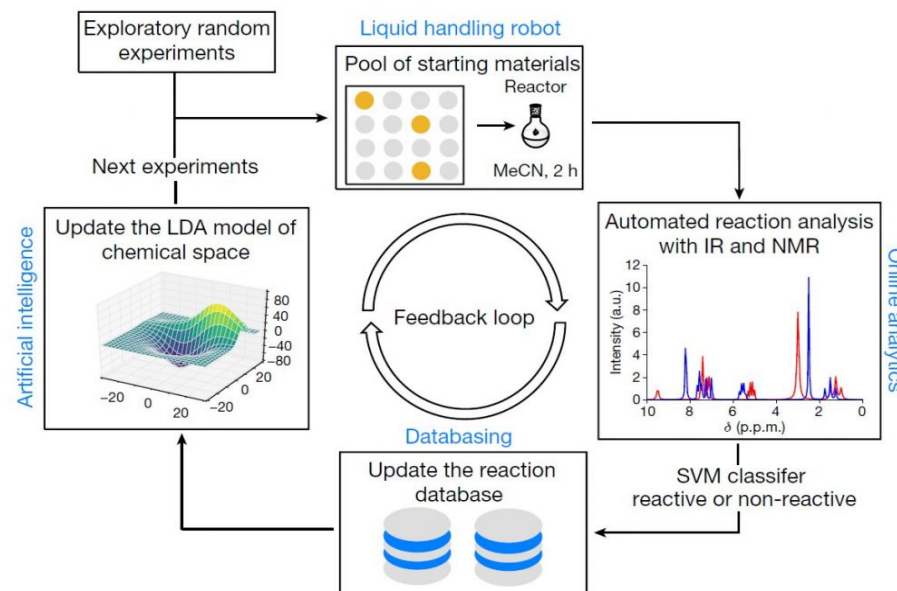


Controlling an organic synthesis robot with machine learning to search for new reactivity; J. M. Granda, L. Donina, V. Dragone, D.-L. Long and L. Cronin, *Nature*, 559, 377–381 (2018)

DOI: [10.1038/s41586-018-0307-8](https://doi.org/10.1038/s41586-018-0307-8)

Automated self-optimization, intensification, and scale-up of photocatalysis in flow; A. Slattery, Z. Wen, P. Tenblad, J. Sanjosé-Orduna, D. Pintossi, T. den Hartog and T. Noël, *Science*, 383, (2024).

DOI: [10.1126/science.adj1817](https://doi.org/10.1126/science.adj1817)



Benchtop NMR applications



Synthesis

Pharma

Macromolecules

Education

Flow chemistry



Batteries

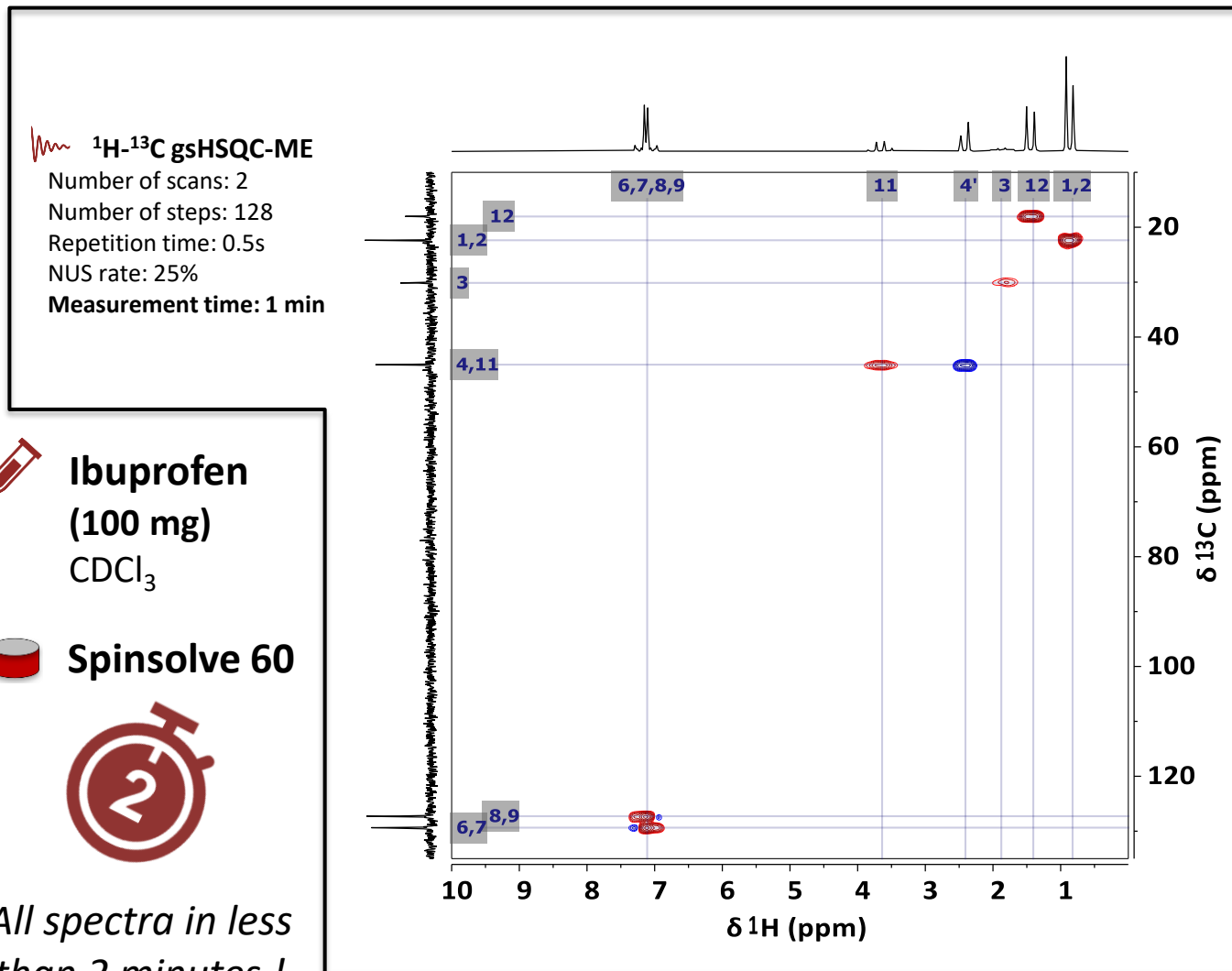
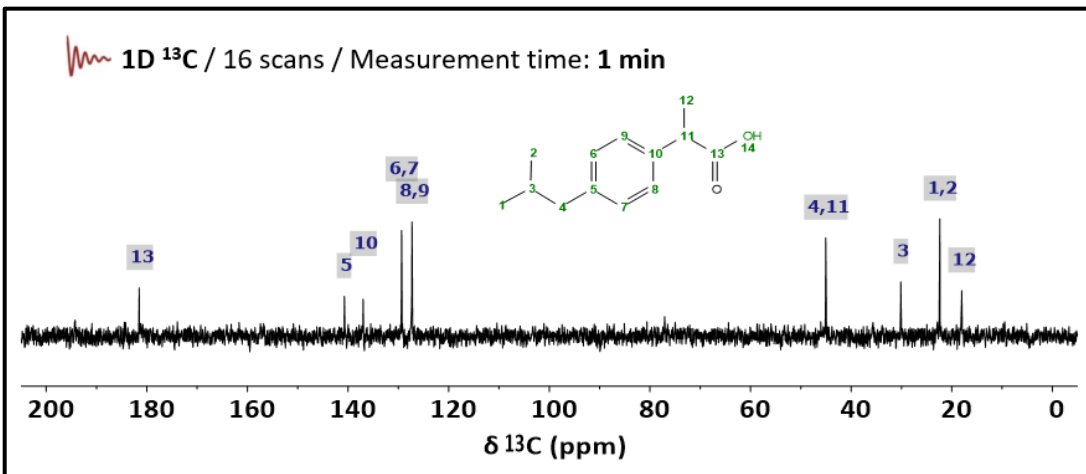
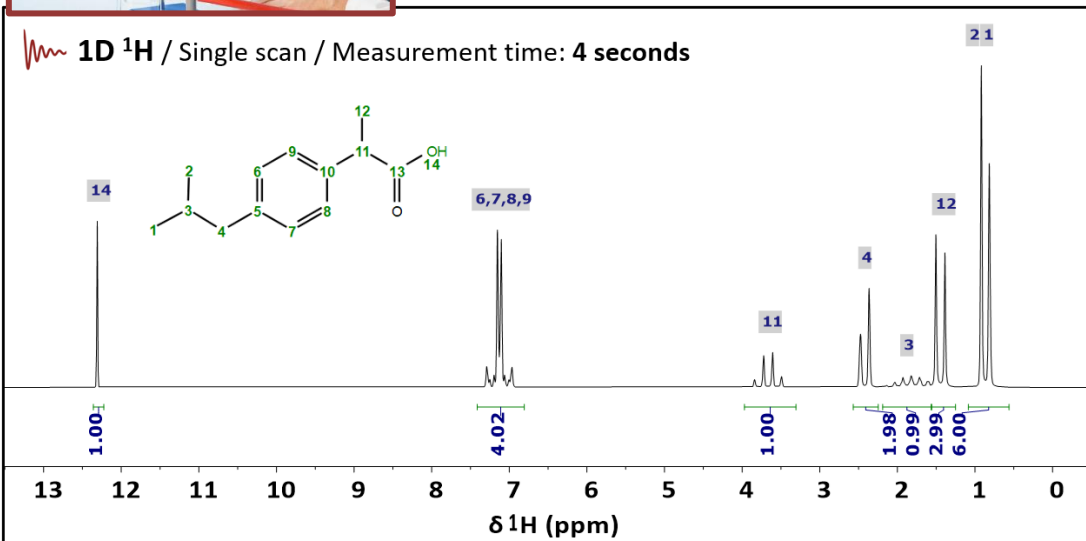
Food & Agriculture


Forensics

Analytics



Teaching structural elucidation



 **Ibuprofen**
(100 mg)
 CDCl_3

 **Spinsolve 60**



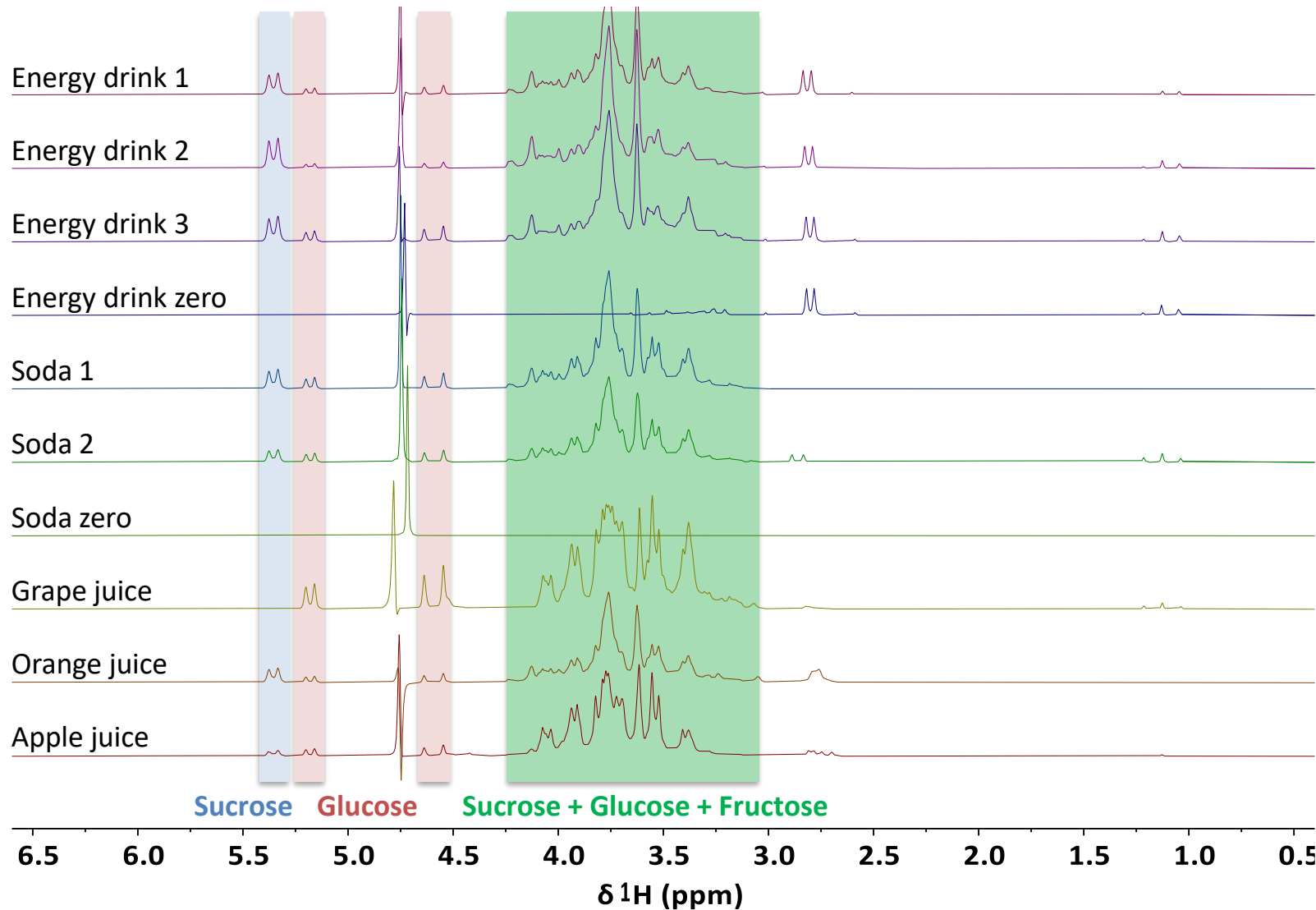
All spectra in less than 2 minutes !



Teaching qNMR - Soft drink analysis

1D ¹H (WET)/Repetition time: 10 s/Number of scans: 8/Measurement time: 2 min each

- Low sample preparation

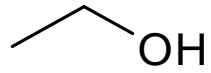


Name	Glucose [g/100 mL]	Sucrose [g/100 mL]	Fructose [g/100 mL]	Total sugars [g/100 mL]
Energy Drink 1	5.20	2.10	2.05	8.88
Energy Drink 2	7.00	1.39	1.94	9.86
Energy Drink 3	6.03	3.24	1.56	10.36
Energy Drink zero	0.01	0.01	0.47	0.01
Cola 1	4.46	3.20	2.91	10.57
Cola 2	2.51	2.33	2.15	6.99
Cola without sugar	0.00	0.01	0.03	0.05
Grape Juice	0.08	8.03	7.99	16.10
Orange Juice	3.77	2.11	2.78	8.65
Apple Juice	1.53	2.47	6.18	10.18

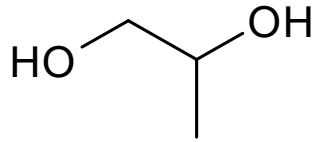


Teaching qNMR - Soft drink analysis

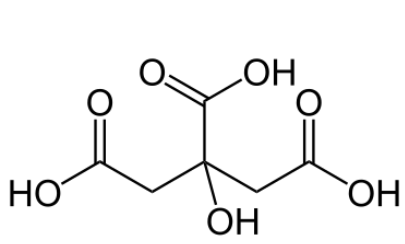
1D ¹H (WET)/Repetition time: 10 s/Number of scans: 128/Measurement time: 20 min each



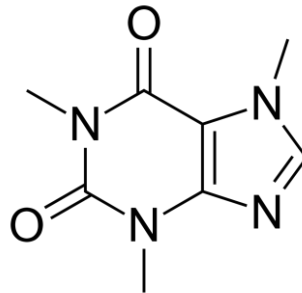
Ethanol



Propylene glycol (E1520)

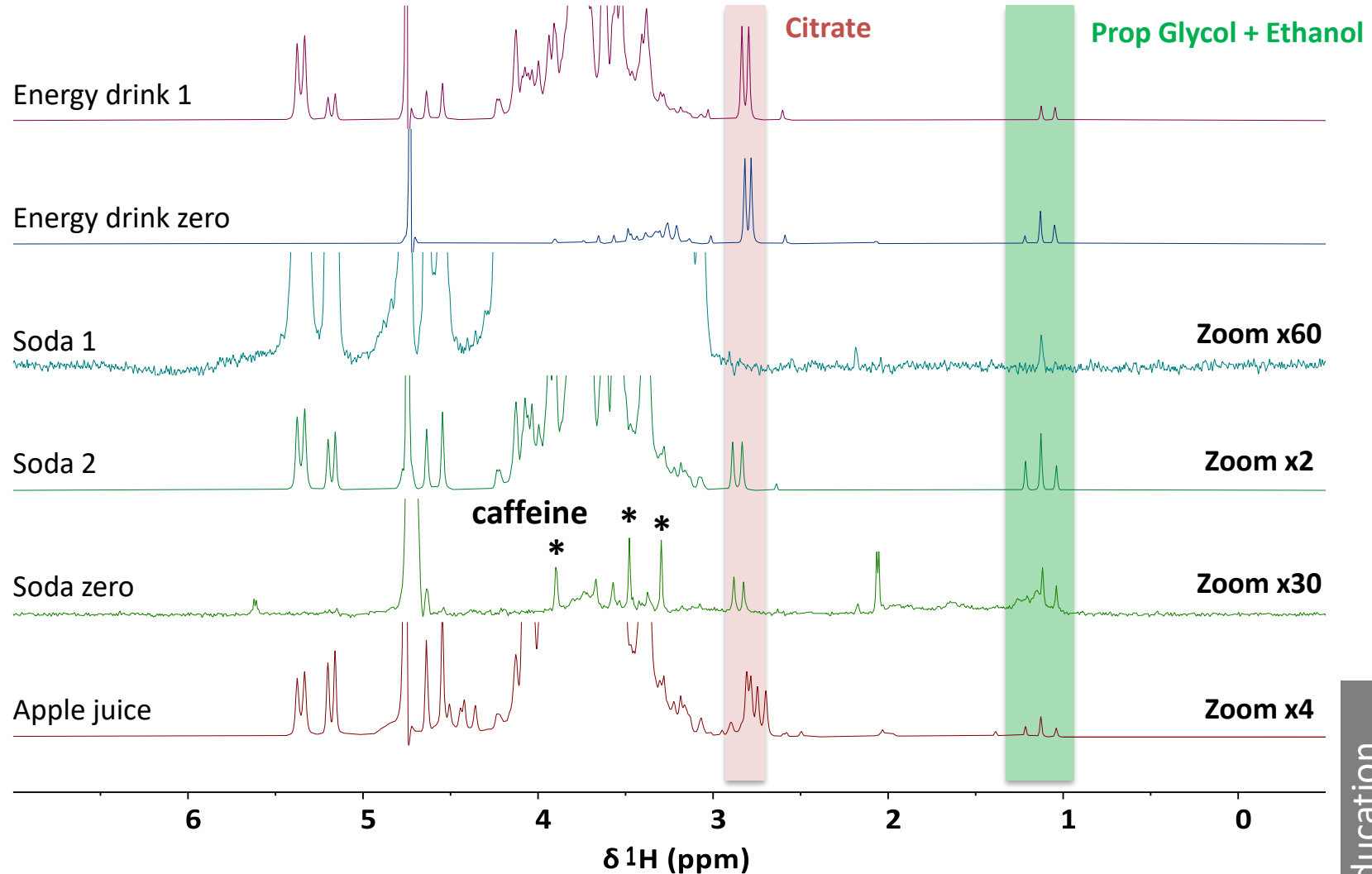


Citrate



Caffeine

- Different quantification approaches possible: internal/external standard, standard addition

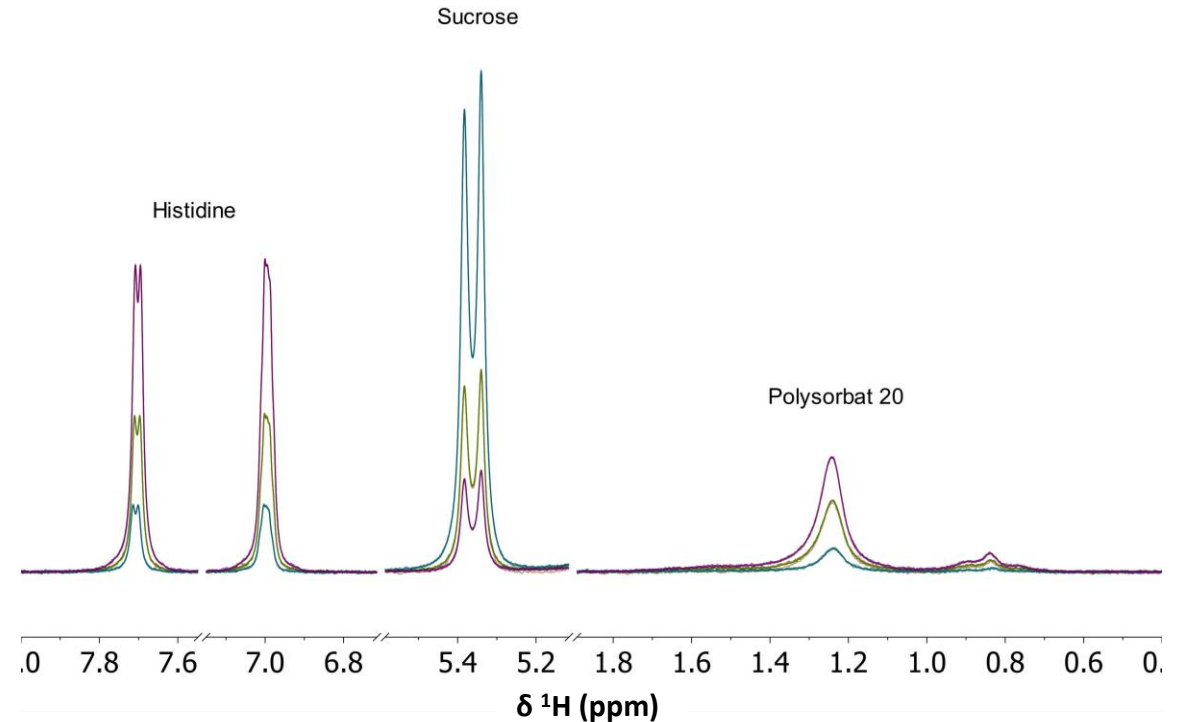
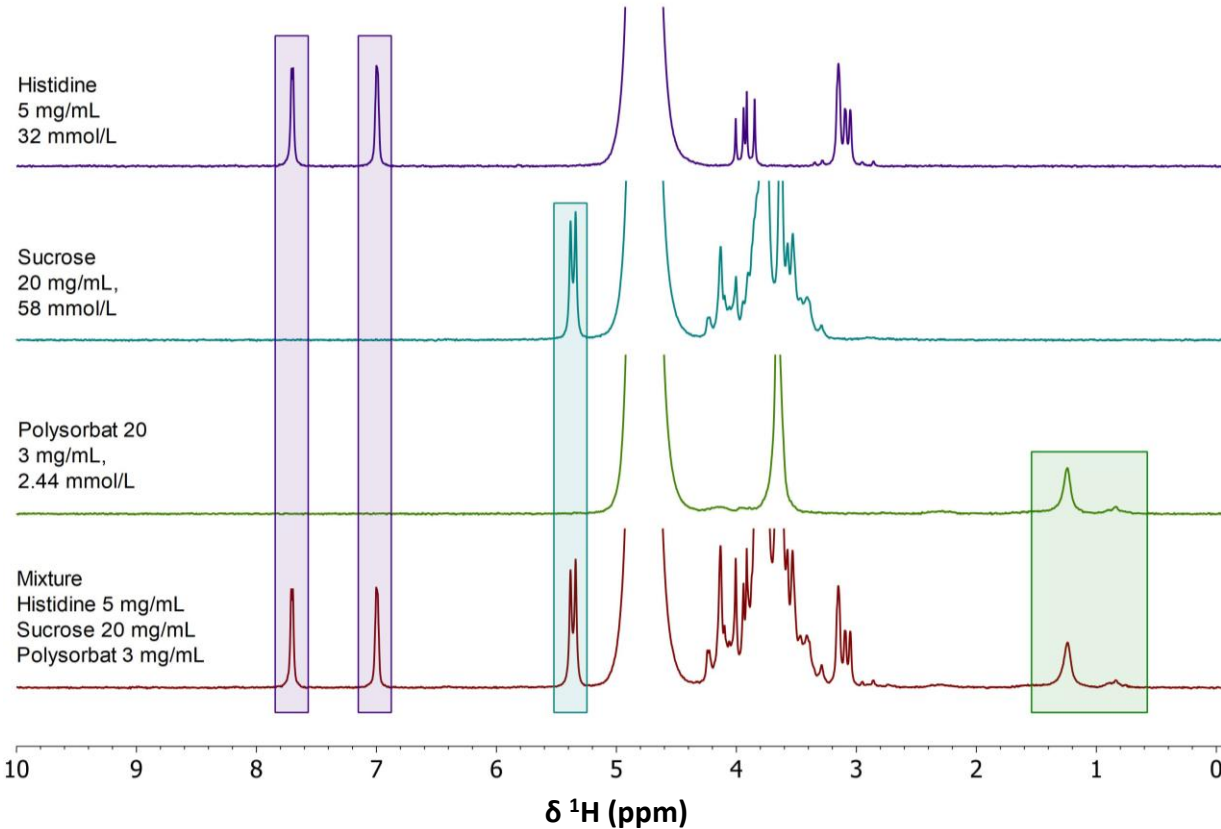


Free resources available online: <https://magritek.com/2023/12/08/quantifying-residual-solvents-in-active-pharmaceutical-ingredients/>



Teaching qNMR - Pharmaceutical formulation

1D ^1H (PRESAT)/Repetition time: 10 s/Number of scans: 32/Measurement time: 6 min each



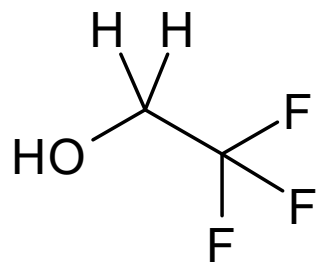
- 3 reference spectra (known concentrations) and a spectrum of a mixture measured on a Spinsolve 80 spectrometer.

- Superposition of three mixtures with different concentrations of histidine, sucrose and polysorbate 20.

Free resources available online: <https://magritek.com/2021/02/22/qnmr-on-samples-from-the-production-plant/>



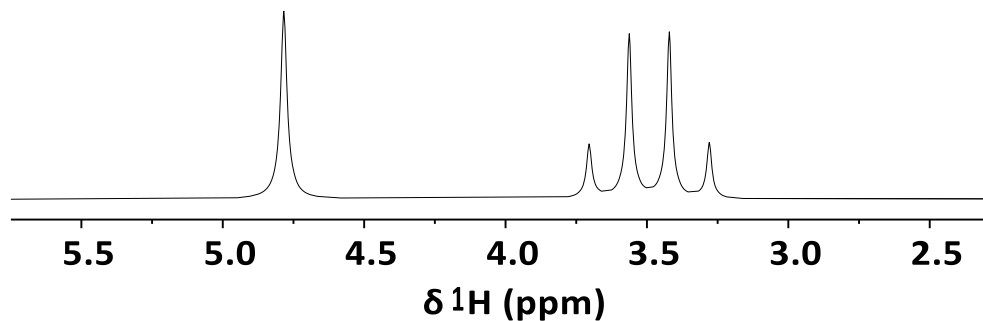
Teaching NMR - Couplings with ^{19}F



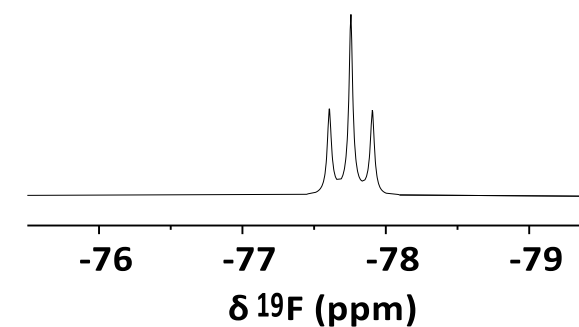
2,2,2-trifluoroethanol

- Full $^1\text{H}/^{19}\text{F}$ decoupling options to understand the couplings, simplify the spectra and get higher SNR

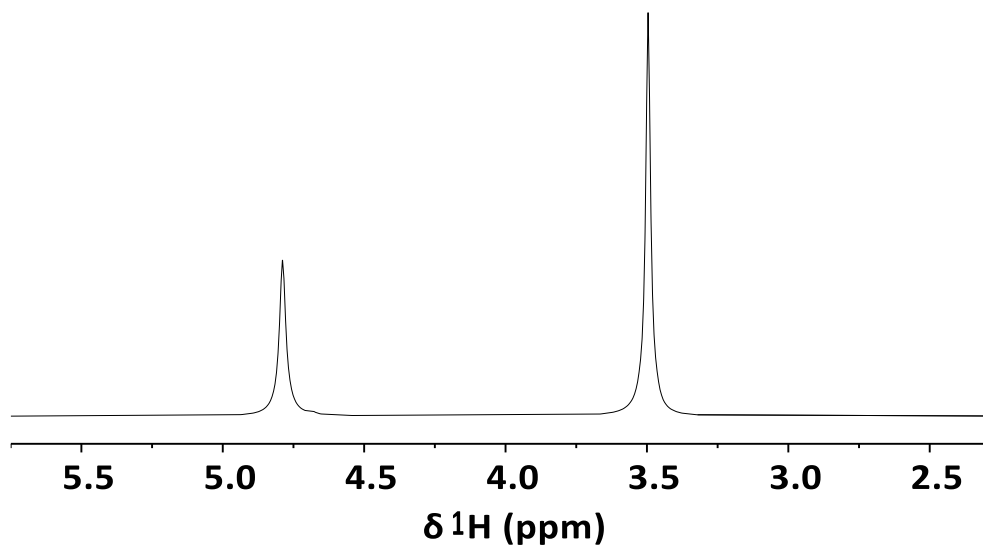
^1H /Single scan/Measurement time: 5 s



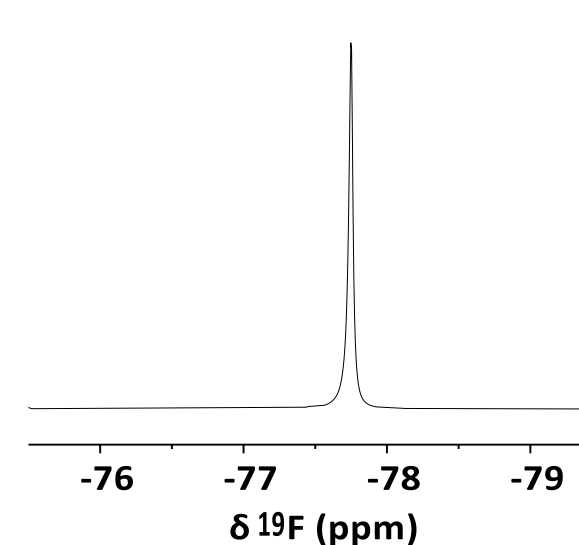
^{19}F /Single scan/Measurement time: 5 s



$^1\text{H}\{^{19}\text{F}\}$ /Single scan/Measurement time: 5 s

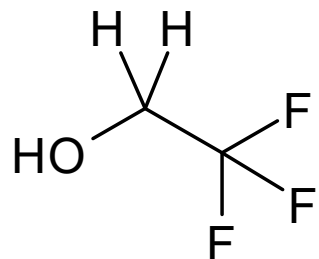


$^{19}\text{F}\{^1\text{H}\}$ /Single scan/Measurement time: 5 s





Teaching NMR - Couplings with ^{19}F



2,2,2-trifluoroethanol

- ^1H **AND** ^{19}F decoupling for ^{13}C spectra
- Speeding up fluorinated molecules analysis

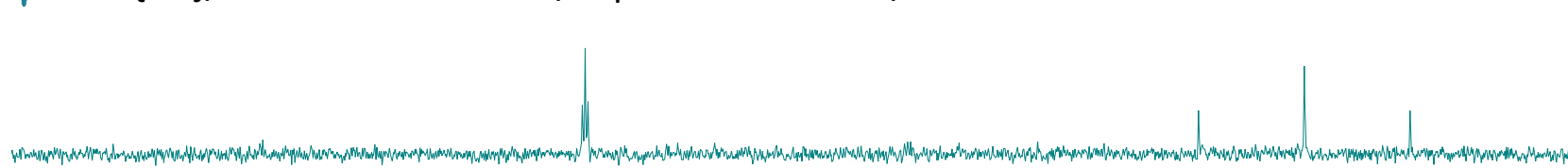
^{13}C /Number of scans: 2048/Repetition time: 3 s/Measurement time: 3h30



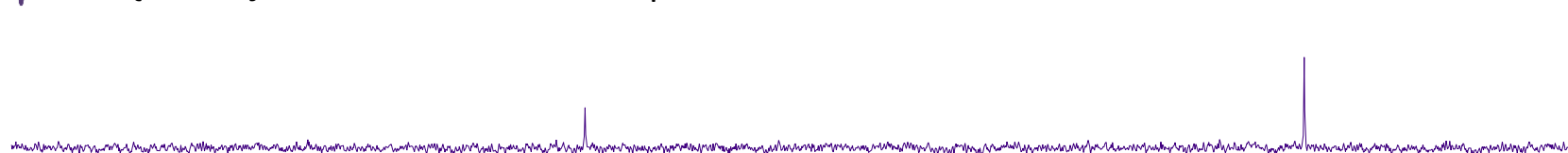
$^{13}\text{C}\{^1\text{H}\}$ /Number of scans: 128/Repetition time: 3 s/Measurement time: 6 min



$^{13}\text{C}\{^{19}\text{F}\}$ /Number of scans: 64/Repetition time: 3 s/Measurement time: 3 min



$^{13}\text{C}\{^1\text{H}+^{19}\text{F}\}$ /Number of scans: 4/Repetition time: 3 s/Measurement time: 12 s



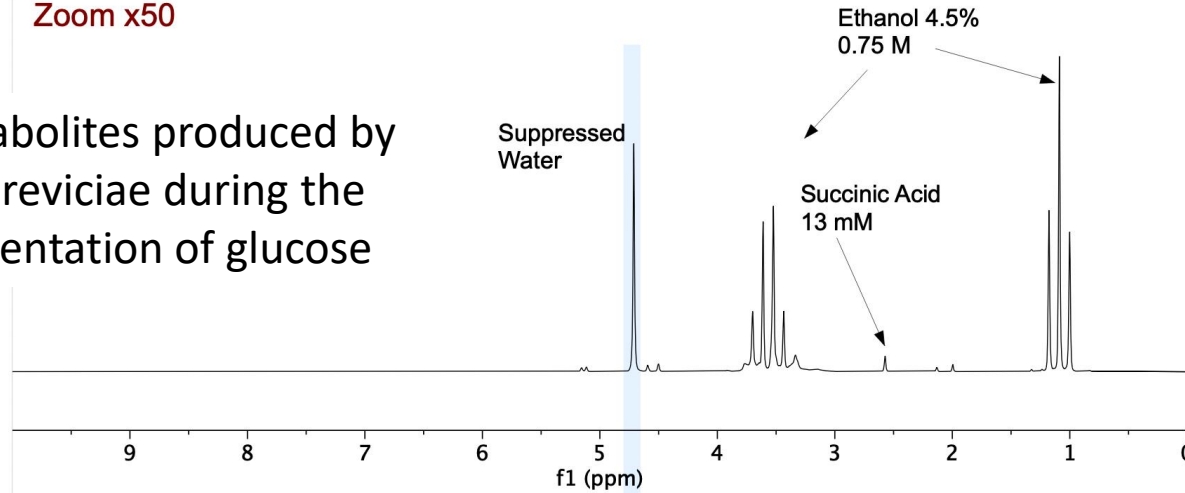
170 160 150 140 130 120 110 100 90 80 70 60 50 40
 $\delta^{13}\text{C}$ (ppm)



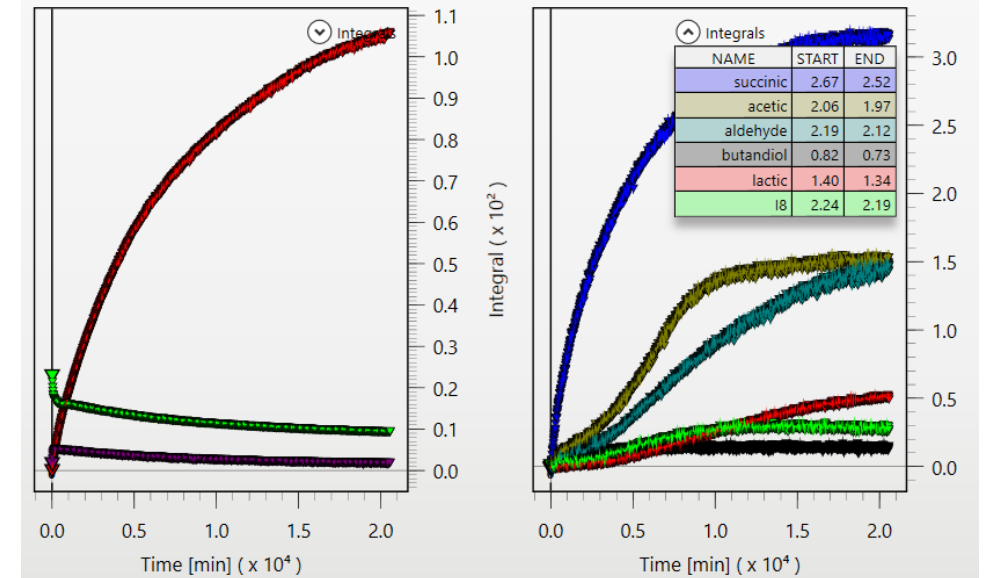
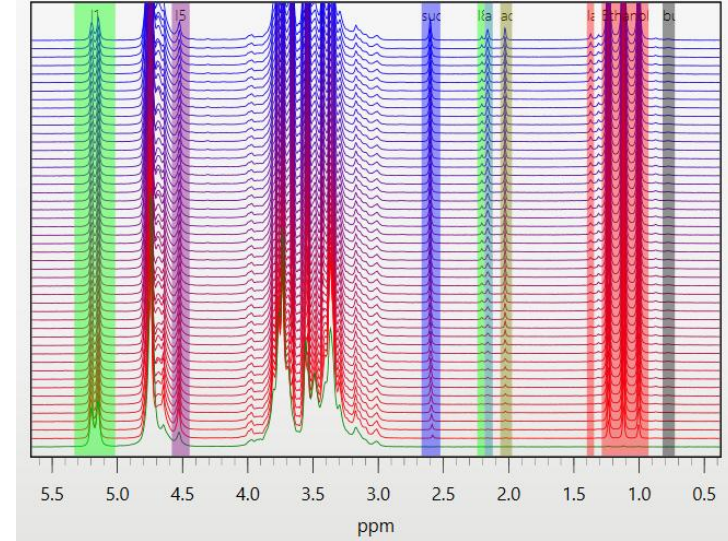
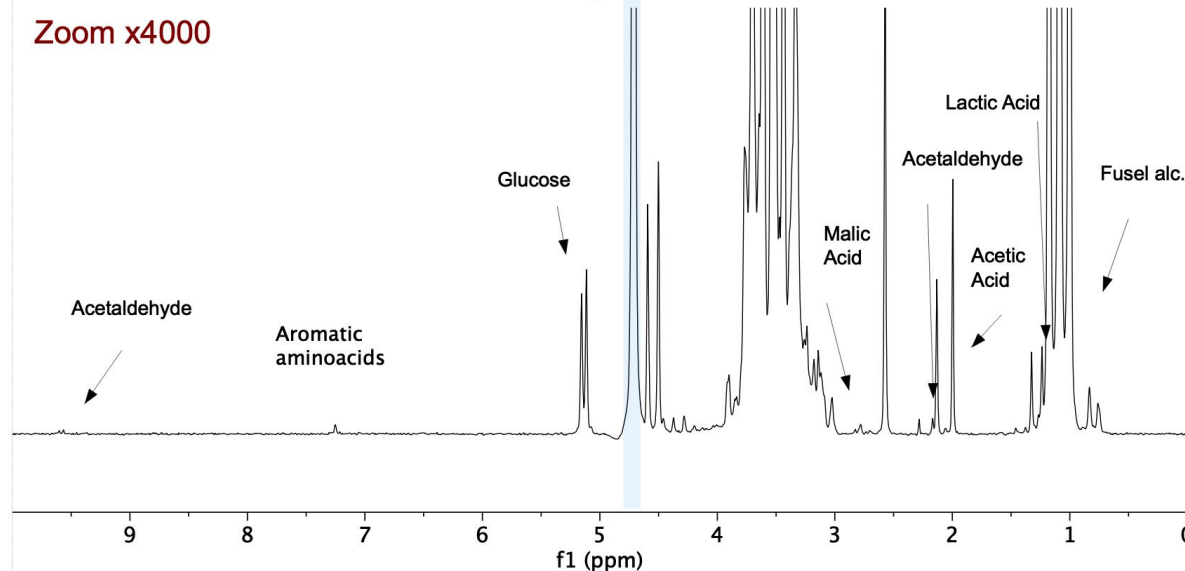
Teaching biochemistry: monitoring fermentations

Zoom x50

Metabolites produced by *S. Cereviciae* during the fermentation of glucose



Zoom x4000



• Monitoring over 35h

Benchtop NMR applications



Synthesis

Pharma

Macromolecules

Education

Flow chemistry



Batteries

Food & Agriculture

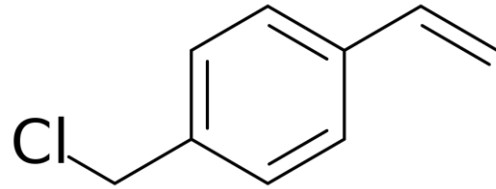
Forensics

Analytics



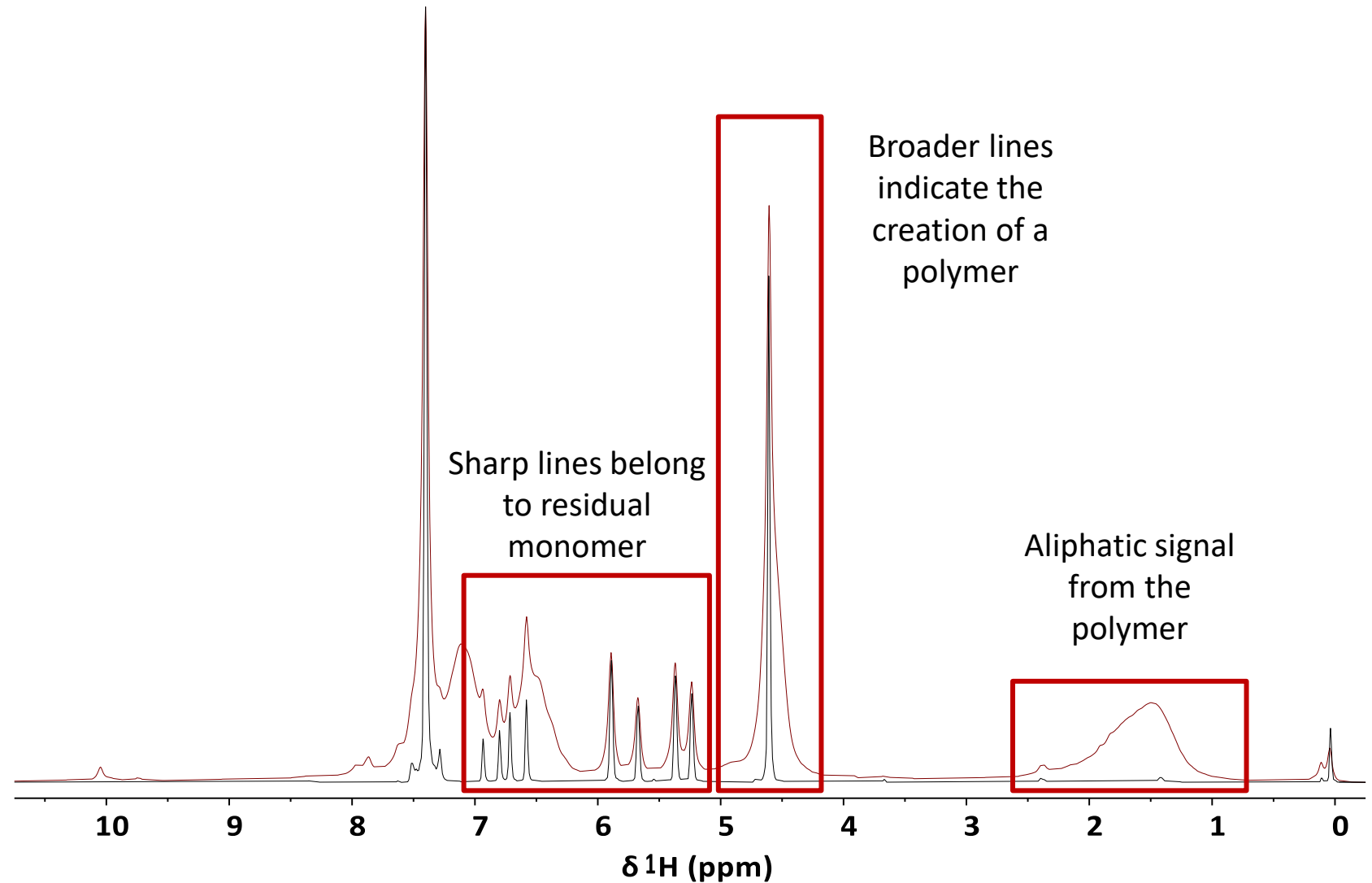
Quality check of a monomer solution

1D ^1H / Repetition time: 10 s / Number of scans: 1 / Measurement time: 10 s



4-vinylbenzyl chloride

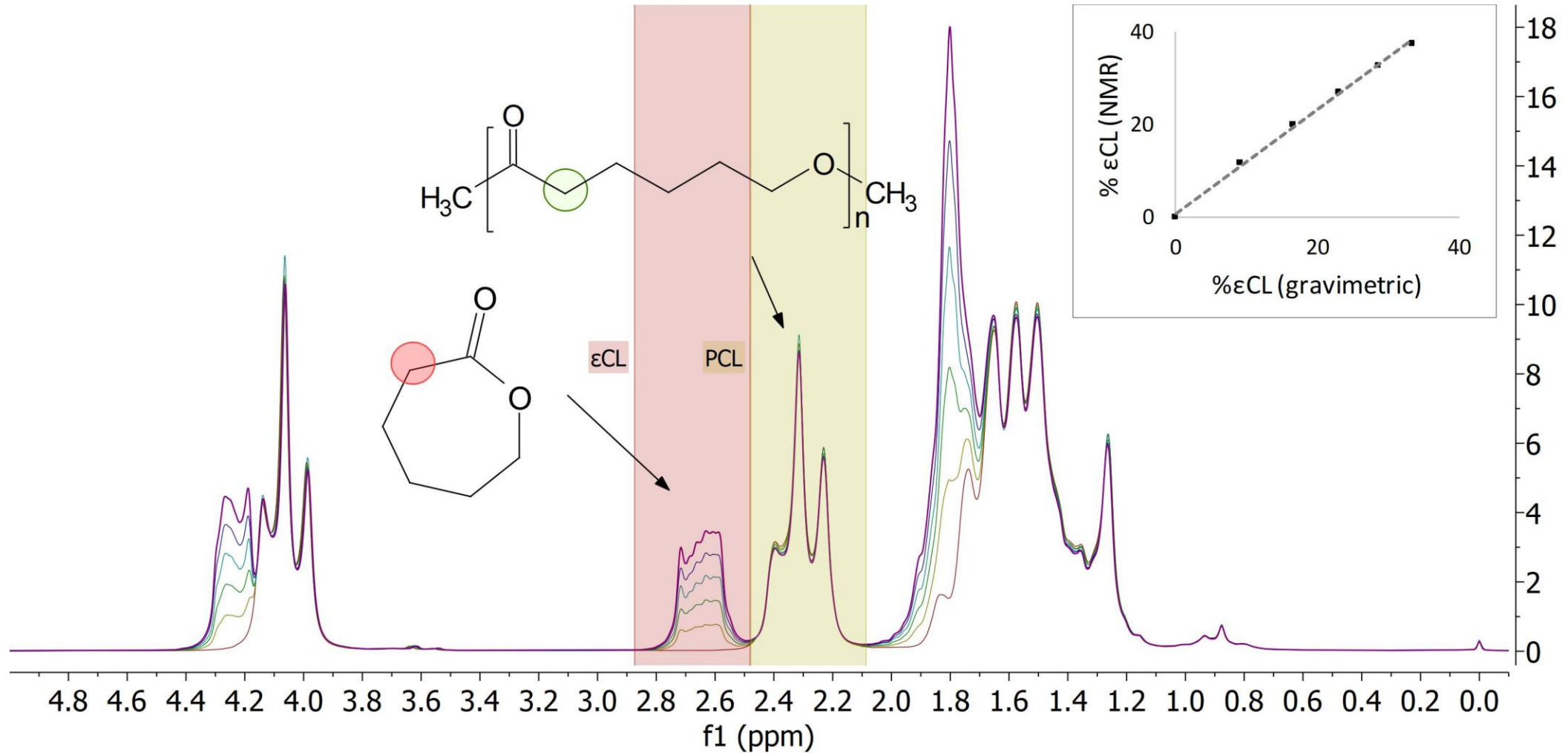
— Fresh sample
— Sample after a few months without storage care (room temperature)





Conversion during the polymerization

Signals of polymer and monomers should be separated in the spectra





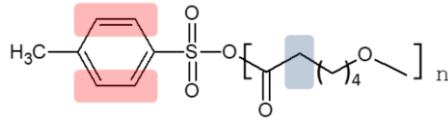
Polymer end-group analysis

$$M_n = DP \times M_{\text{repeating unit}}$$

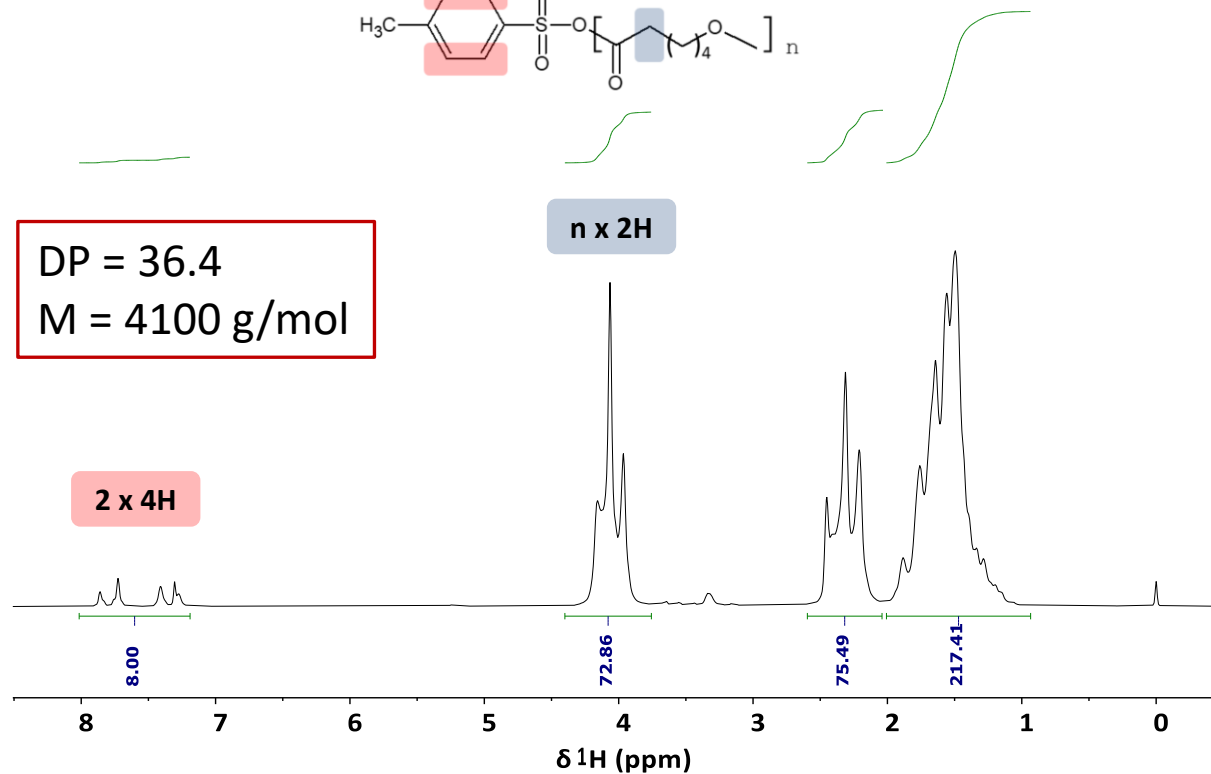
$$DP = \frac{\text{Integral of the repeating unit}}{\text{number of protons of the repeating unit}} \times \frac{\text{number of protons of the end group}}{\text{integration of the end group}}$$

Polycaprolactone

1D ¹H/Repetition time: 10 s/Number of scans: 16/Measurement time: 3 min

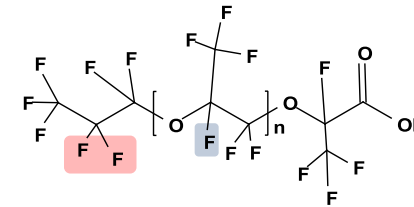


DP = 36.4
M = 4100 g/mol

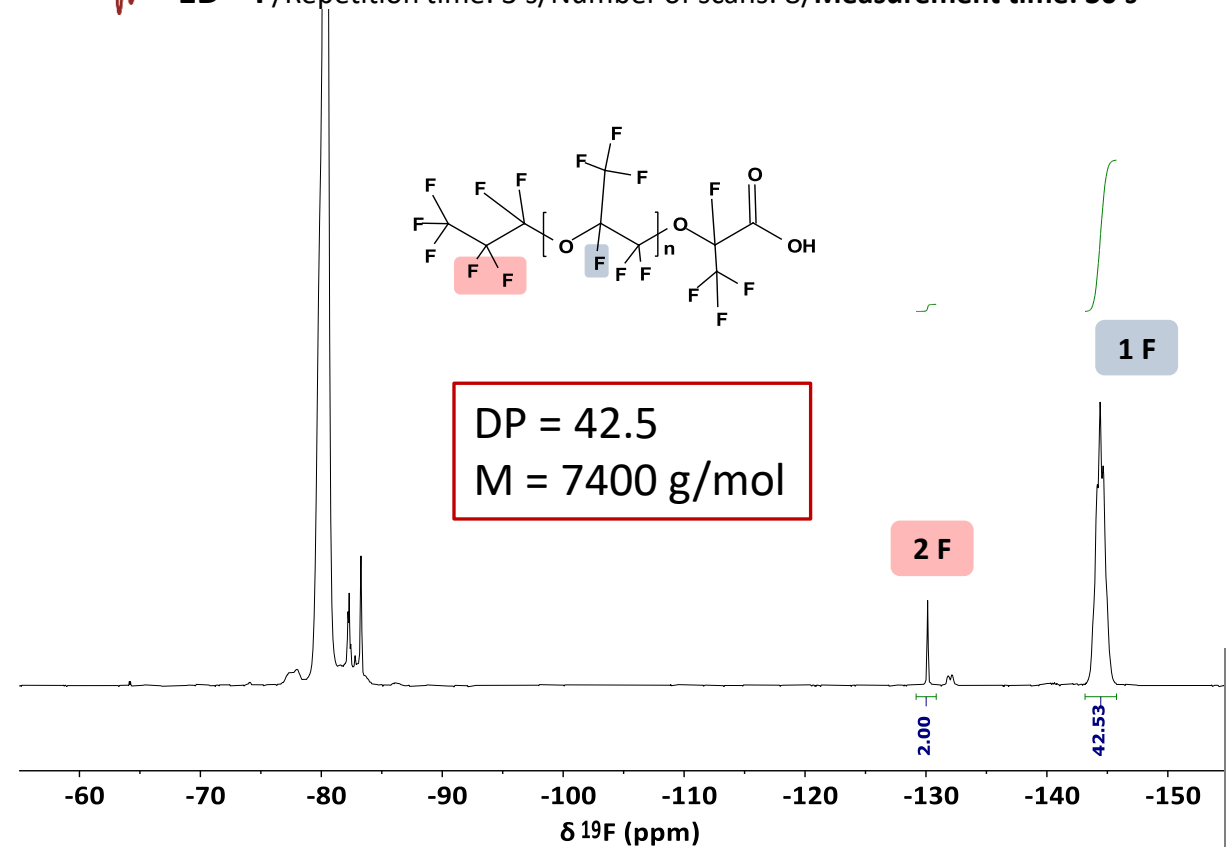


Perfluoropolyether

1D ¹⁹F/Repetition time: 3 s/Number of scans: 8/Measurement time: 30 s



DP = 42.5
M = 7400 g/mol



Free resources available online: <https://magritek.com/applications/polymers/>

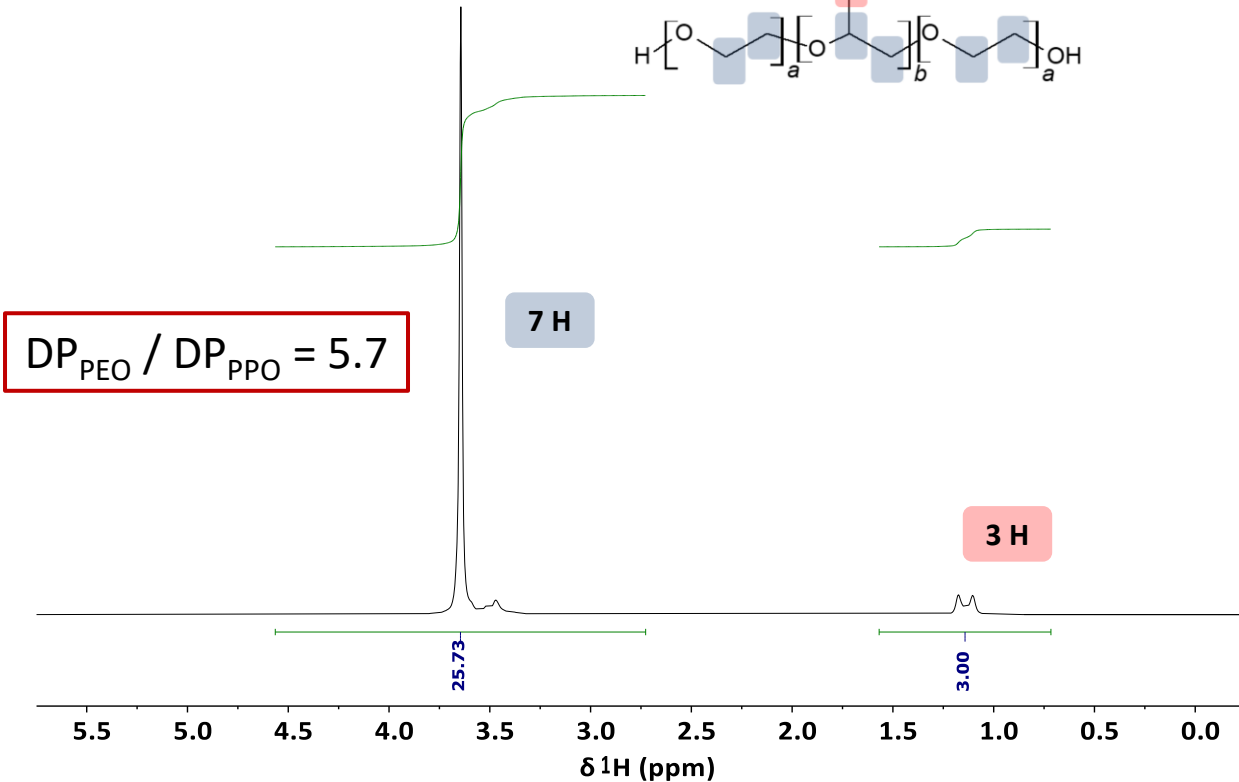
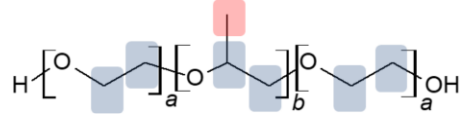


Copolymer ratio

$$\frac{DP_1}{DP_2} = \frac{\text{Integral of the repeating unit 1}}{\text{number of } ^1\text{H of the repeating unit 1}} \times \frac{\text{number of } ^1\text{H of the repeating unit 2}}{\text{Integral of the repeating unit 2}}$$

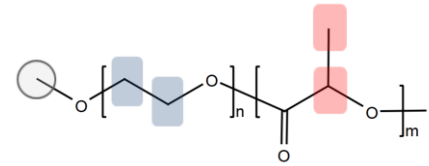
Poloxamer

1D ¹H/Repetition time: 10 s/Number of scans: 16/Measurement time: 3 min

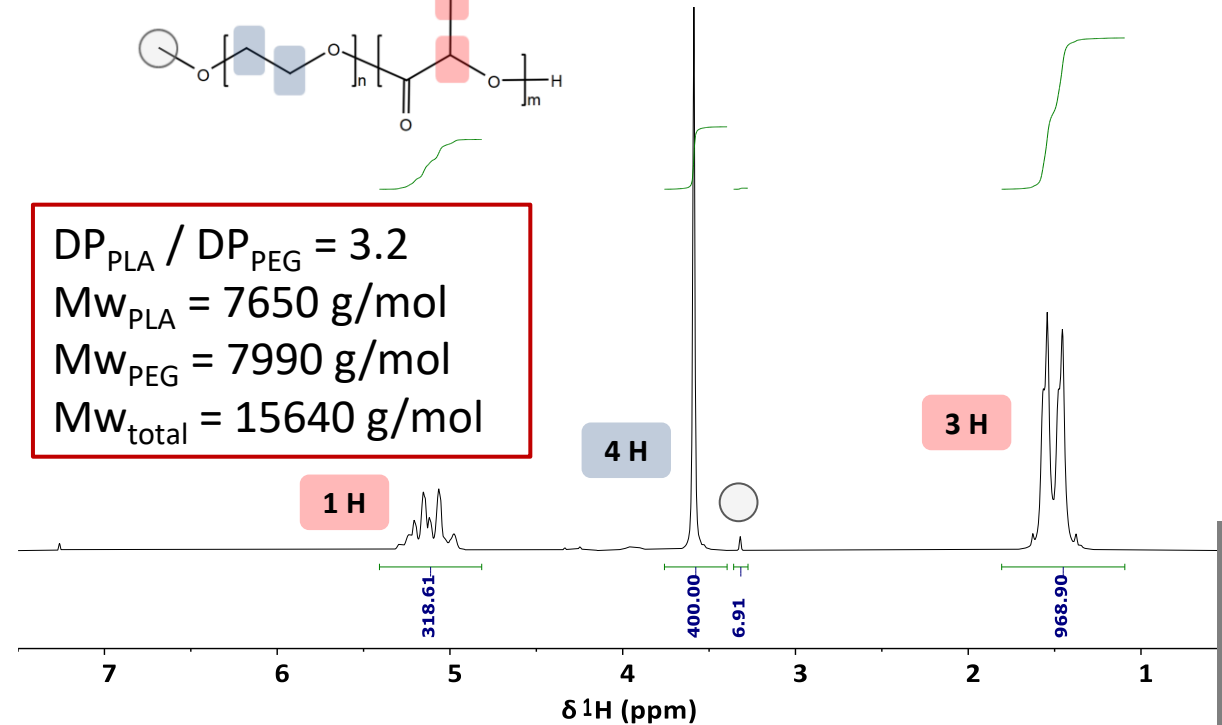


Poly(ethylene glycol)-b-poly(lactic acid)

1D ¹H/Repetition time: 10 s/Number of scans: 16/Measurement time: 3 min



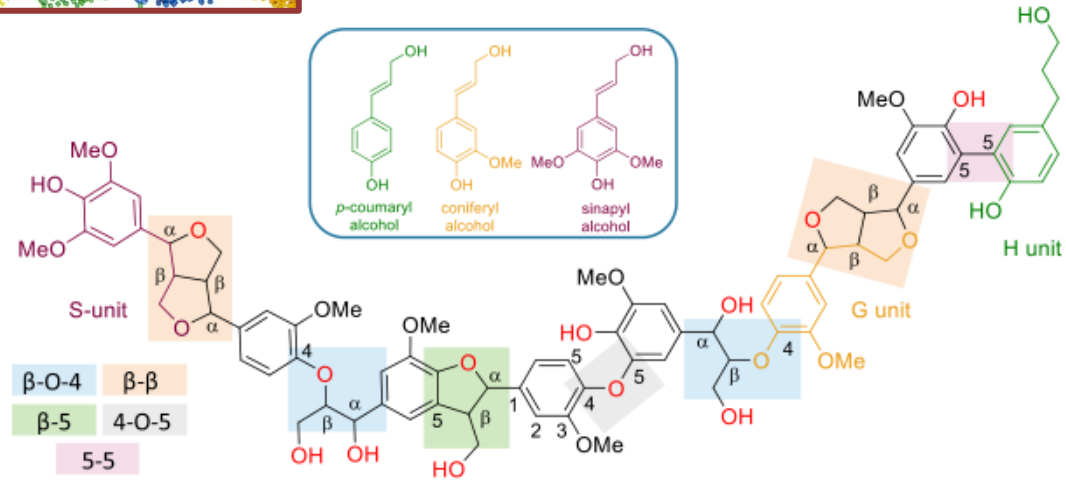
$DP_{PLA} / DP_{PEG} = 3.2$
 $Mw_{PLA} = 7650 \text{ g/mol}$
 $Mw_{PEG} = 7990 \text{ g/mol}$
 $Mw_{total} = 15640 \text{ g/mol}$



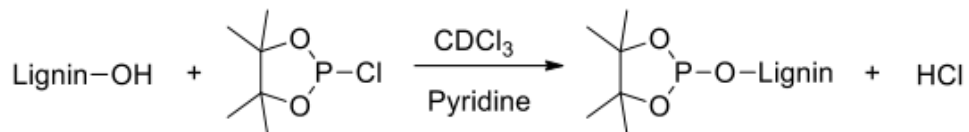
Free resources available online: <https://magritek.com/applications/polymers/>



Quantification of lignin -OH groups

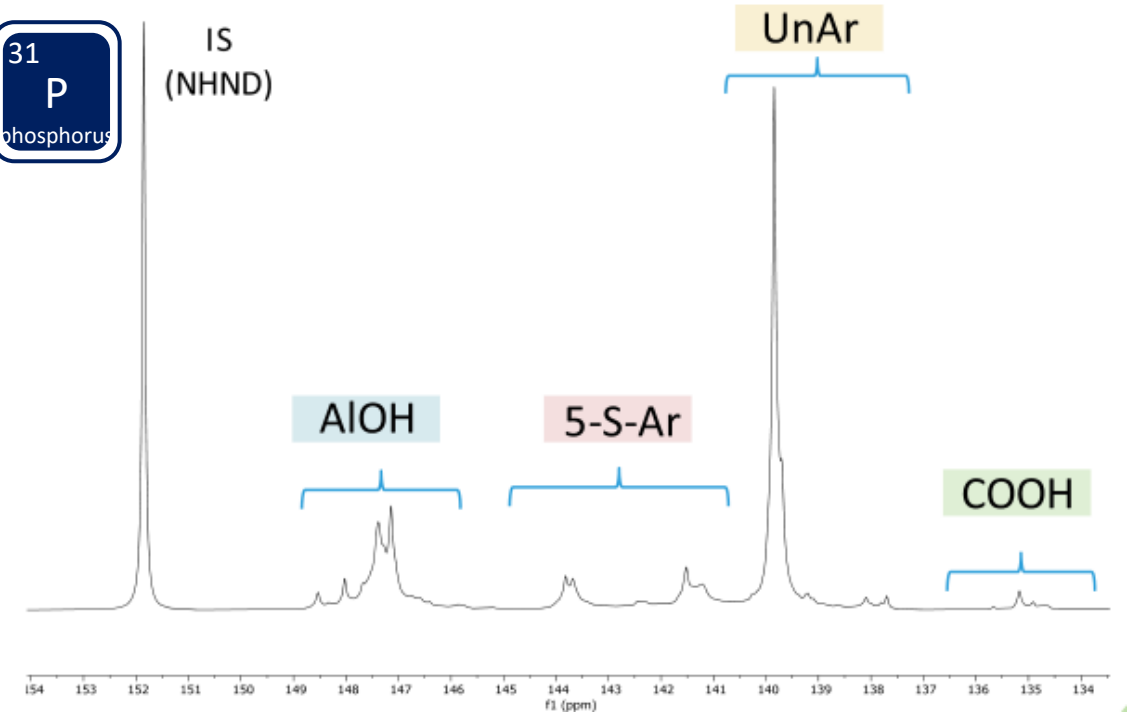


Representative structure of lignin with the most common bonds



Derivatization of -OH group to introduce a phosphorous group

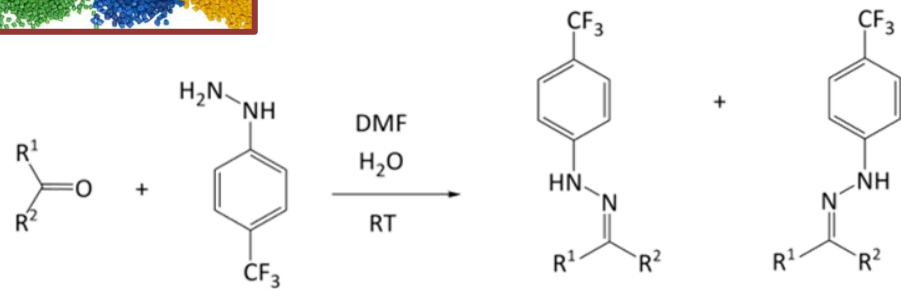
Spinsolve 80



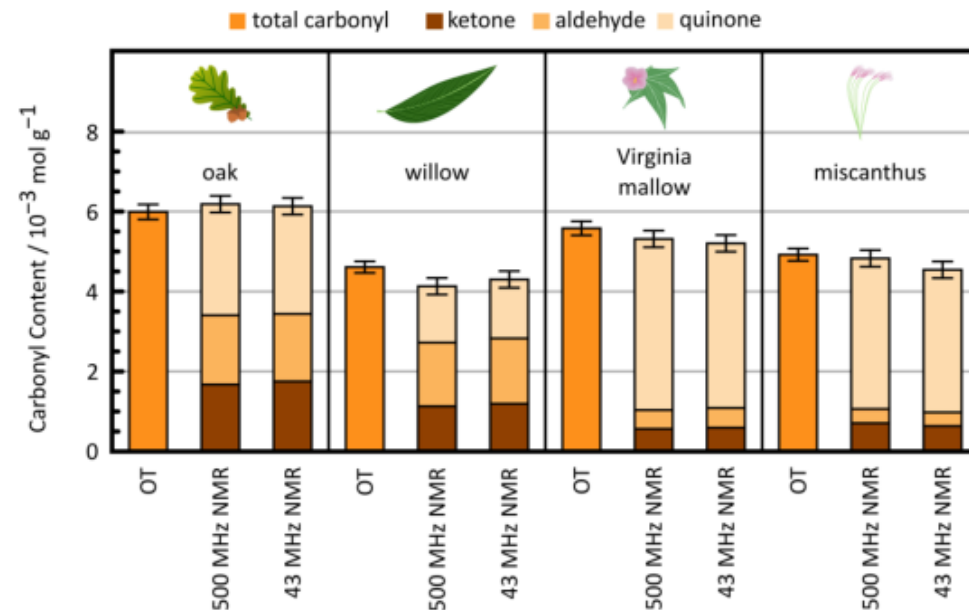
- Determination of lignin -OH groups
- Results in accordance with High-Field NMR



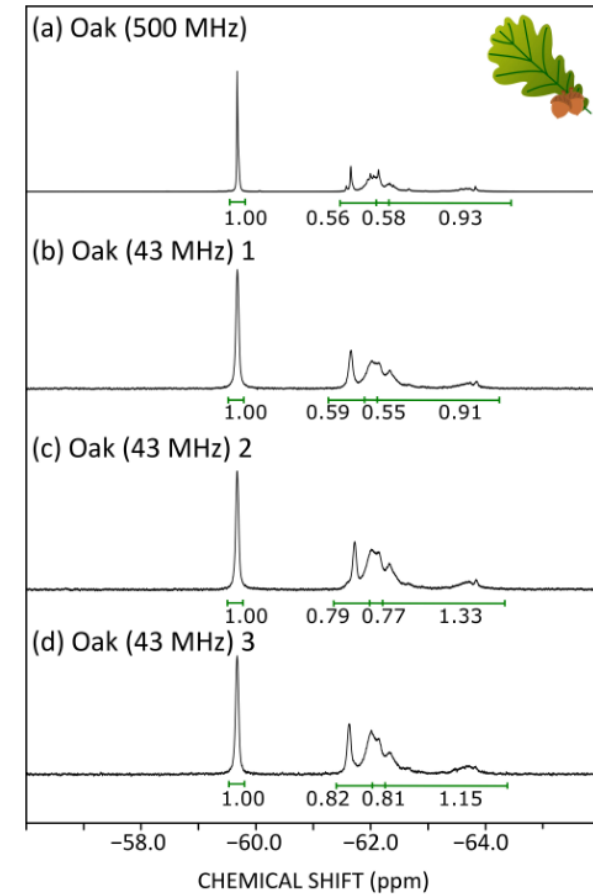
Quantification of pyrolysis oils carbonyl groups



Derivatization of $-C=O$ group to introduce a fluorinated group



Spinsolve 43



- Quantification of carbonyl groups
- Results in accordance with High-Field NMR & titration

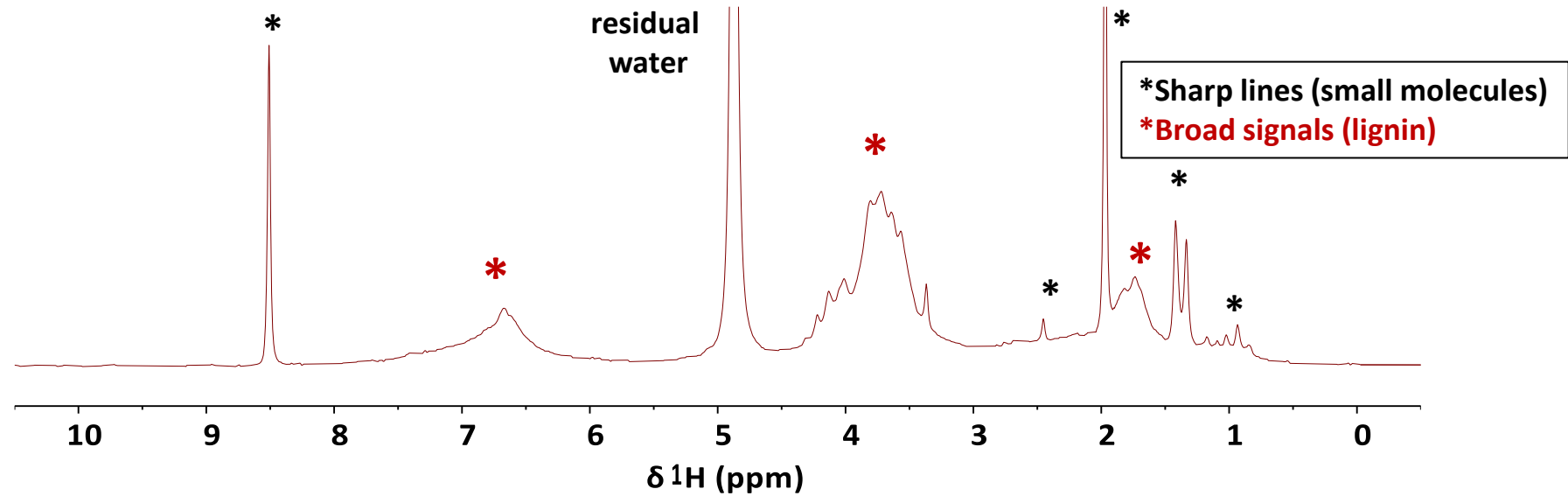



Black liquor analysis

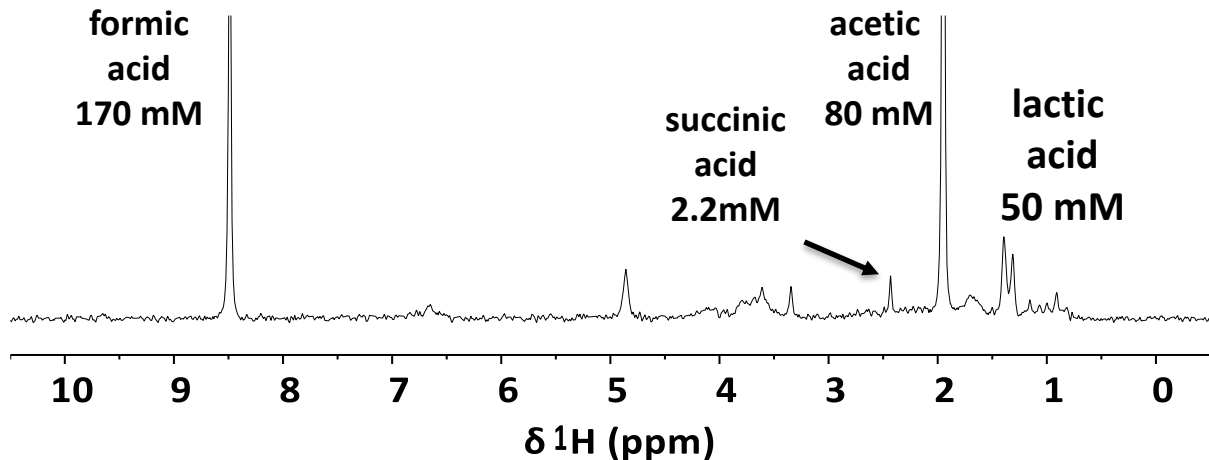
 **Spinsolve 80**

 **¹H WET (solvent suppression)**

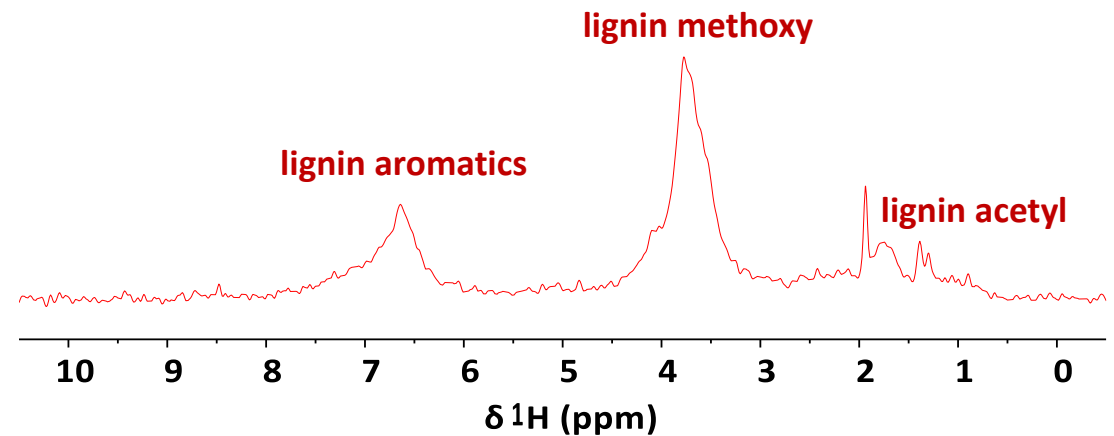
- Black liquor is a waste of the kraft paper industry



 **WET with T2 filter (small molecules only)**

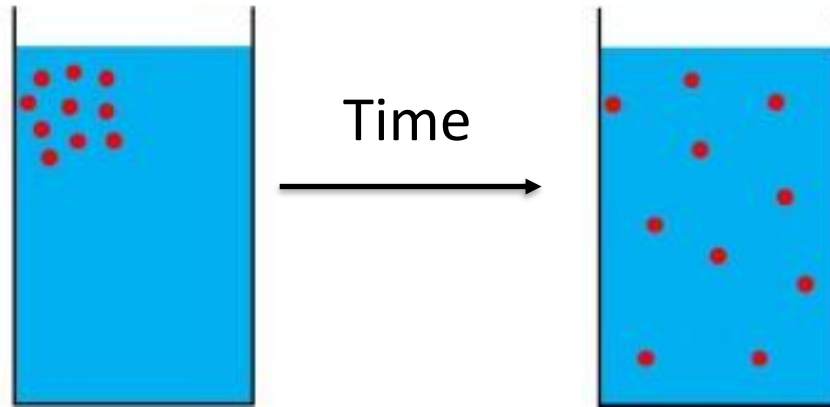


 **¹H with diffusion filter (large molecules only)**





Diffusion of (macro)molecules



Diffusion - Brownian motion

- In solution, every (macro)molecule diffuse
- Speed of diffusion depend on different parameters, and the Stokes-Einstein equation give the value of the diffusion coefficient

$$D = \frac{kT}{6\pi\eta R_H}$$

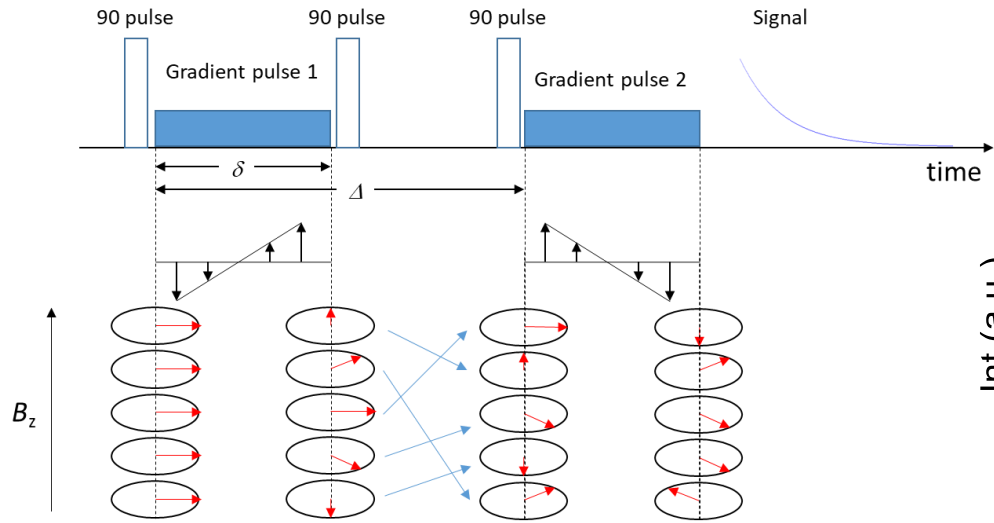
Can be determined by NMR with a pulse field gradient method



NMR method – pulse sequence

Sample: PMMA + MMA in CH₂Cl₂

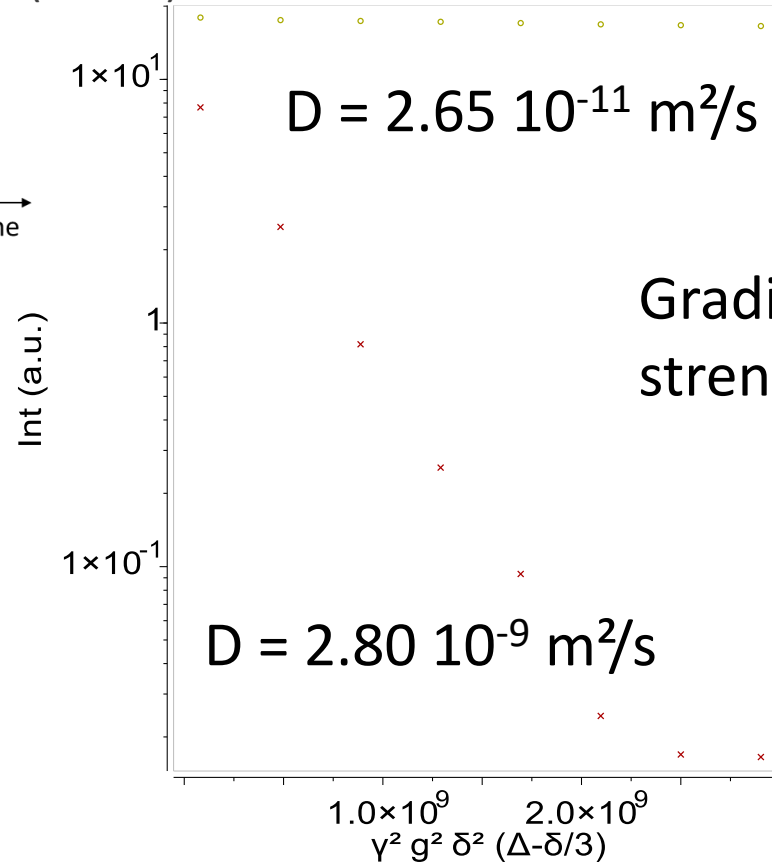
Pulse sequence: Pulsed Gradient Stimulated Echo (PGSTE)



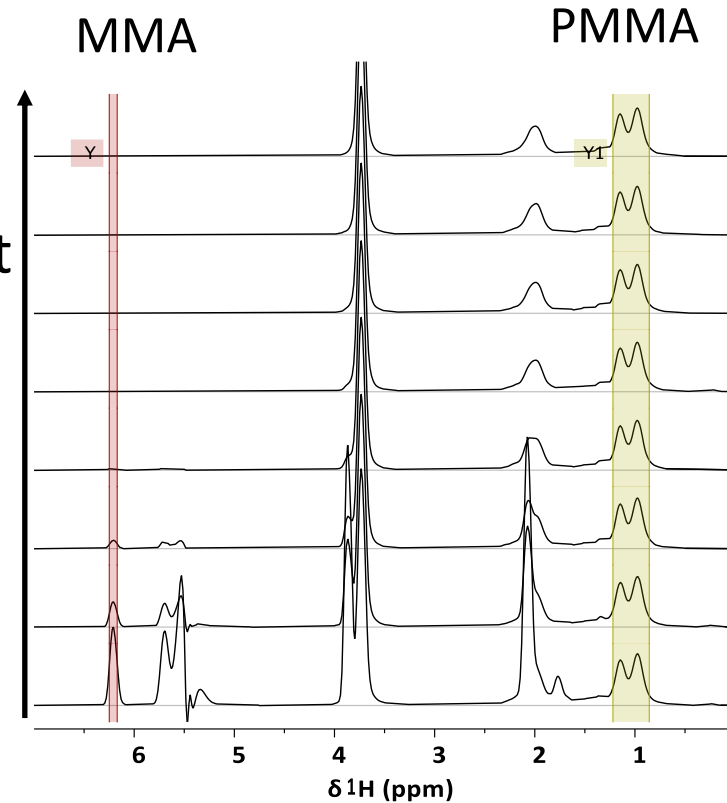
Stejskal and Tanner equation

$$\ln \left(\frac{S}{S_0} \right) = -D \cdot \gamma^2 \cdot g^2 \cdot \delta^2 \cdot \left(\Delta - \frac{\delta}{3} \right)$$

Stejskal, E.O., and Tanner, J.E., *Chem. Phys.* **42**, 288 (1965).



Gradient strength



How to connect D to MW?

Stokes-Einstein equation

$$D = \frac{kT}{6\pi\eta R_H}$$

D: the diffusion coefficient

K: the Boltzmann constant

T: the absolute temperature

η : the solvent viscosity

R_H : the hydrodynamic radius

Rouse-Zimm model

$$R_H \sim bM^\nu$$

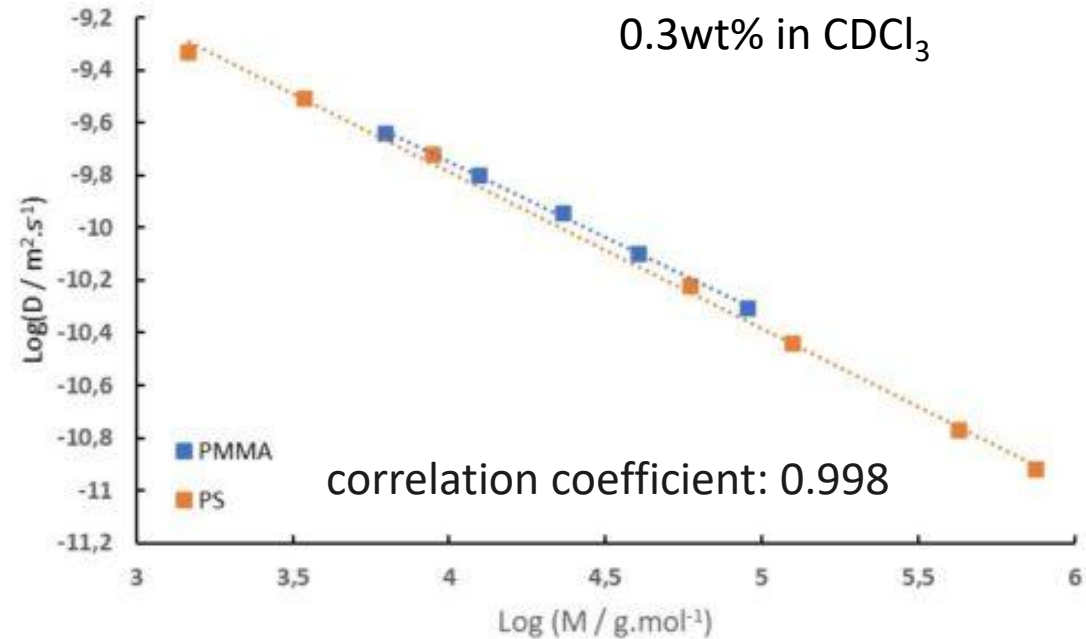
rewriting of the Stokes-Einstein equation (if T and η are constant)

$$D = b'M^{-\nu}$$

$$\log(D) = \log(b') - \nu \log(M)$$

MW Determination of PS and PMMA via PFG

PS		PMMA	
MW (g/mol)	D (m ² /s)	MW (g/mol)	D (m ² /s)
1470	4,64 10 ⁻¹⁰	6270	2,29 10 ⁻¹⁰
3460	3,12 10 ⁻¹⁰	12500	1,58 10 ⁻¹⁰
8900	1,89 10 ⁻¹⁰	23200	1,13 10 ⁻¹⁰
58900	6,02 10 ⁻¹¹	40300	7,96 10 ⁻¹¹
125000	3,63 10 ⁻¹¹	90600	4,92 10 ⁻¹¹
426000	1,69 10 ⁻¹¹		
753000	1,20 10 ⁻¹¹		



$\delta = 10$ ms; $\Delta = 300$ ms; G = 50%; meas. time < 10 min

- ✓ MW as high as 750 kg/mol
- ✓ D as low as 1.2 10⁻¹¹ m²/s
- ✓ Far from the limit of the benchtop NMR
- ✓ Fast, less than 10 min

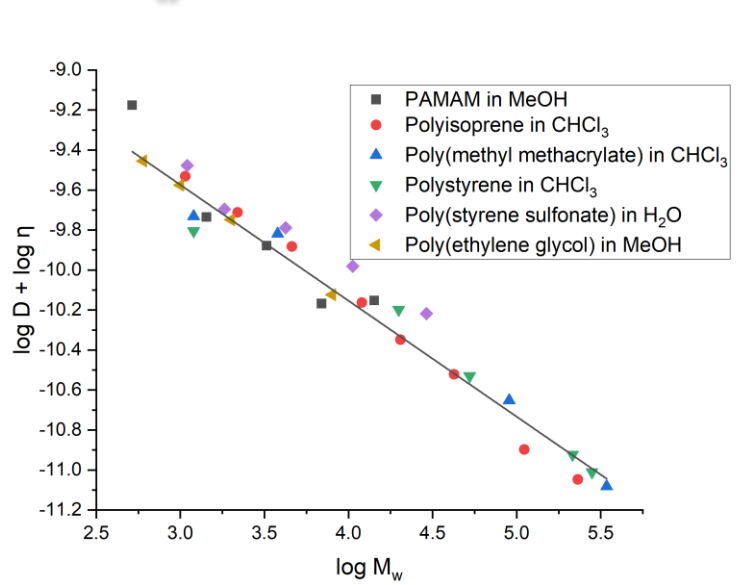
Fit the literature
(determined on high field
NMR) with the
same concentration



Online polymer mass detector through diffusion NMR

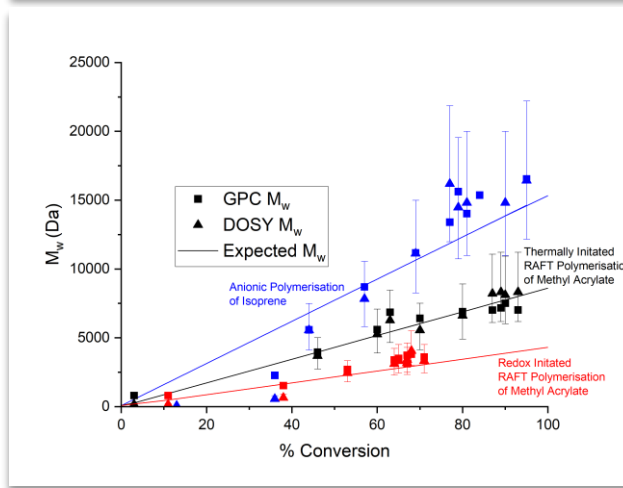
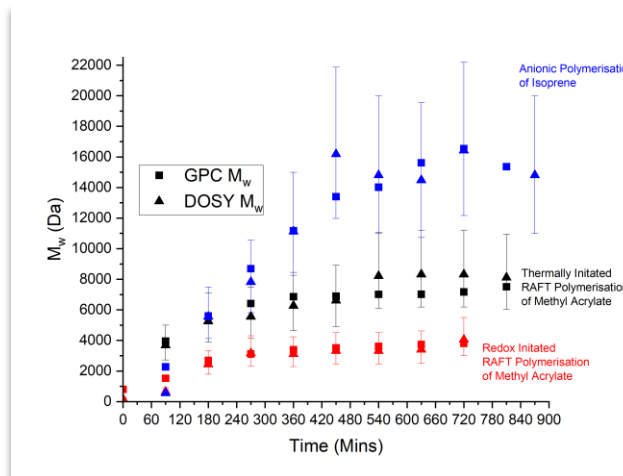
Calibrate

$$M_w = 10^{\frac{(\log D + \log \eta) + 7.74}{-0.597}}$$

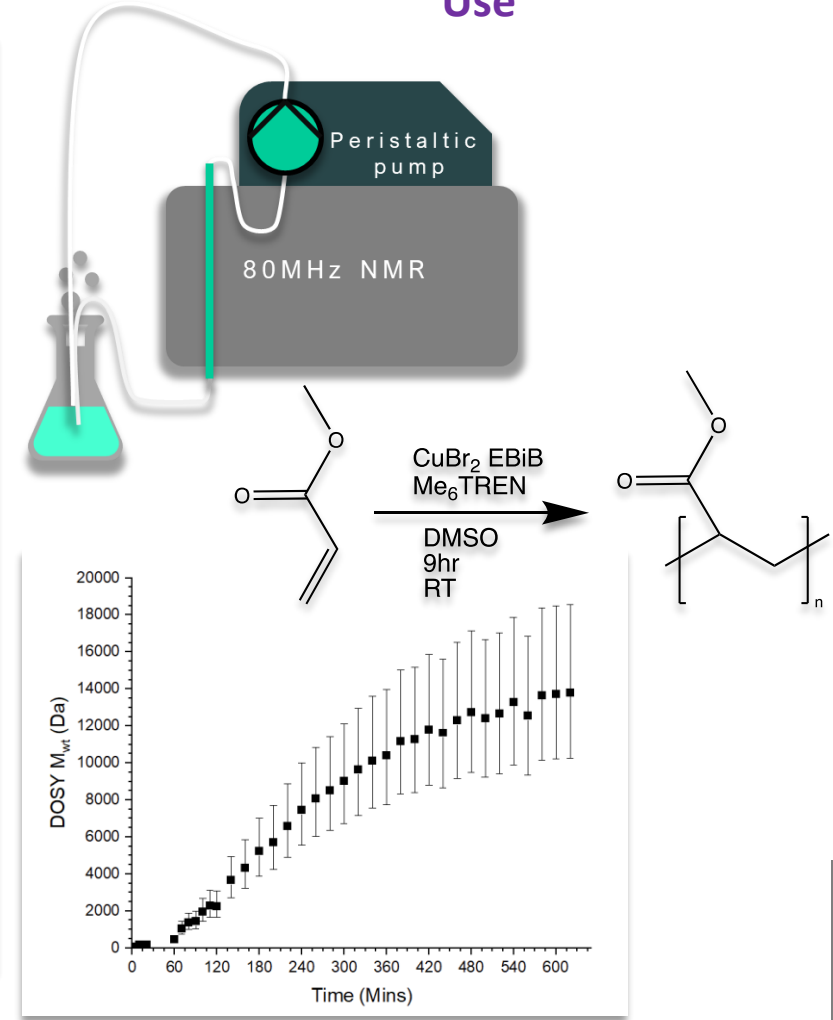


- Using a range of different polymer standards in different solvents and correcting for the solvent viscosity

Compare with SEC-GPC



Use



MaDDOSY (Mass Determination Diffusion Ordered Spectroscopy) using an 80 MHz Bench Top NMR for the Rapid Determination of Polymer, Macromolecular Molecular Weight; O. Tooley, W. Pointer, R. Radmall, M. Hall, V. Beyer, K. Stakem, T. Swift, J. Town, T. Junkers, P. Wilson, D. Lester, D. Haddleton, (2024) DOI: [10.1002/marc.202300692](https://doi.org/10.1002/marc.202300692)



D to M_{wt} calculator

Group Members ▶ Owen Tooley ▶ DOSY Molecular Weight Calculator

DOSY Molecular Weight Calculator

Calculate M_{wt} from DOSY

Please Consider Citing: <https://doi.org/10.1002/marc.202300692>

Diffusion Constant ($m^2 s^{-1}$):

Solvent Bulk Viscosity (mPa.s):

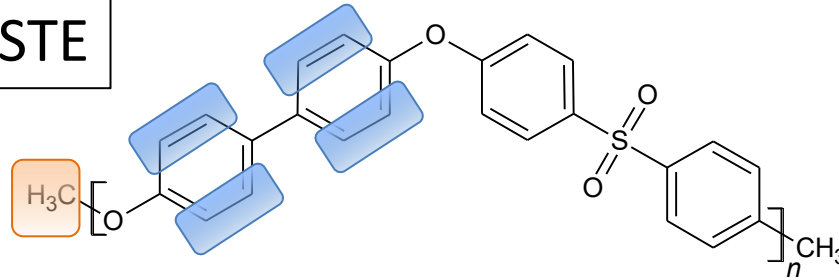
Number of Scans:

Calculate

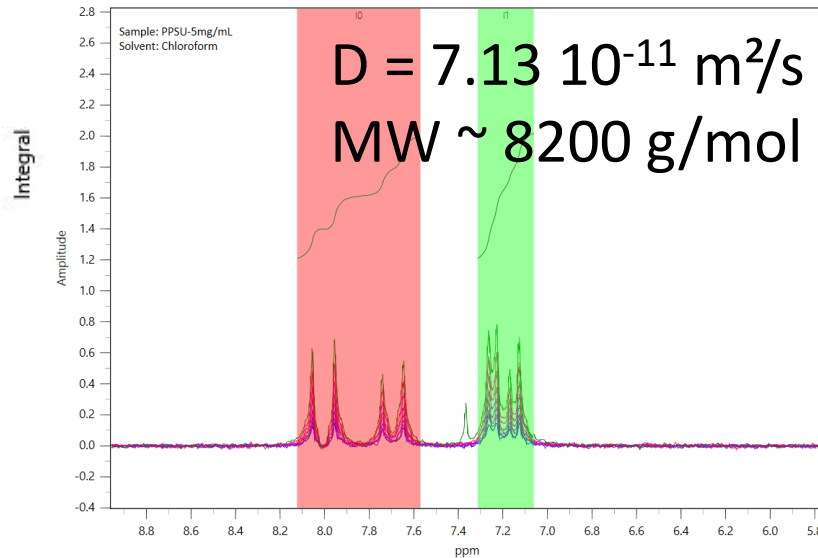
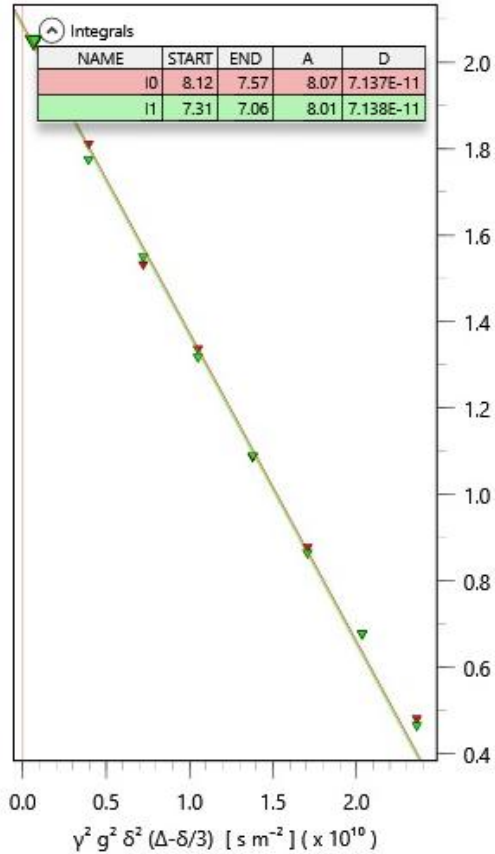


PPSU MW - end-group analysis vs PFG

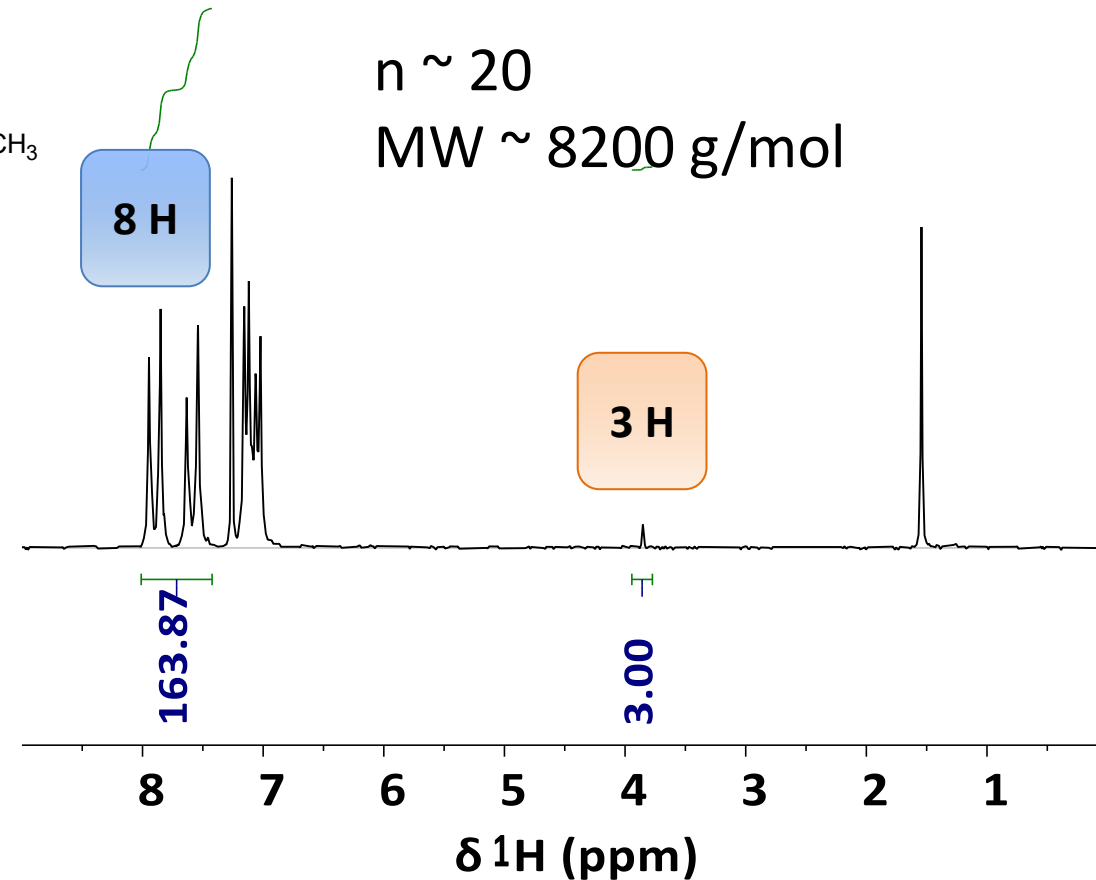
PGSTE



1D 1H - End-group analysis



$n \sim 20$
 $MW \sim 8200 \text{ g/mol}$



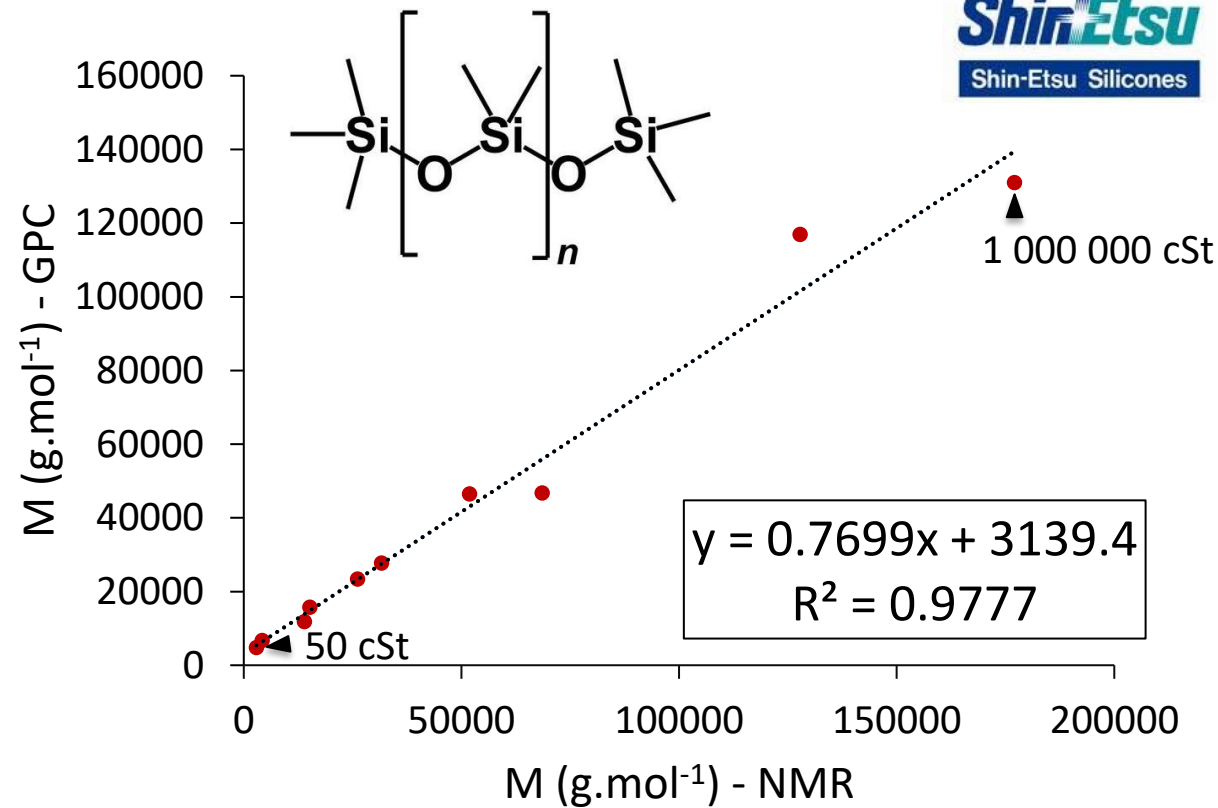
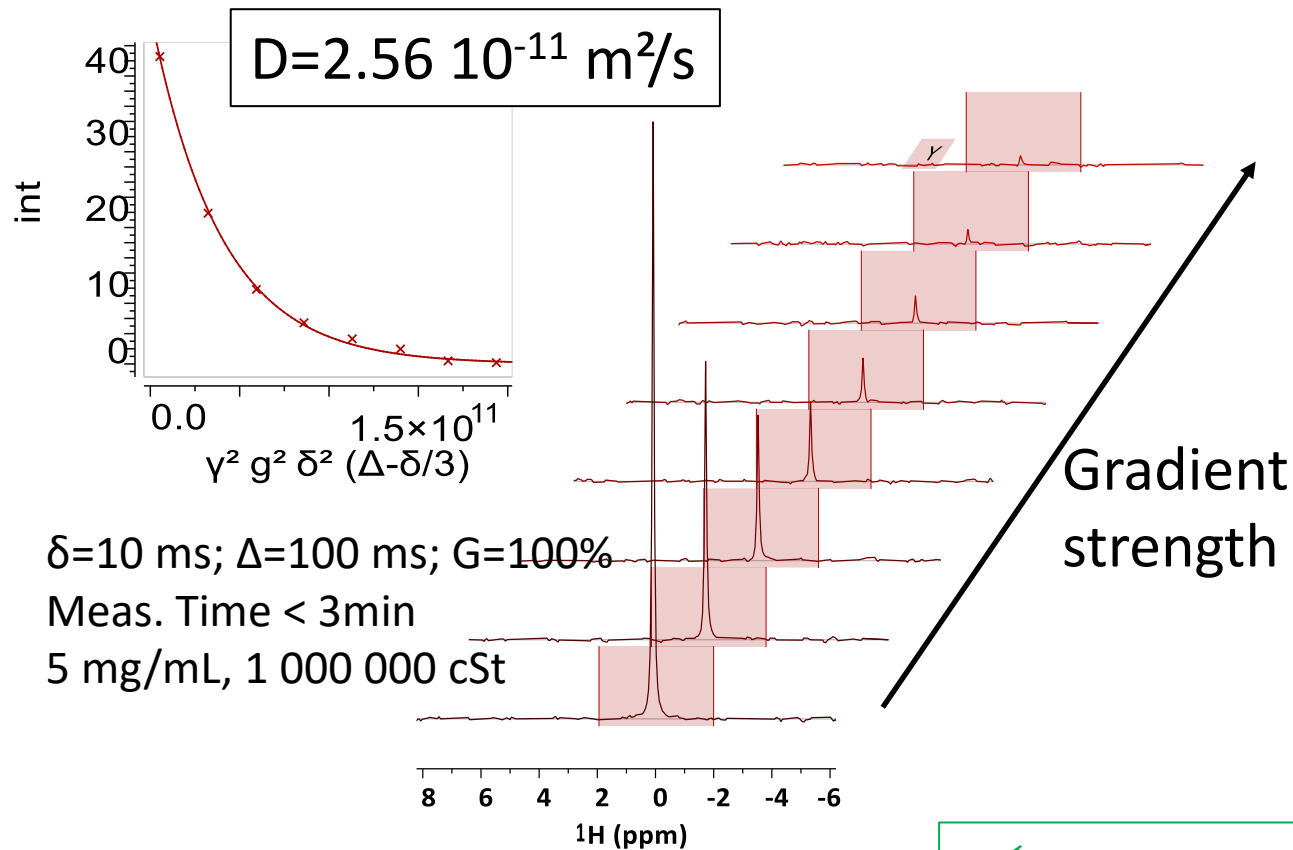
$\delta=5 \text{ ms}$; $\Delta=30 \text{ ms}$; $G=75\%$

✓ Both methods give the same MW
✓ Far from the limit of the benchtop NMR

<https://magritek.com/wp-content/uploads/2025/01/App-Note-Polymer-MW-Calculation.pdf>



PDMS MW - GPC vs PFG analysis

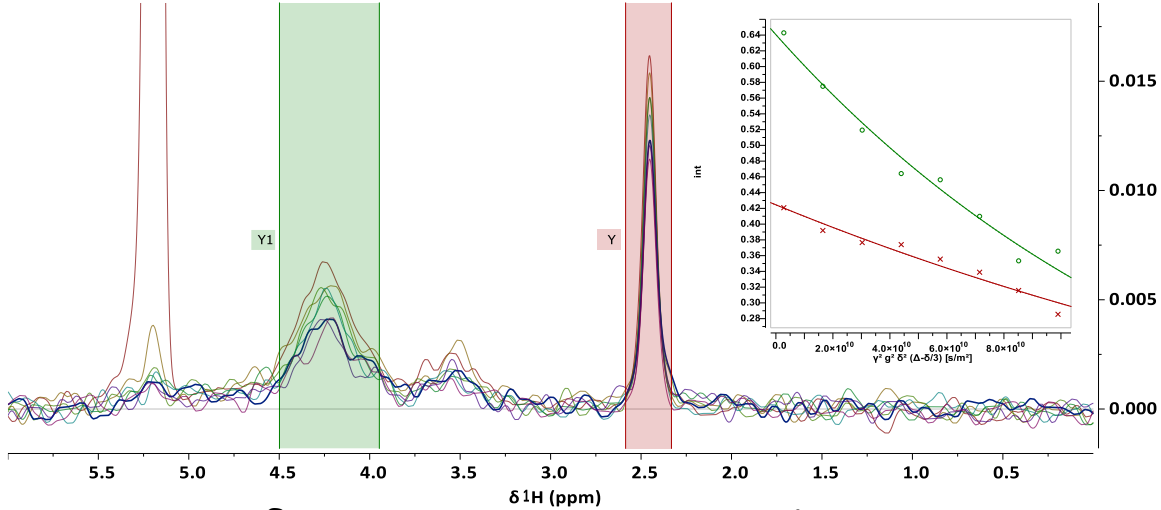


- ✓ Good correlation between GPC and NMR values
- ✓ Fast, 3 min measurement time
- ✓ Sustainable alternative, only 0.5 mL of Solvent

https://magritek.com/wp-content/uploads/2025/12/AppNote_Mw-determination-of-PDMS-by-diffusion_final-1.pdf

Natural polymer: chitosan

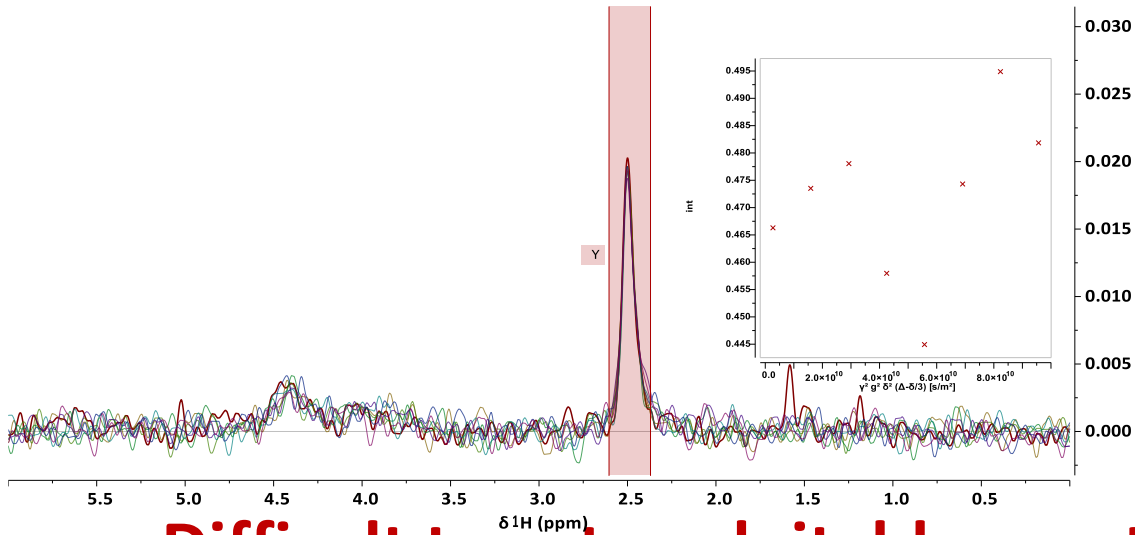
#scans = 128, RT = 7 s, Meas time = 119.71 min, $\delta = 7$ ms, $\Delta = 100$ ms, max gradient = 100 %



128 scans; $\delta=7$ ms; $\Delta=100$ ms; G=100%

MW = 105 kg/mol; C = 10 mg/mL in D₂O with 2% formic acid

$$D = 3.50 \cdot 10^{-12} \text{ m}^2/\text{s}$$



Higher MW of chitosan solution!

Difficult to get exploitable spectra to extract the diffusion coefficient

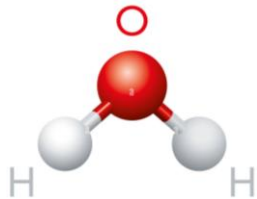
Advantages of higher T measurements:

- Solvent is less viscous

Bulk viscosity of H₂O

25 °C	0.89 mPa s
60 °C	0.47 mPa s

- Brownian motion of a (macro)molecule in solution increase



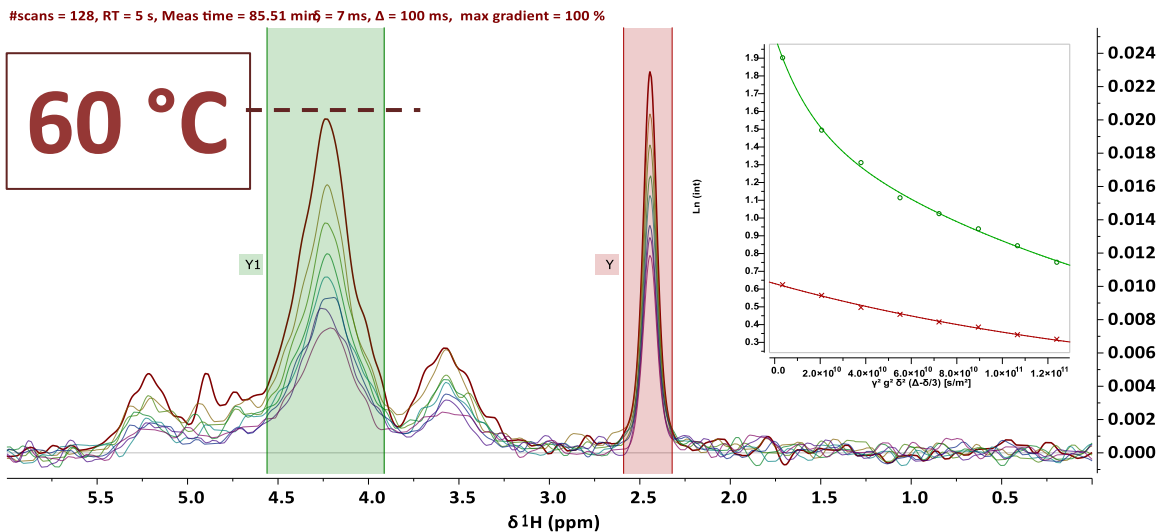
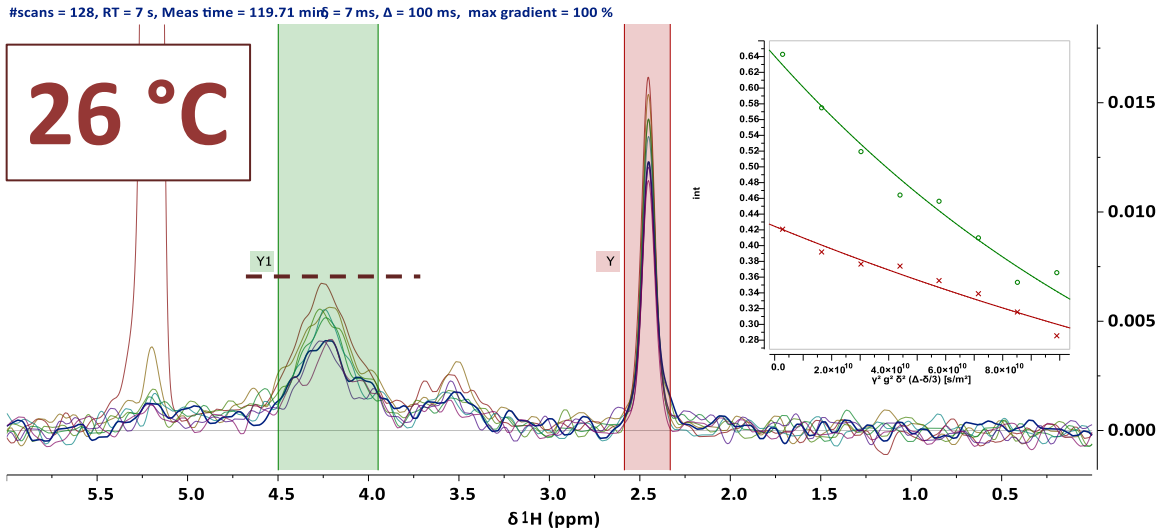
Diffusion coef. of water

25 °C	$2.30 \cdot 10^{-9} \text{ m}^2/\text{s}$
60 °C	$4.70 \cdot 10^{-9} \text{ m}^2/\text{s}$

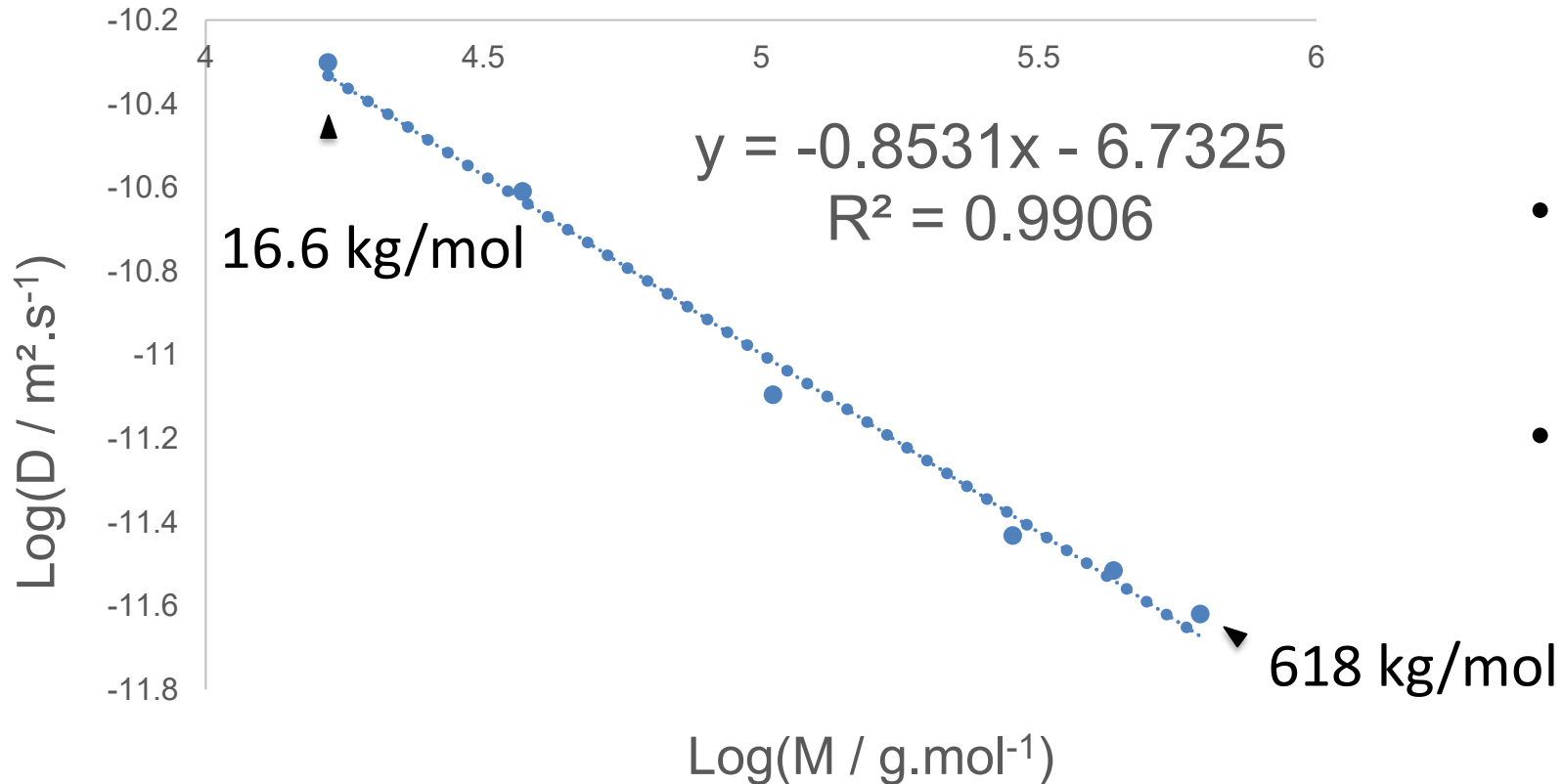
- Higher value of T_2 -> less signal loss due to T_2 effect

PFG of chitosan: 26 vs 60 °C

MW = 105 kg/mol; C = 10 mg/mL in D₂O with 2% formic acid



✓ Gained in signal intensity
✓ Difference by a factor of 2 in the diffusion coefficient

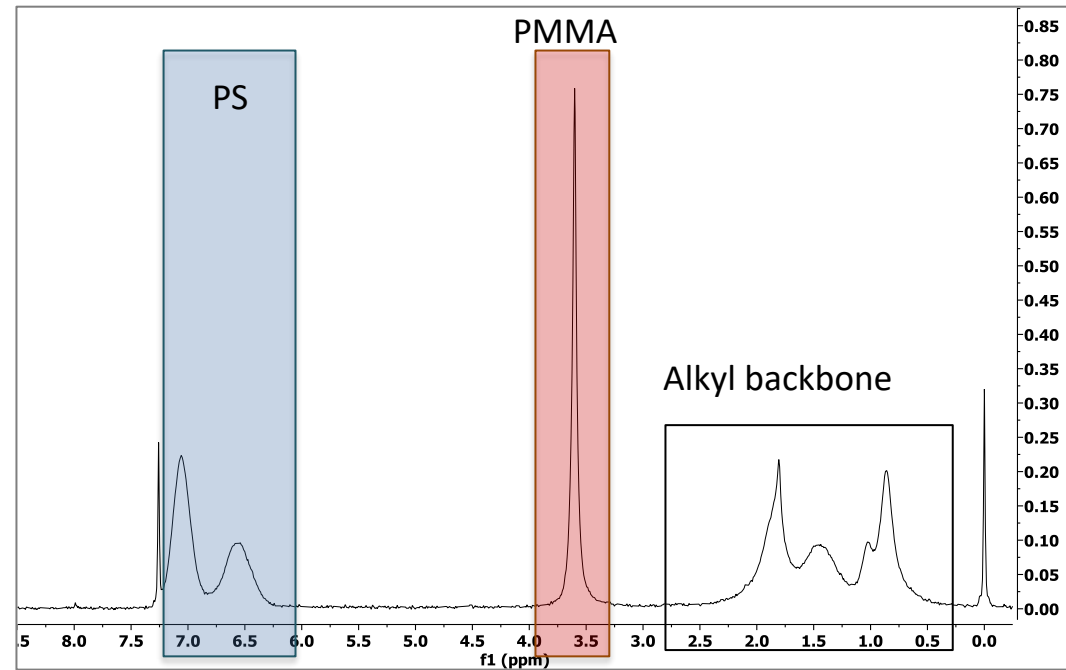
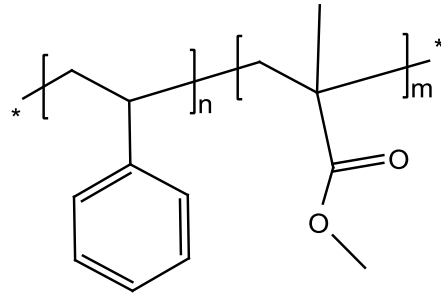


- Diffusion coefficient measured at 60 °C
- Average of 3 measurements

✓ Possibility to build a calibration curve with chitosan standard



Quality of the copolymer/copolymerization

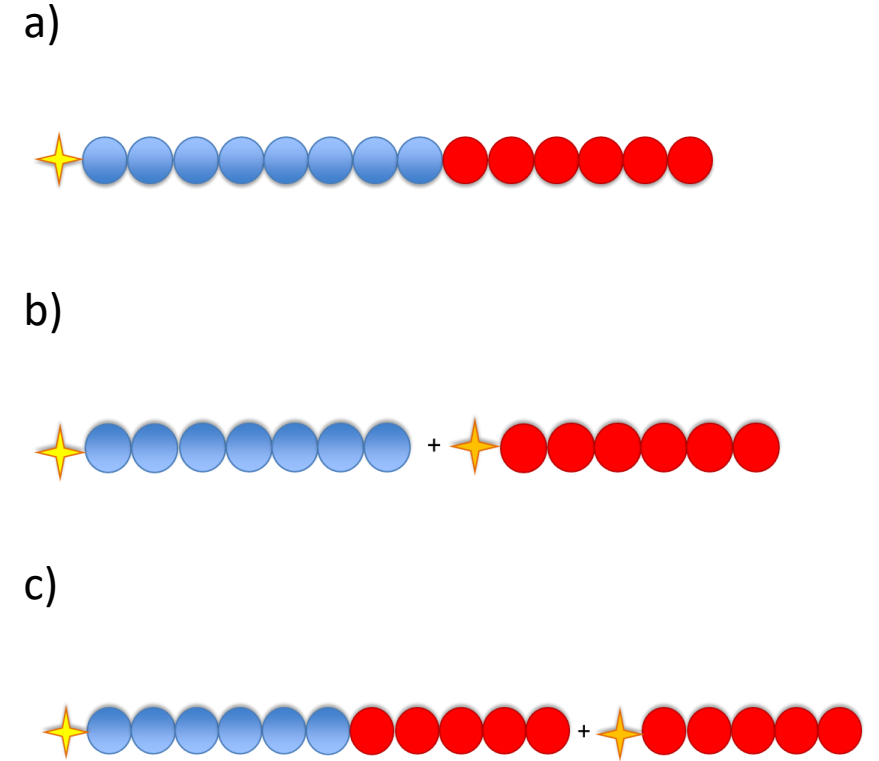
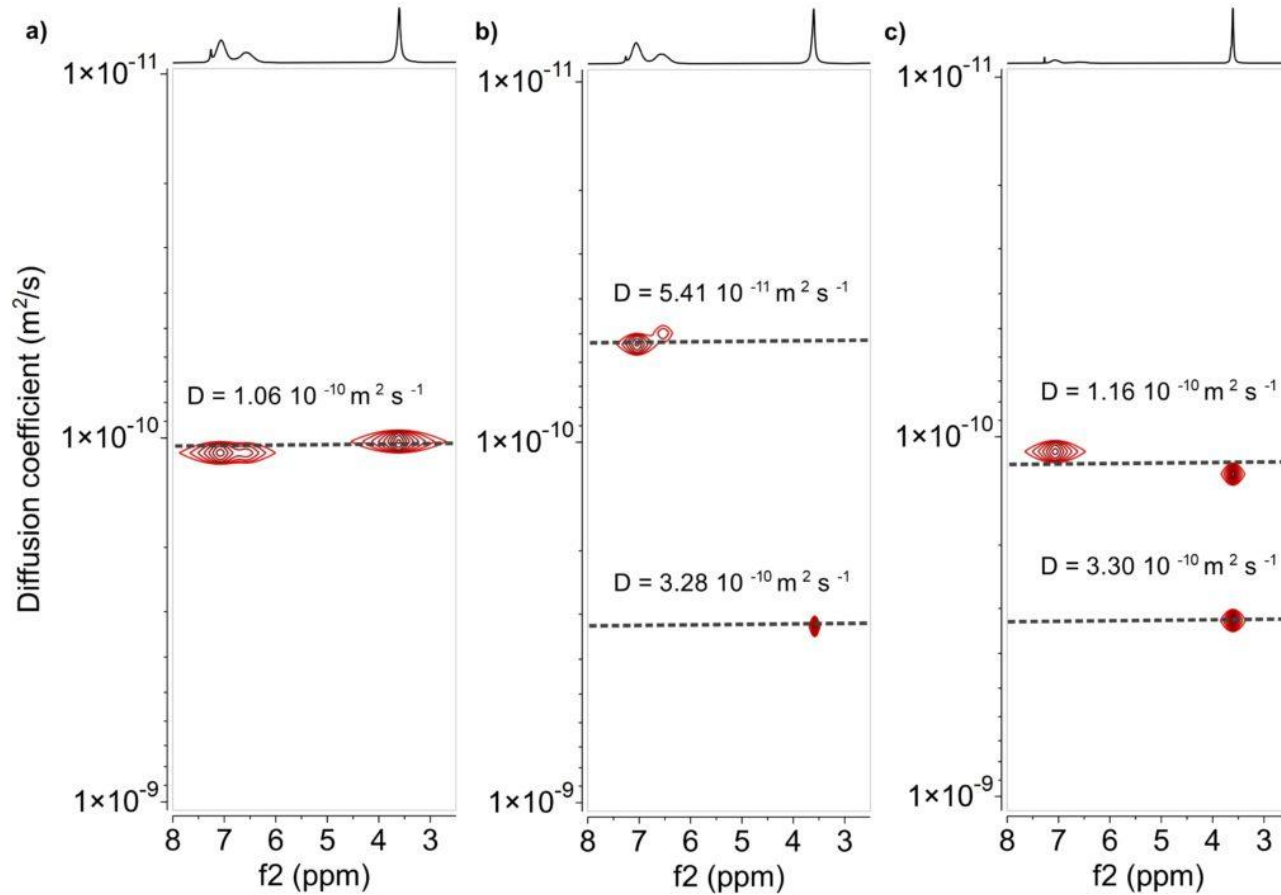


What do you have in your solution that 1D 1H does not reveal but DOSY does?

- Only PS-*b*-PMMA Block copolymer?
- PS homopolymer and PMMA homopolymer?
- PS-*b*-PMMA Block copolymer and PMMA homopolymer?
- ...



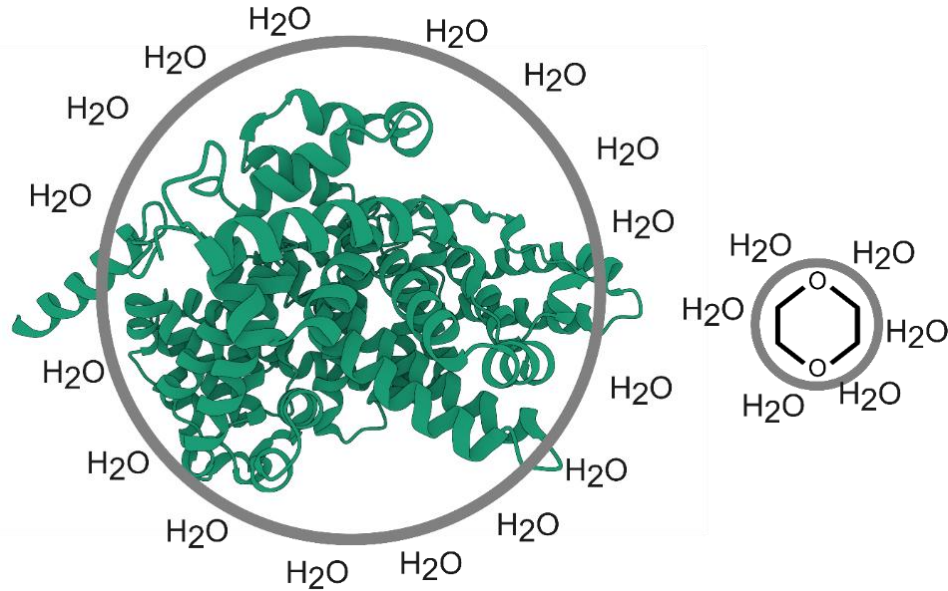
Quality check by diffusion measurements





Protein Radius of Hydration by Diffusion NMR

$$\ln(I/I_0) = \gamma^2 \delta^2 g^2 [\Delta - \delta/3] D. [1]$$



$$D = \frac{k_B T}{6\pi\eta R_H}$$

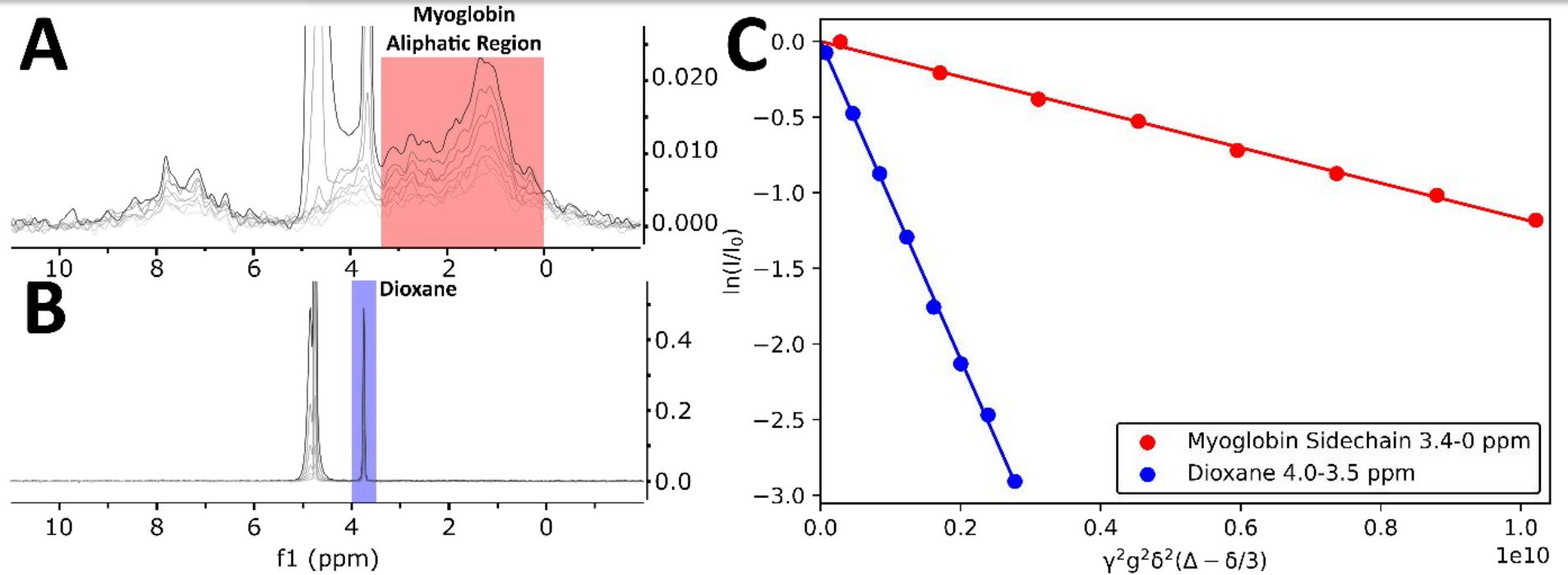
$$D_{standard} R_{H standard} = \frac{k_B T}{6\pi\eta} = D_{unknown} R_{H unknown}$$

$$R_H = R_{H dioxane} \frac{D_{dioxane}}{D_{sample}}$$

Internal diffusion standard (dioxane) and protein -> same bulk solution properties



Protein Radius of Hydration by Diffusion NMR



Protein	Dioxane Diffusion	Protein Diffusion	R_H^*	Literature R_H
Bovine Serum Albumin	$9.86 \times 10^{-10} \text{m}^2/\text{s}$	$6.12 \times 10^{-11} \text{m}^2/\text{s}$	$35 \pm 1 \text{ \AA}$	37.4 \AA^5
Equine Heart Myoglobin	$1.05 \times 10^{-9} \text{m}^2/\text{s}$	$1.17 \times 10^{-10} \text{m}^2/\text{s}$	$19.4 \pm 0.2 \text{ \AA}$	20.4 \AA^8

Benchtop NMR applications



Synthesis

Pharma

Macromolecules

Education

Flow chemistry



Batteries

Food & Agriculture

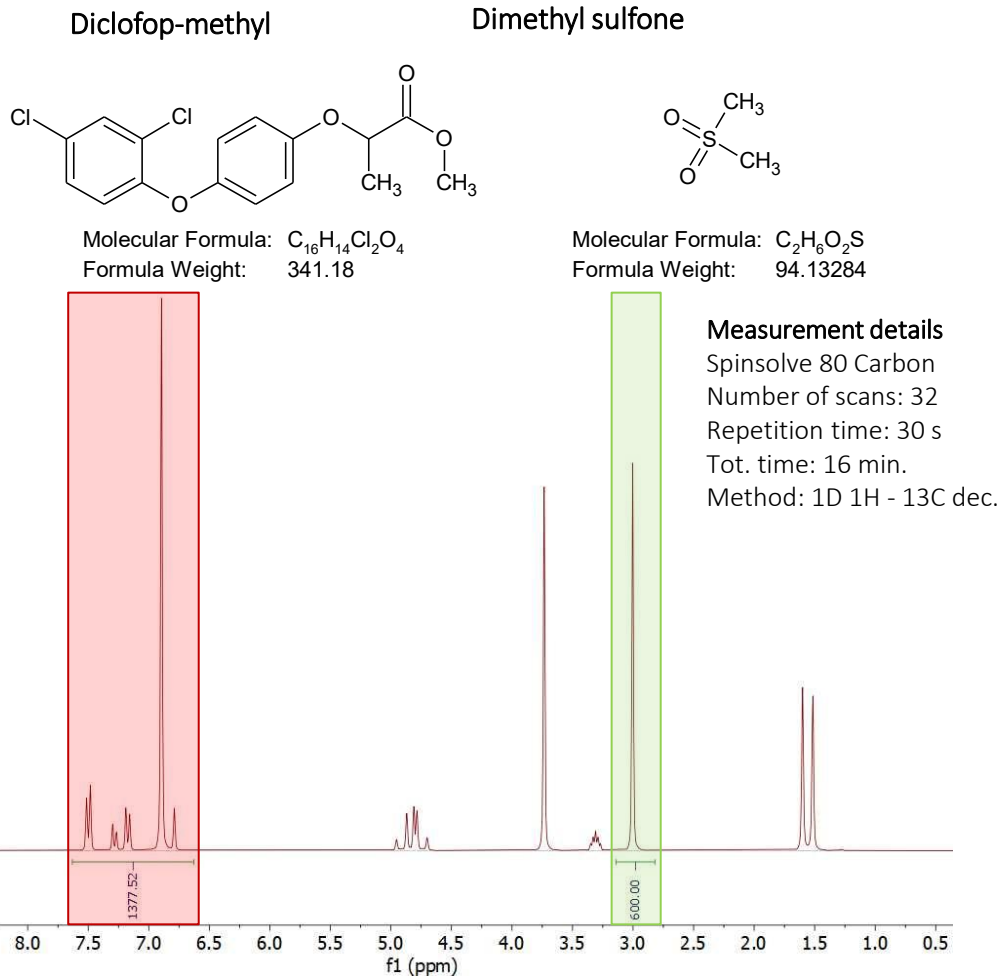
Forensics

Analytics



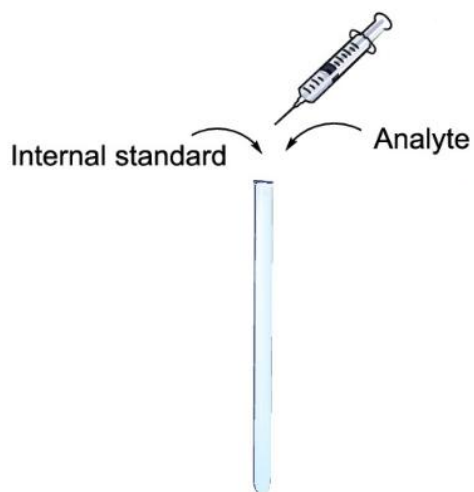
Absolute quantification (assay) with internal standard

- NMR is typically used to measure product assay with the use of an internal standard



HPC
HPC Standards GmbH

The HPC Standards GmbH is a Manufacturer and distributor of high-purity analytical standards for residue analysis.



- Access to the assay (not purity) of the compound

Sample preparation details

Diclofop-methyl:	34.9 mg
Internal std (Dimethyl sulfone):	4.9 mg
Solvent (MeOH-d4):	1 mL
Sample volume:	500 µL

Concentrations:

Diclofop-methyl	100 mM
DMS	50 mM

$$Assay_{Prod} [\%] = \frac{Assay_{Std} W_{Std} M_{Prod}}{100 M_{Std} W_{Prod}} \frac{Int_{Prod} N_{Std}}{Int_{Std} N_{Prod}}$$

$Assay_{Prod} [\%] = 99.7 \%$

<https://magritek.com/2020/12/04/quantitative-nmr-with-internal-standard-on-a-spinsolve-benchtop-nmr-spectrometer/>



Microplastics quantification

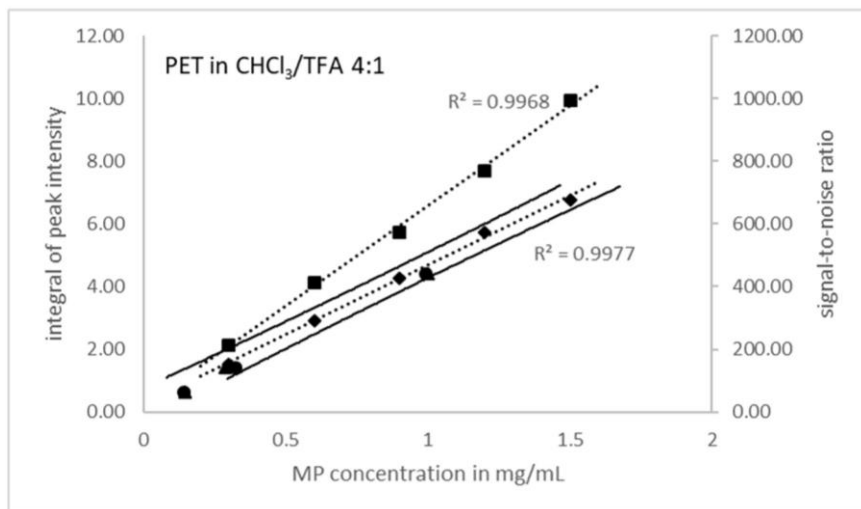


RESEARCH ARTICLE | [Open Access](#) |

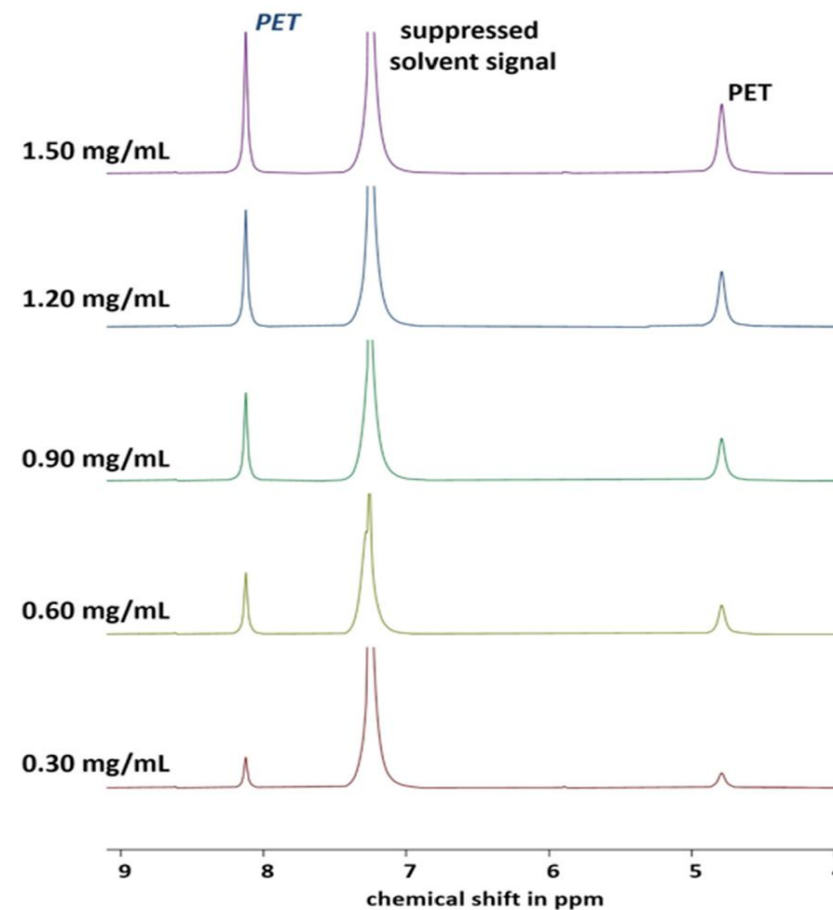
Applicable and cost-efficient microplastic analysis by quantitative ^1H -NMR spectroscopy using benchtop NMR and NoD methods

Nadine Peez, Torsten Rinesch, Jürgen Kolz, Wolfgang Imhof

First published: 20 August 2021 | <https://doi.org/10.1002/mrc.5210>



• External qNMR method



^1H -NMR stack plot of 5 different concentrations (0.25–1.5 mg/ml)



Microplastics quantification


LOD

PET	4 µg/ml (29.87 min)
PVC	19 µg/ml (136.52 min)
PS	21 µg/ml (34.13 min)
LOQ	
PET	12 µg/ml (29.87 min)
PVC	57 µg/ml (136.52 min)
PS	64 µg/ml (34.13 min)

- Routine analysis of environmental samples
- No cleaning step between samples
- Competitive against thermoanalytical methods like Thermal Extraction - Desorption Gas Chromatography Mass Spectrometry (TED-GC-MS), pyrolysis-GCMS, and Thermogravimetry Gas Chromatography Mass Spectrometry (TGA-GC-MS)) and even
- Faster than spectroscopic methods (FTIR and Raman), which take up to 1 day

Solvent traces quantification



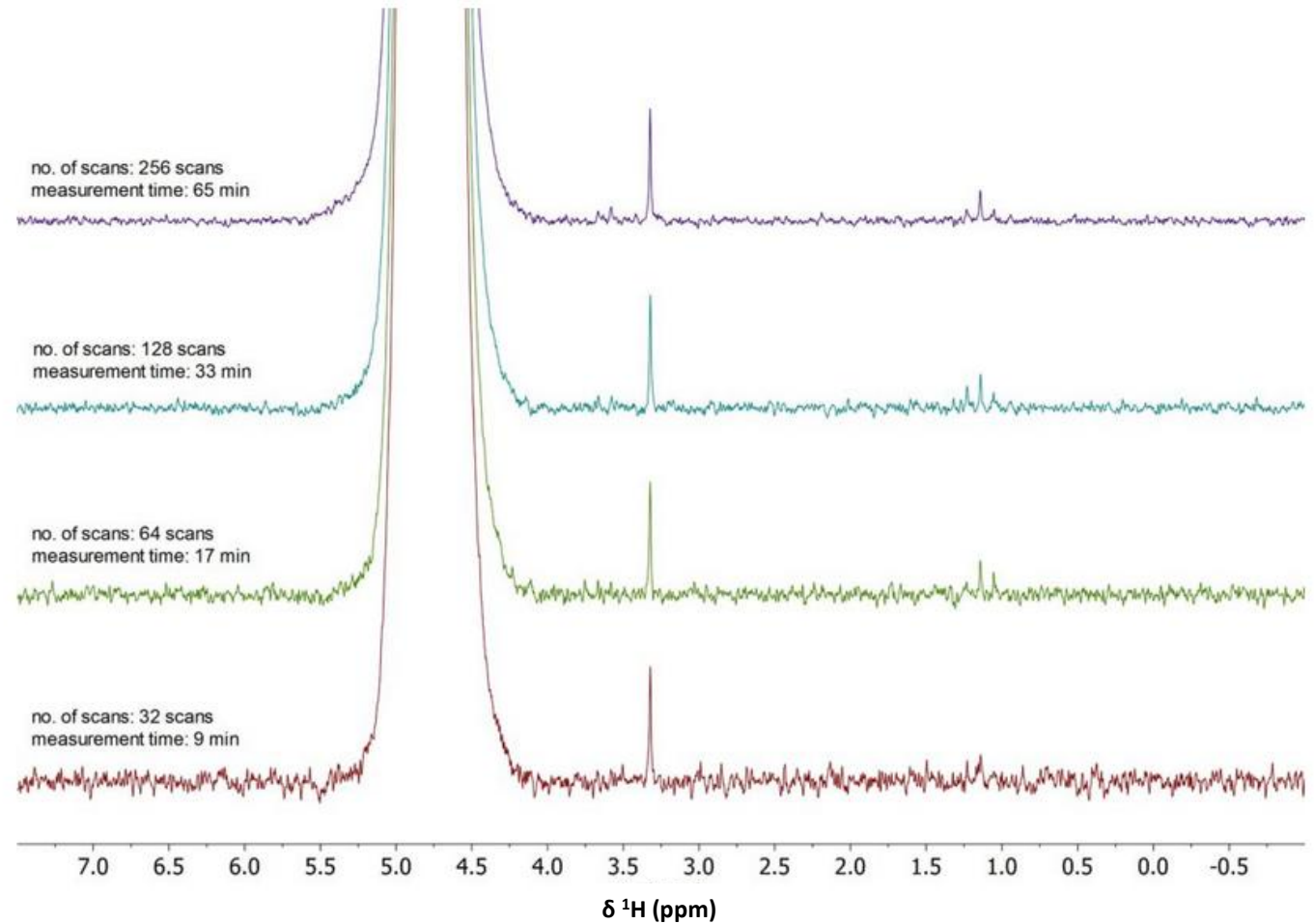
 1D ^1H (PRESAT)

 10 ppm MeOH/EtOH in H_2O

 Spinsolve 80 ULTRA

$$c(\text{LOD, MeOH}) = \frac{3}{4.29749} = 0.70 \text{ ppm} = 22 \mu\text{M}$$

$$c(\text{LOD, EtOH}) = \frac{3}{1.32488} = 2.26 \text{ ppm} = 50 \mu\text{M}$$



Residual solvent quantification in API sample



Quantitative T2-filter NMR protocol for residual solvent trace quantification (in the ppm order of magnitude)

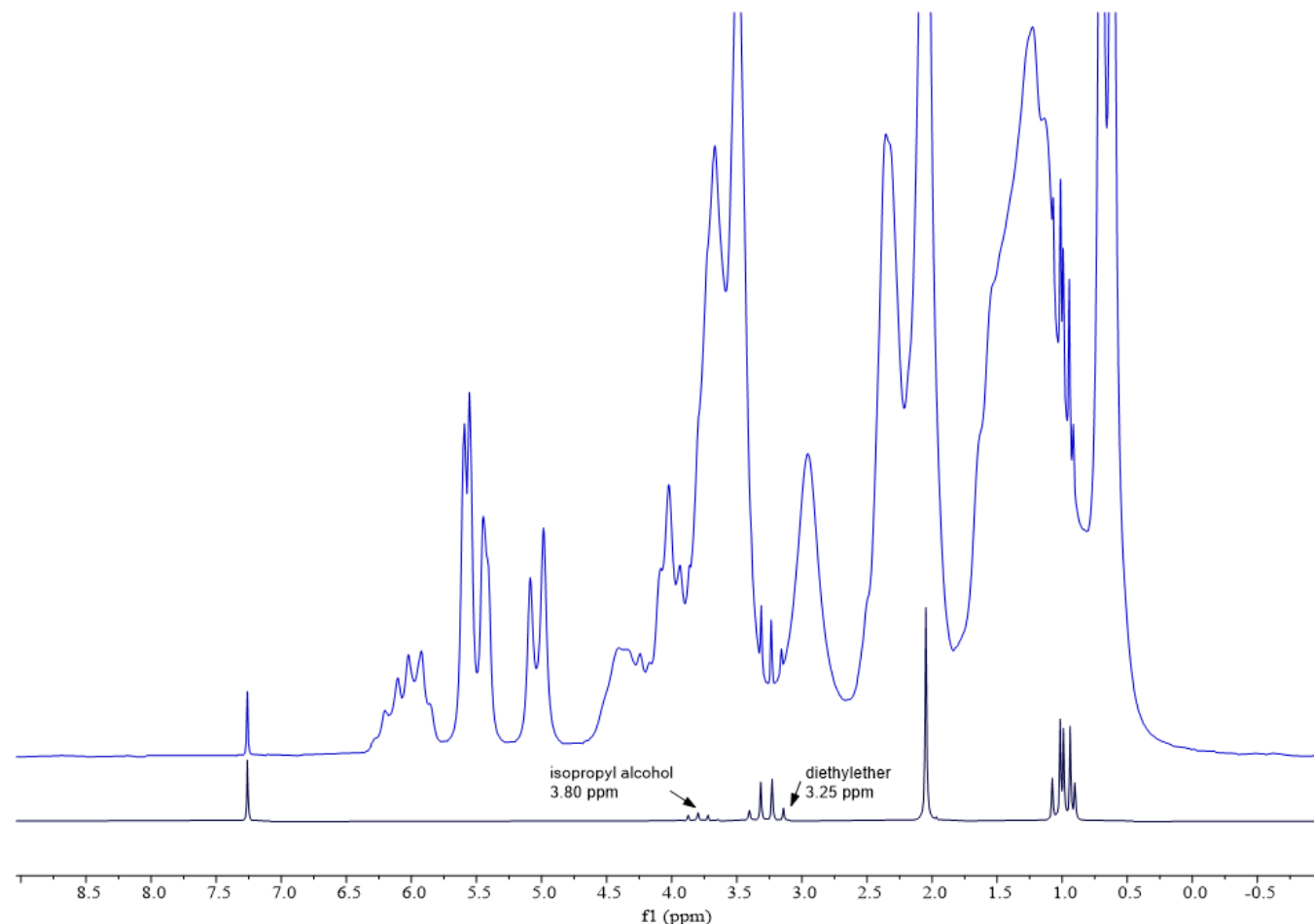
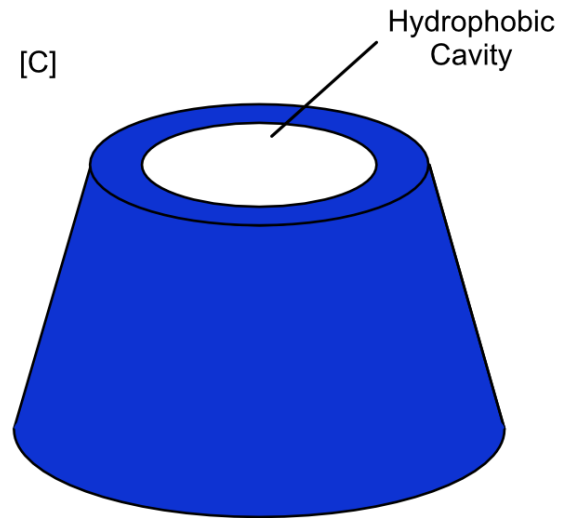
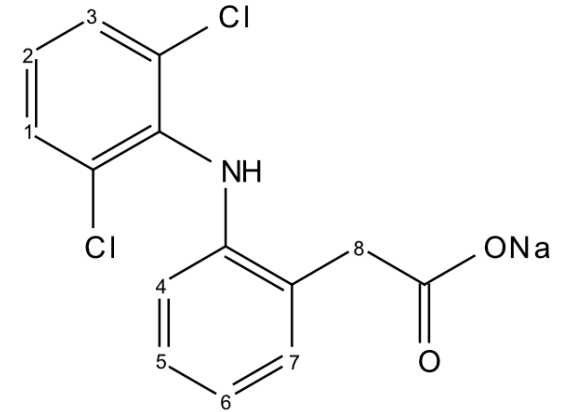


Figure 1: Stacked spectra of the API scaffold without (blue) and with (black) the relaxation filter applied before the NMR protocol.

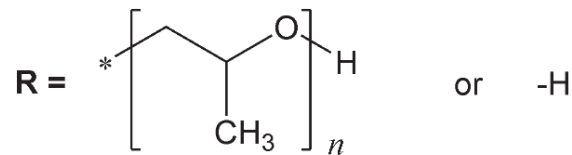
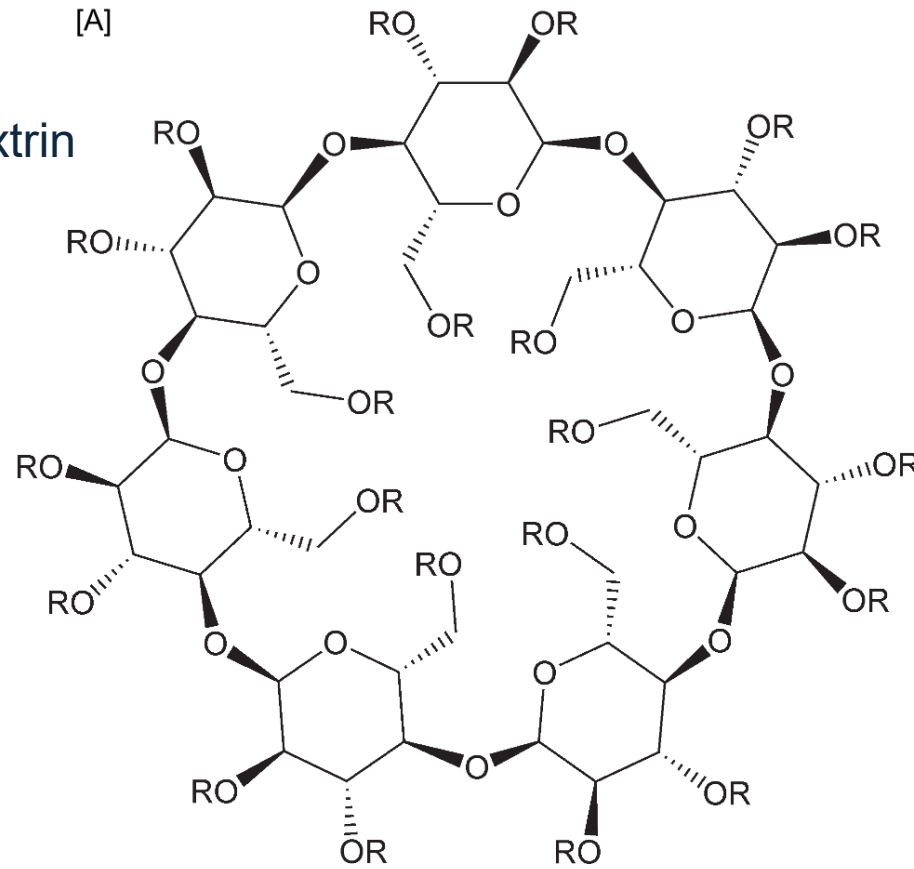
Cyclodextrin Drug Inclusion



sodium diclofenac



2-hydroxypropyl-β-cyclodextrin



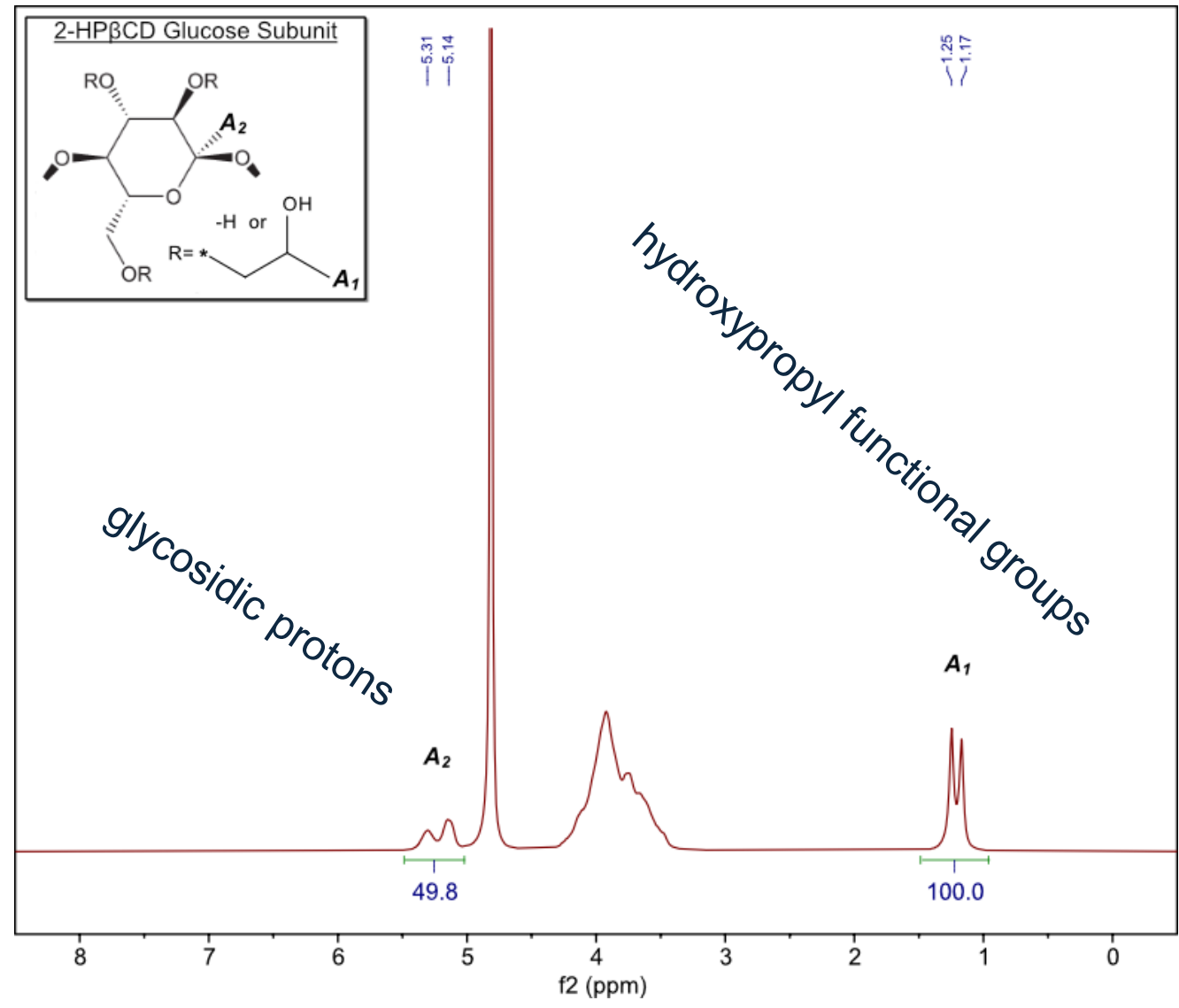
Challenge: rapid and reliable confirmation of inclusion complex formation and geometry during early-stage screening.

Molar Substitution Determination of 2-HP-B-CD



$$MS = \frac{A_1}{3 \times A_2}$$

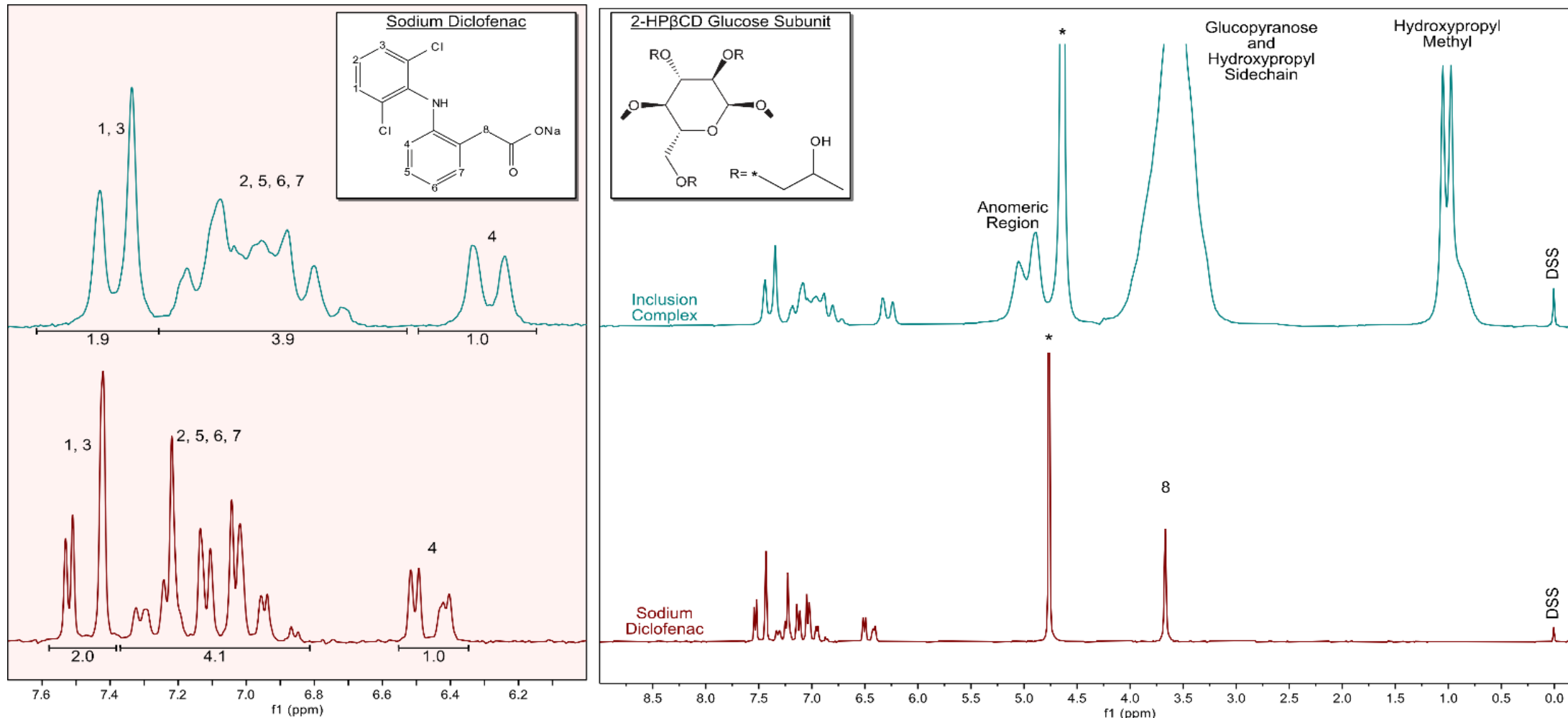
MS = 0.67
Labeling: MS=0.7



1D ¹H Spectroscopy as an Inclusion Screening Tool

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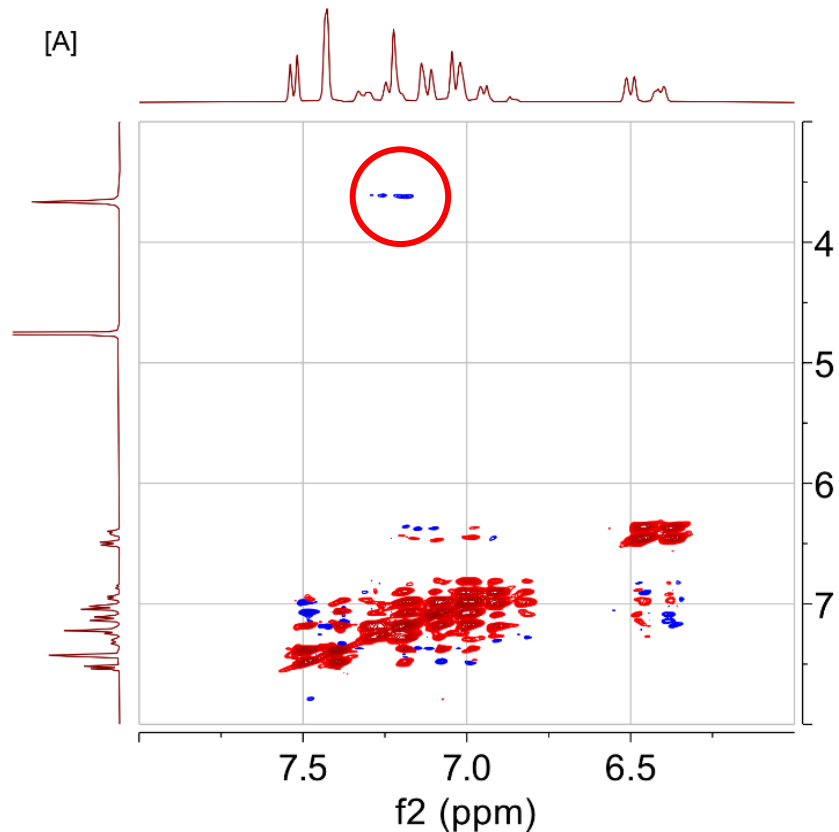
75 mg of sodium diclofenac was added to 1 mL of D₂O, heated to 60 °C, and stirred while 160 mg of 2-HPβCD in 2 mL of D₂O was added dropwise over 5 minutes



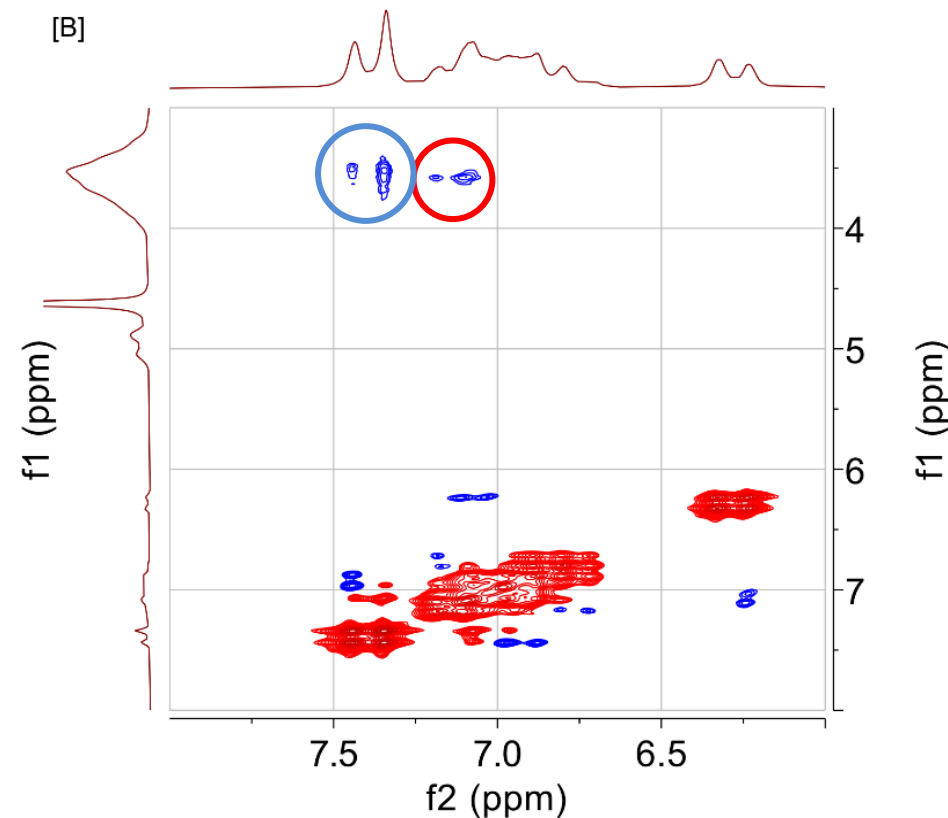
Through-space 2D NMR technique : ROESY



Sodium Diclofenac



Cyclodextrin Drug Inclusion



Red circle: intramolecular coupling between the methylene protons (3.7 ppm) and aromatic protons (7.3 ppm)

Blue circle: coupling between the 2-HPβCD cavity protons (3.6 ppm) and the sodium diclofenac H-1 and H-3 protons (7.4 ppm)

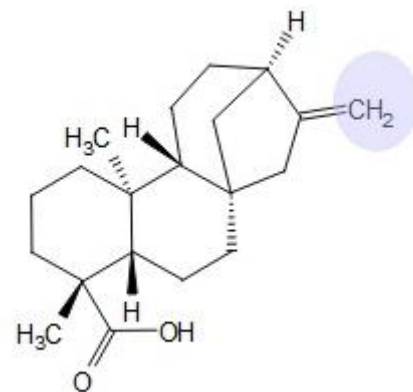
- ✓ Rapidly assessing drug product inclusion complexes
- ✓ Further work to quantify binding interactions (e.g., via NMR titration)
- ✓ Directly in the formulation laboratory, on the bench



Quantitation by ^1H , ^{13}C HSQC of Kaurenoic Acid in Copaiba Extracts

 1D ^1H

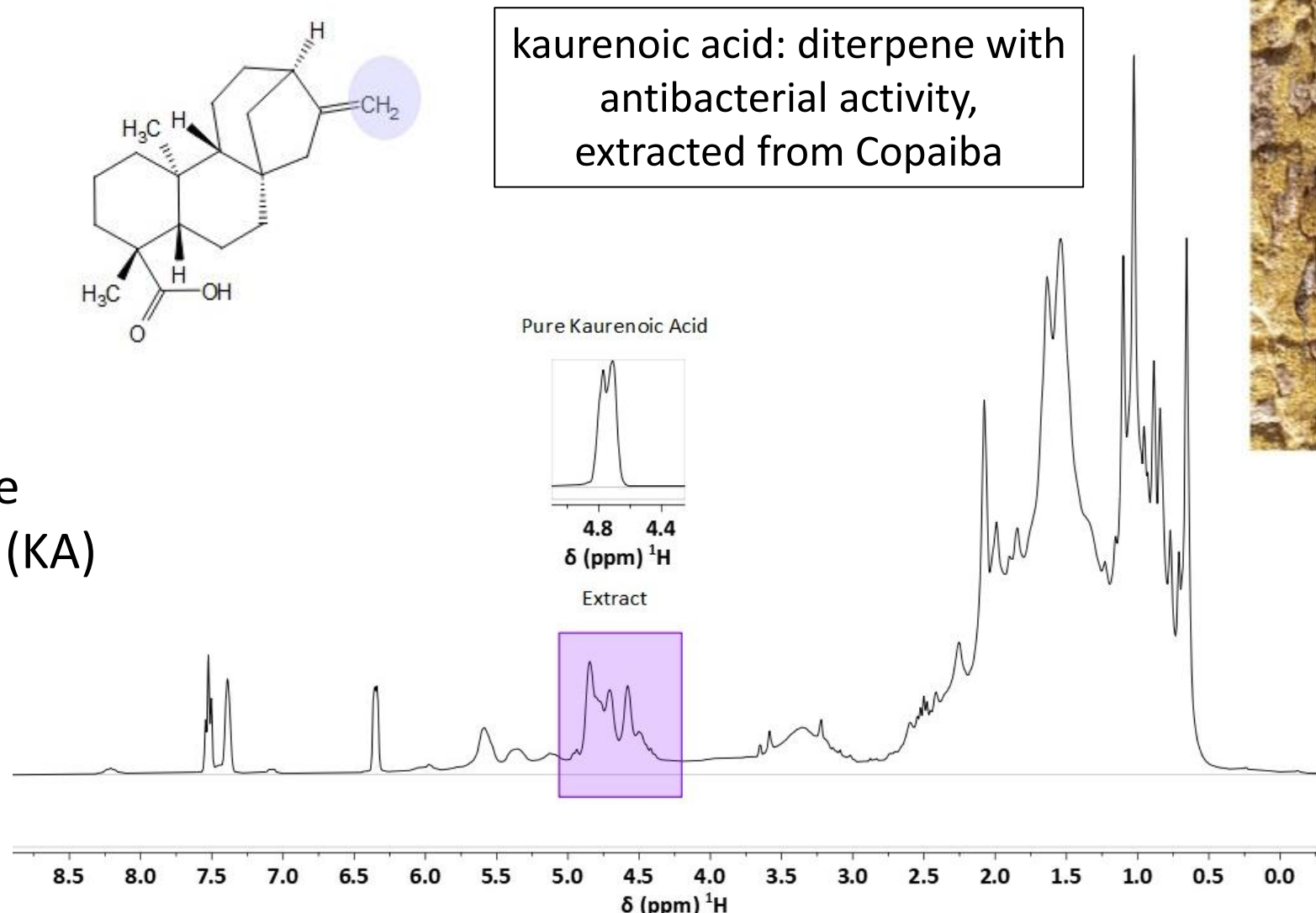
 Spinsolve 80 ULTRA



kaurenoic acid: diterpene with antibacterial activity, extracted from Copaiba



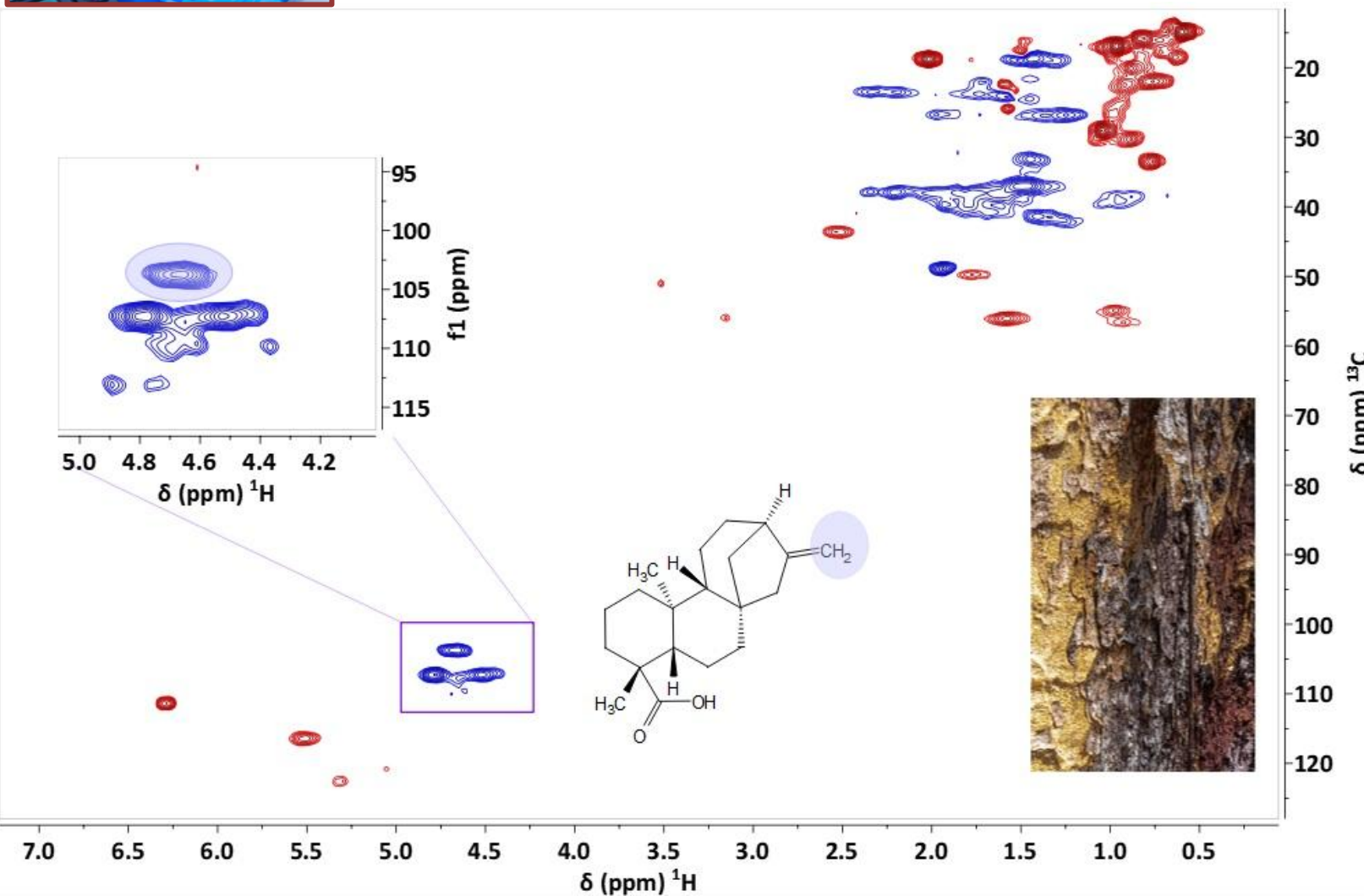
Goal: determination of the amount of kaurenoic acid (KA) in the Copaiba extract (define the quality)



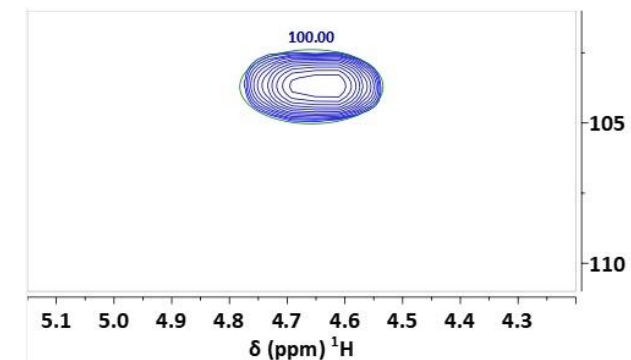


Quantitation by ^1H , ^{13}C HSQC of Kaurenoic Acid in Copaiba Extracts

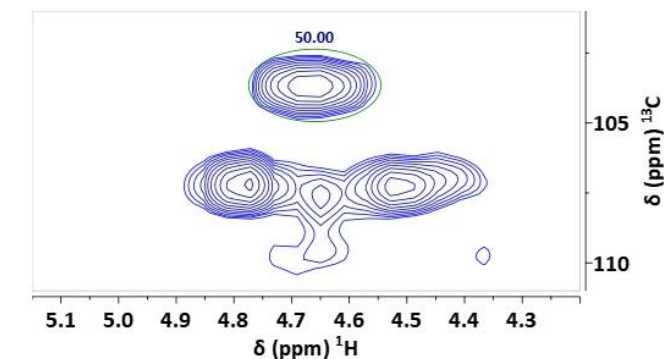
2D ^1H , ^{13}C HSQC-ME  Spinsolve 80 ULTRA



KA External standard



Extract

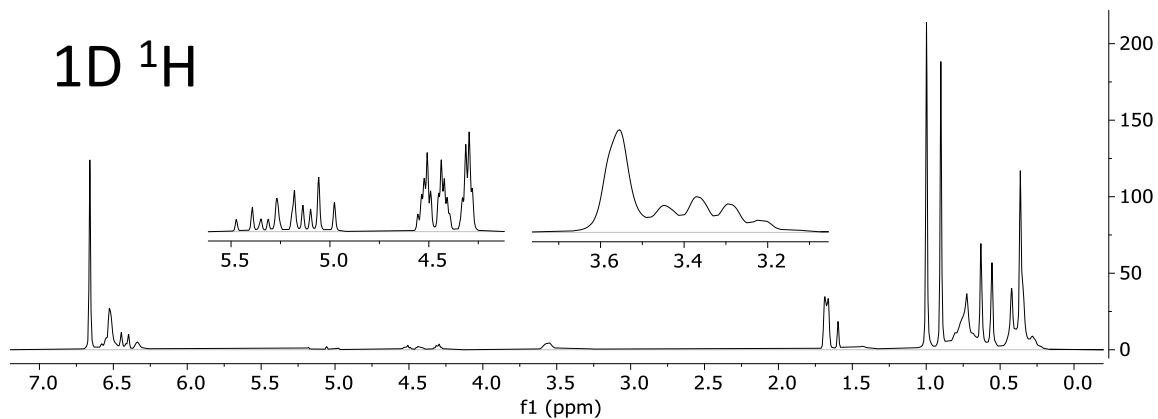
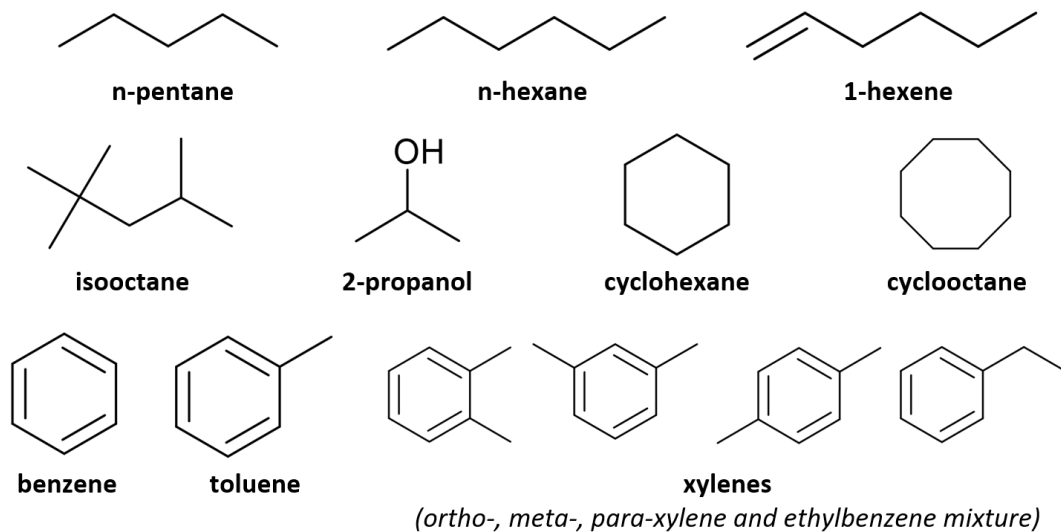


This sample contains 9.2% KA

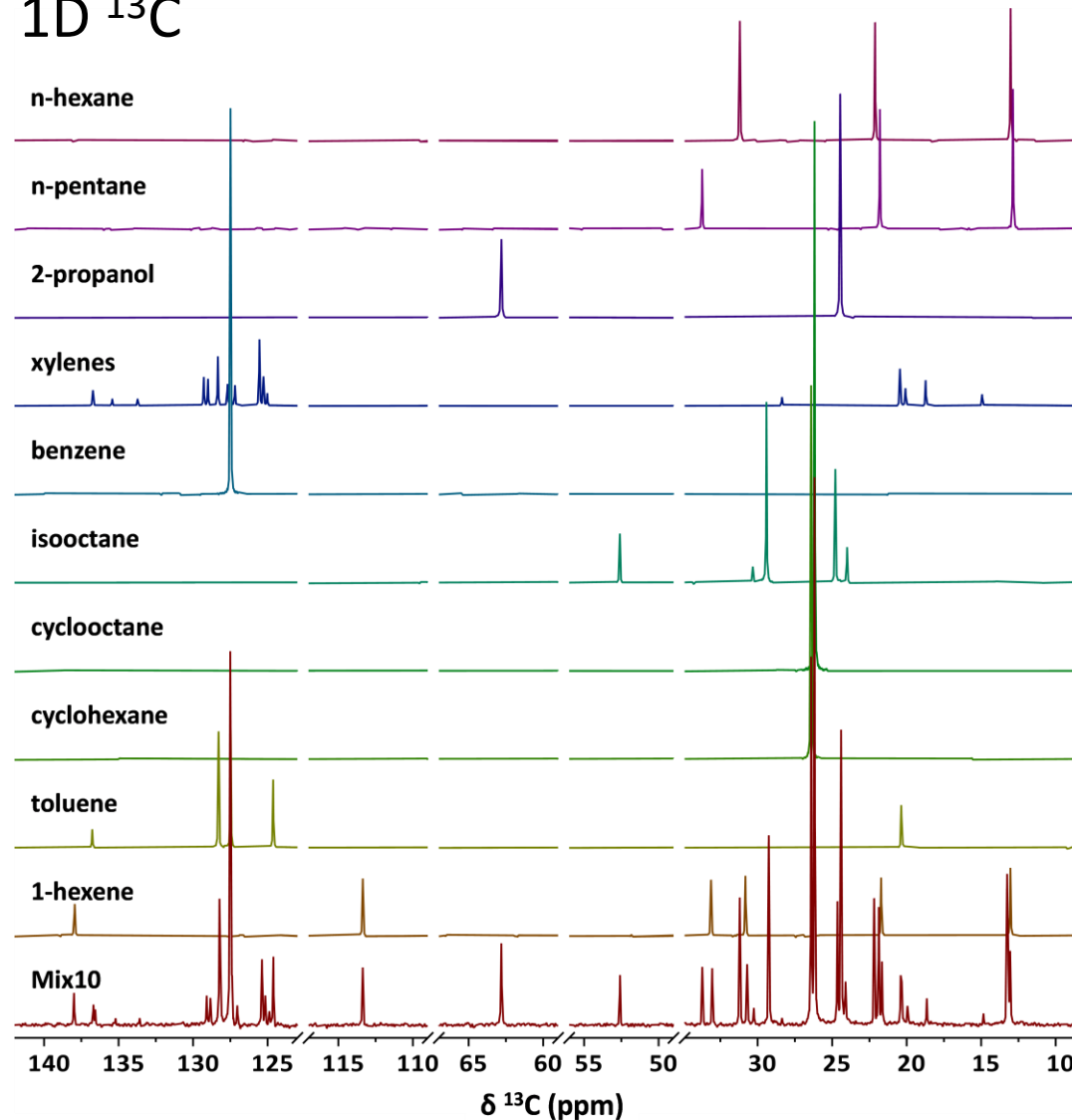
Free resources available online: <https://magritek.com/wp-content/uploads/2025/04/App-Note-Quantitation-bei-HSQC-kaurenoic-acid.pdf>



Quantification of hydrocarbon mixtures



1D ^{13}C

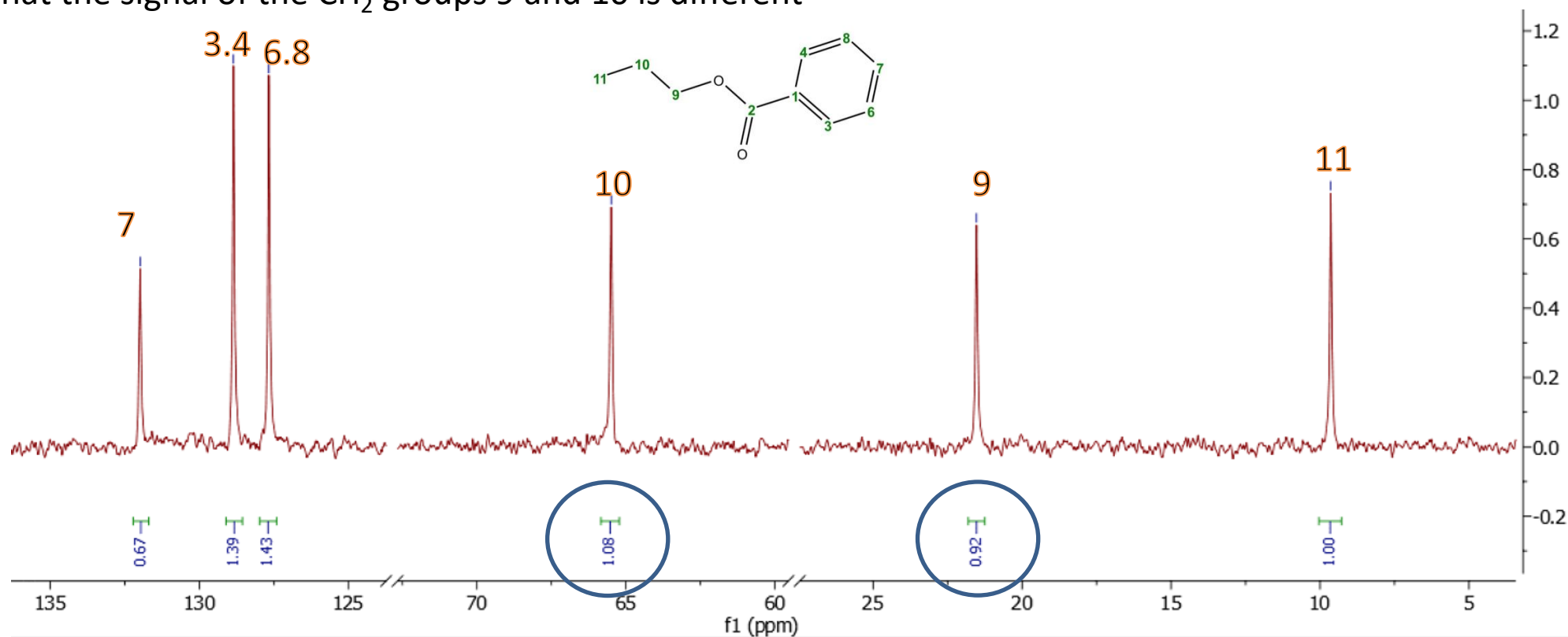


<https://magritek.com/wp-content/uploads/2022/02/App-Note-Quantification-of-single-components-in-complex-mixture-by-13C-NMR-1.pdf>

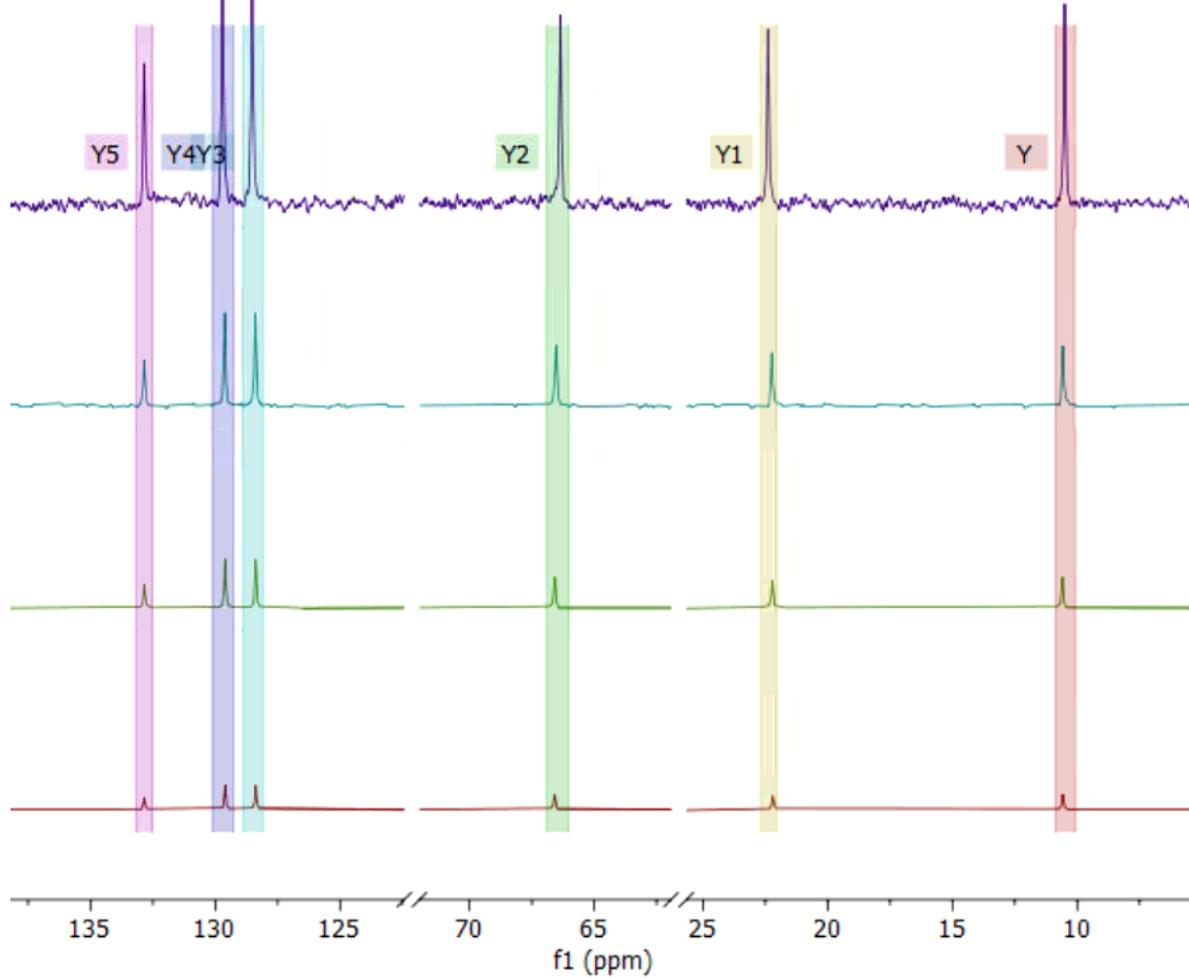


Quantification with ^{13}C NMR

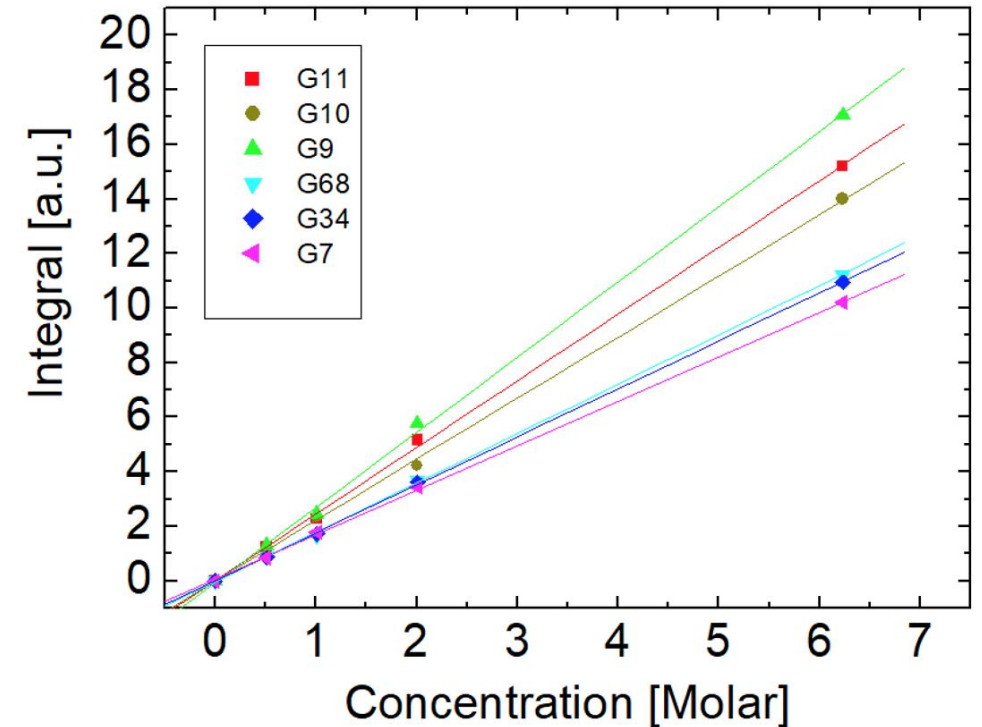
- ^{13}C can be run under so-called quantitative conditions but measurement times will be long
- In ^{13}C NMR methods like polarization transfer from ^1H to ^{13}C decouple ^1H from ^{13}C during the acquisition of the ^{13}C signal
- The integral of the carbon signals of each chemical group IS NOT proportional to the number of carbons
- This is due to the fact that the efficiency of the polarization transfer depends on the number of ^1H coupled to the ^{13}C
- **Notice** that the signal of the CH_2 groups 9 and 10 is different



The signal of each group is still proportional to the concentration

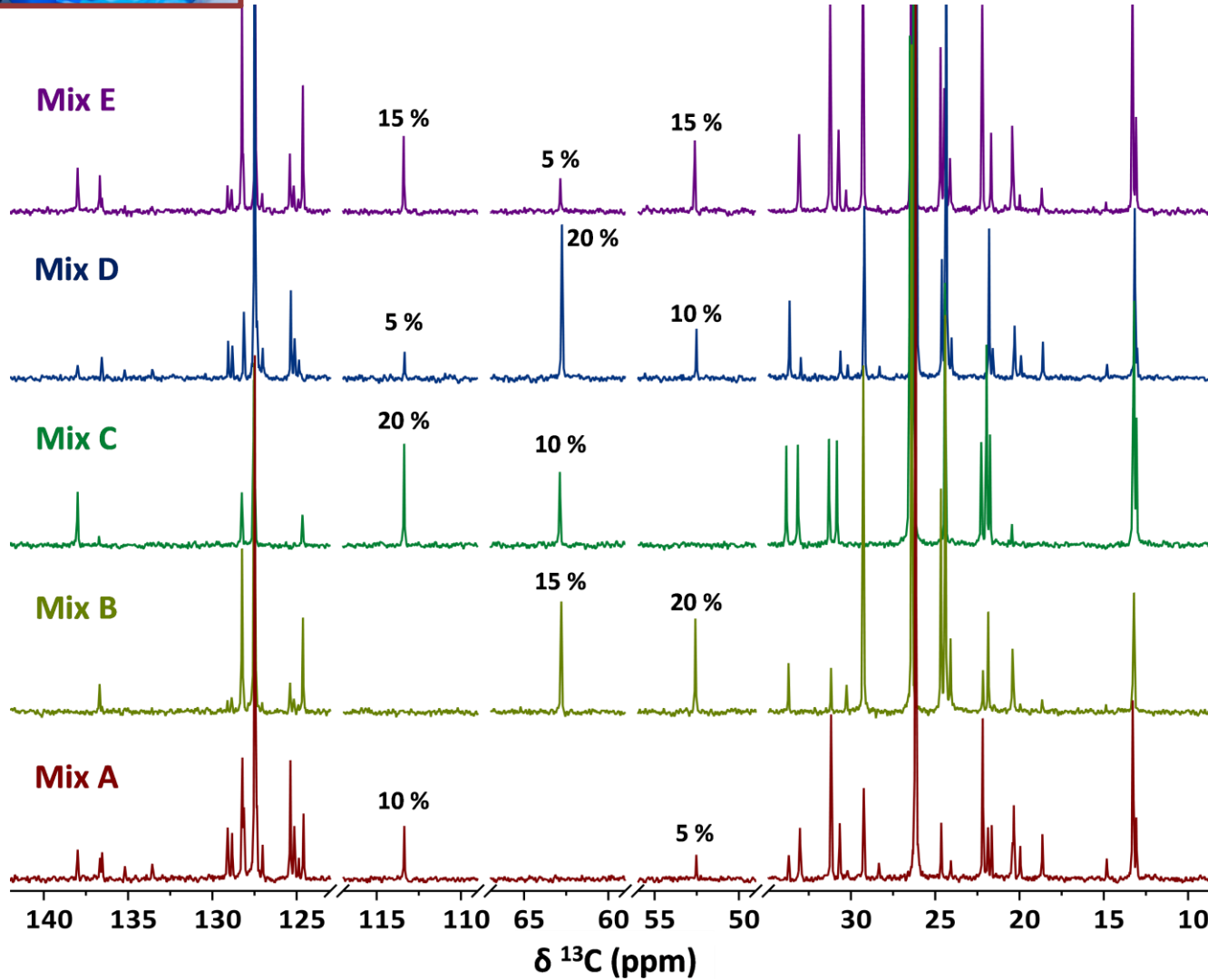


- Each carbon has a different calibration
- Even similar chemical groups like the CH2s 9 and 10 cannot be compared
- **BUT all are linear with concentration**





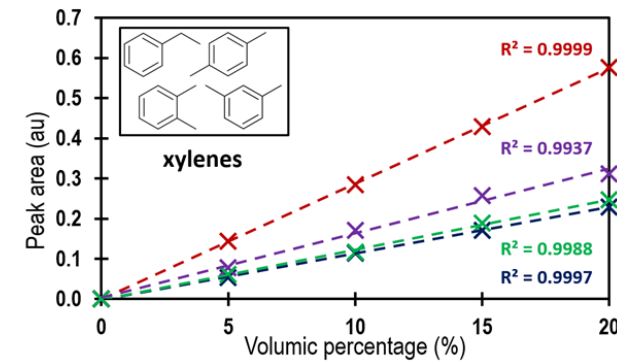
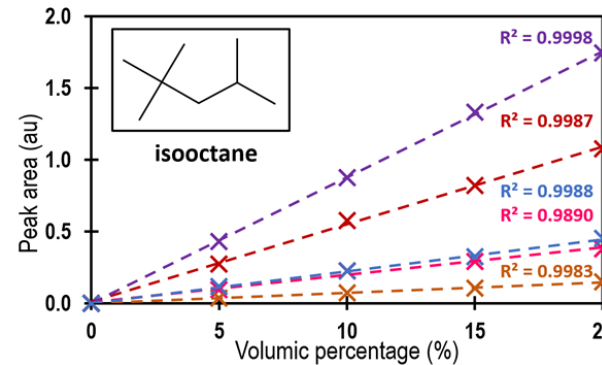
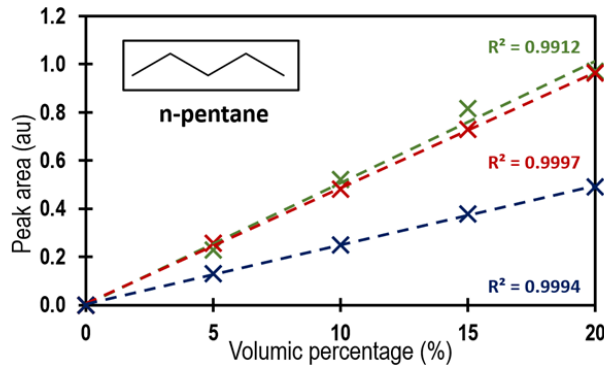
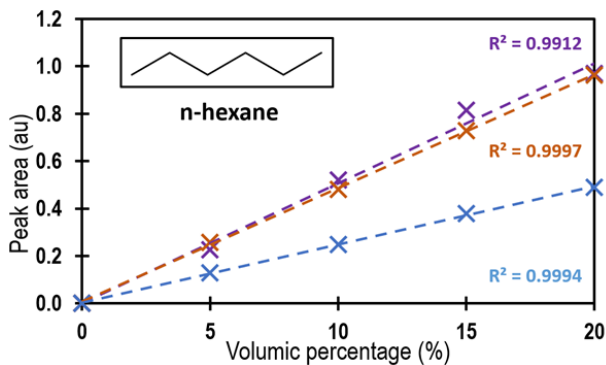
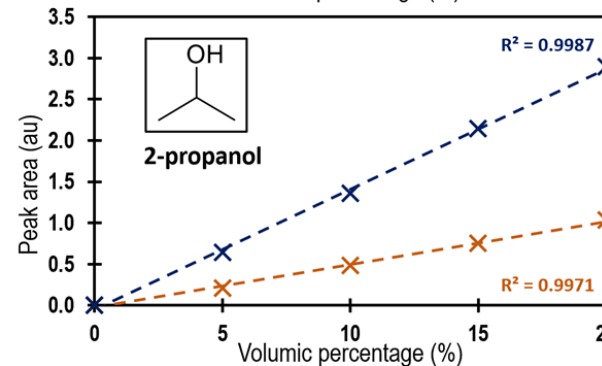
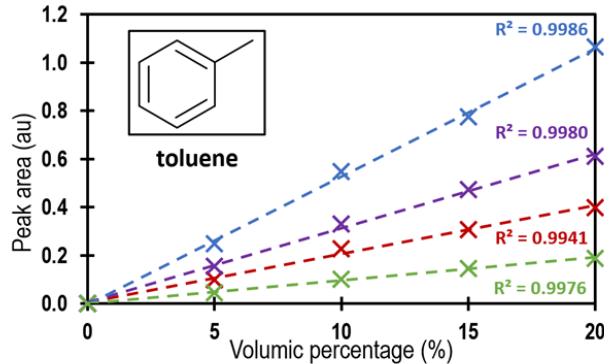
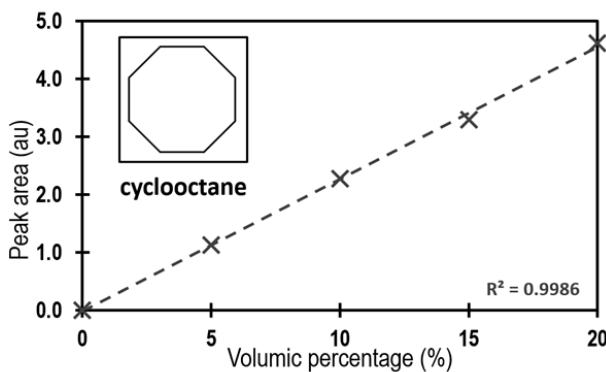
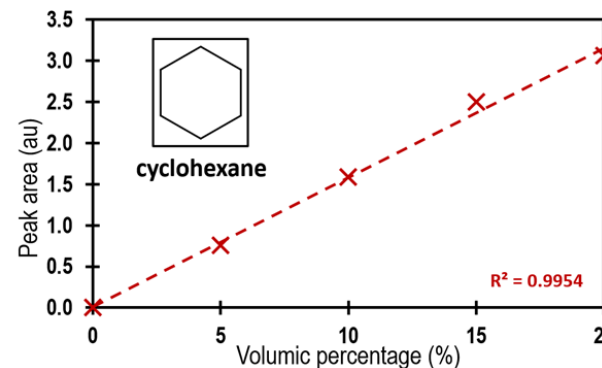
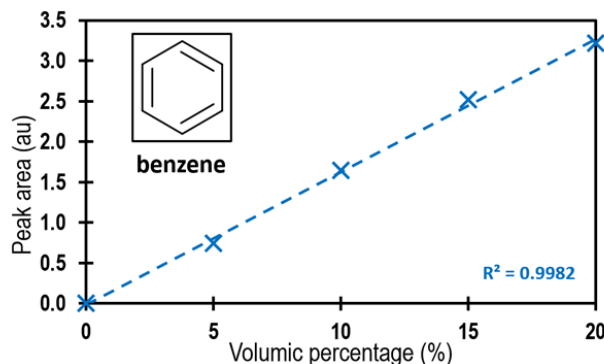
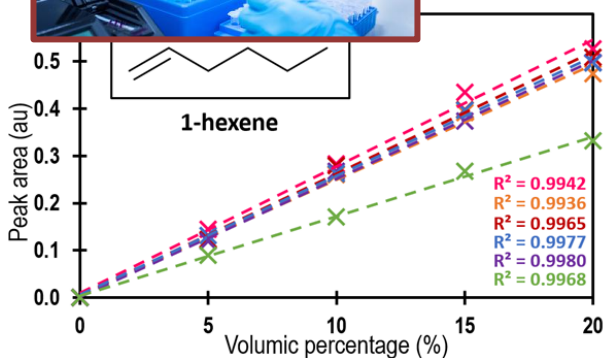
Calibration mixtures



- 5 calibration mixtures were prepared and used as external standards
- The concentration of each component was varied between samples so that for each component the whole calibration range was covered
- Spectra were acquired with NOE and decoupling
- Repetition time = 4 s; scans = 1024

	Mix A	Mix B	Mix C	Mix D	Mix E
cyclooctane	20	0	15	10	5
n-pentane	5	10	20	15	0
n-hexane	15	5	10	0	20
cyclohexane	0	20	15	5	10
isooctane	5	20	0	10	15
2-propanol	0	15	10	20	5
1-hexene	10	0	20	5	15
toluene	10	15	5	0	20
xylenes	20	5	0	15	10
benzene	15	10	5	20	0

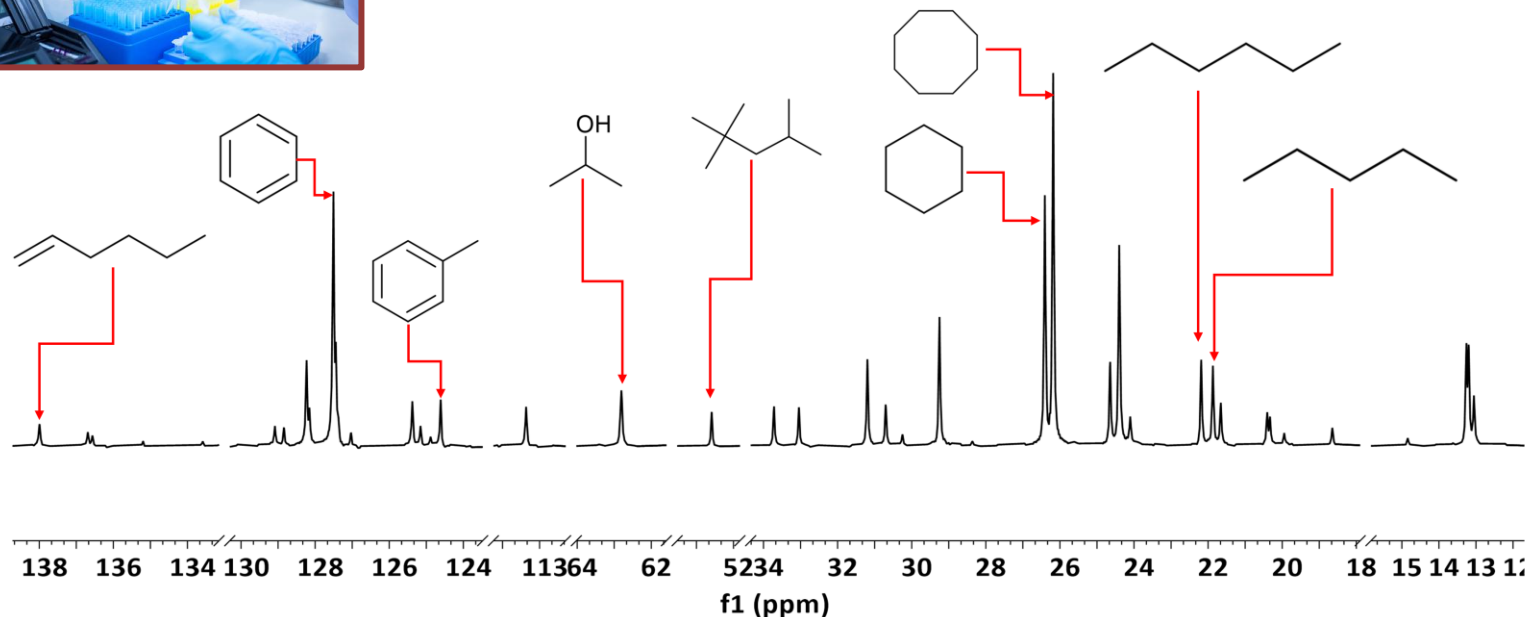
Calibration graphs



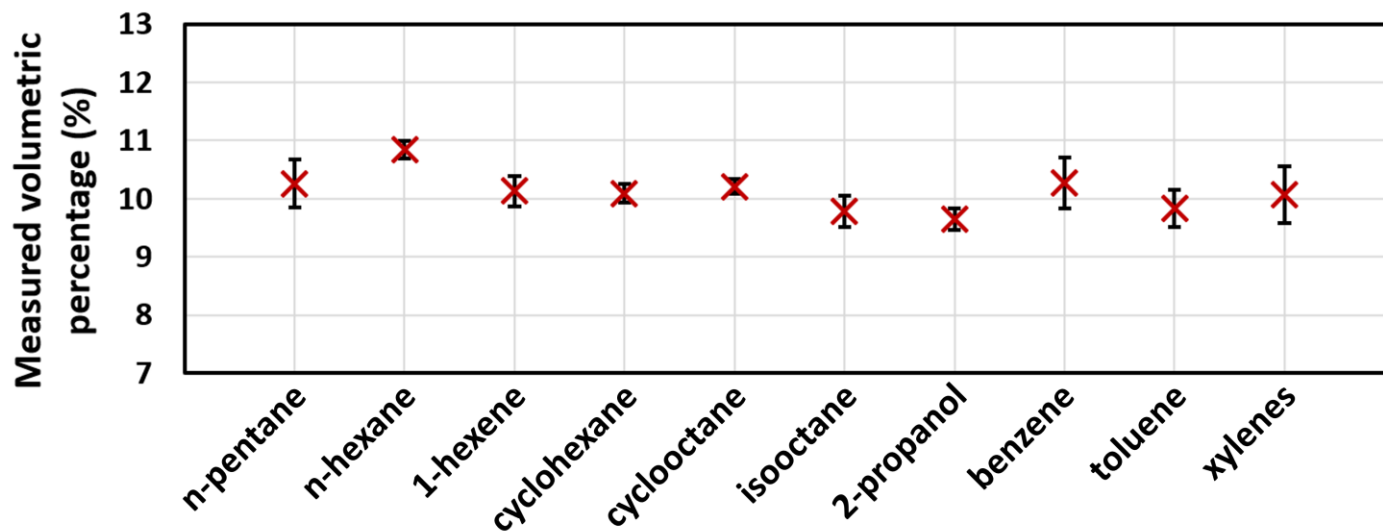
- Each component shows at least one peak that can be used for quantifications
- For all components a linear behavior is observed



Validation



- Mixture with 10 % of each component
- Spectrum was acquired with 1024 scans and repetition time of 4 seconds
- Standard deviation < 0.5% for all components



	v/v (% Vol)	std (% Vol)
n-pentane	10.26	0.41
n-hexane	10.85	0.14
1-hexene	10.13	0.26
cyclohexane	10.09	0.15
cyclooctane	10.21	0.12
isooctane	9.79	0.27
2-propanol	9.65	0.18
benzene	10.28	0.43
toluene	9.84	0.32
xylenes	10.06	0.49

Benchtop NMR applications



Synthesis

Pharma

Macromolecules

Education

Flow chemistry



Batteries

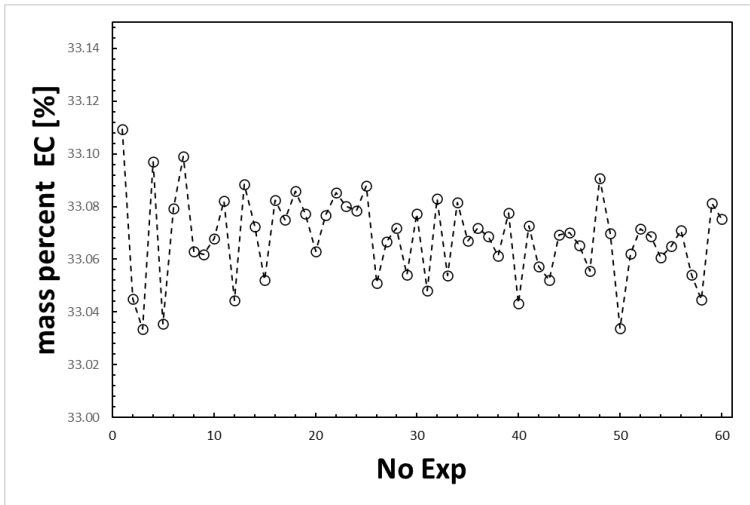
Food & Agriculture

Forensics

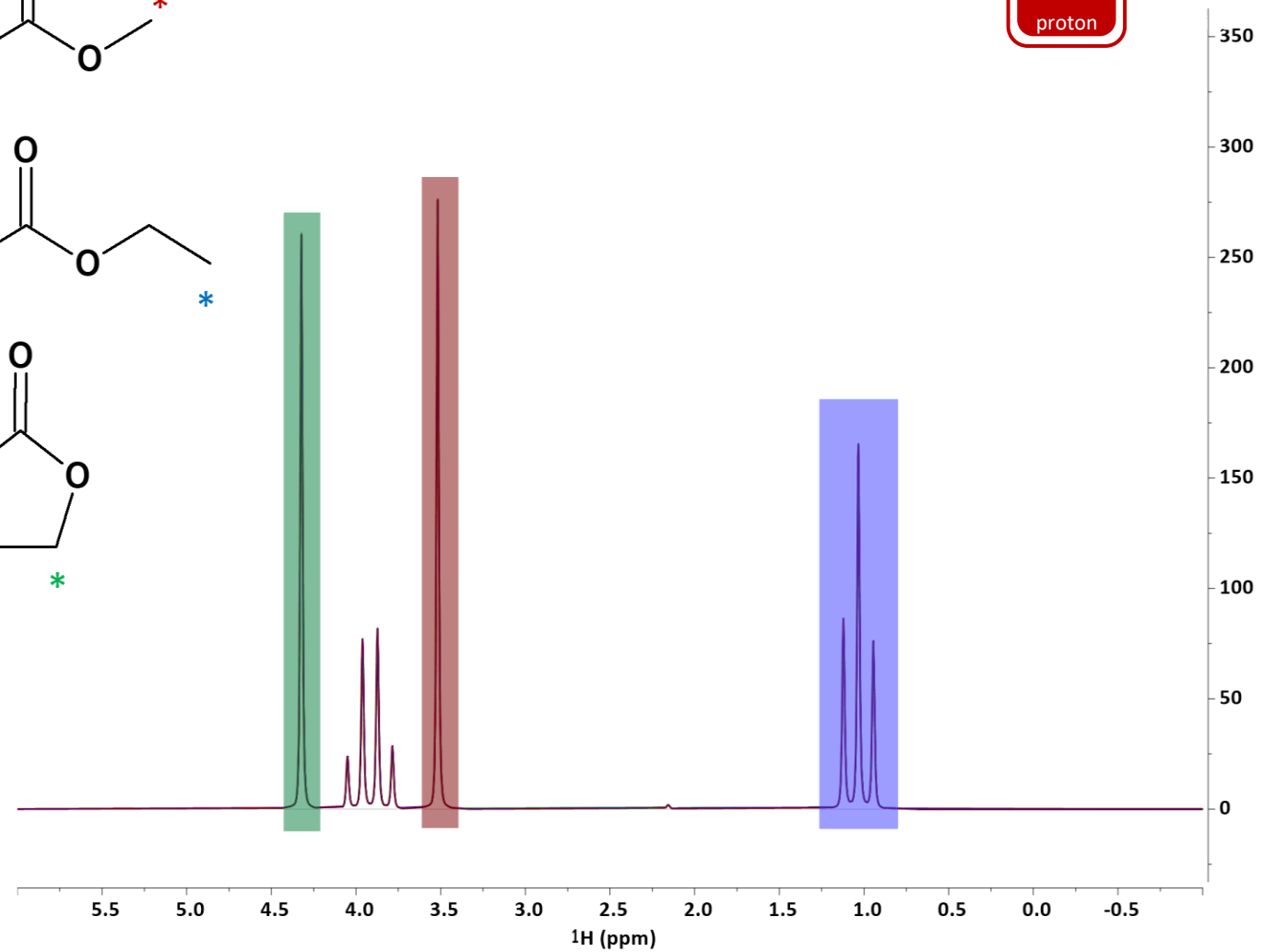
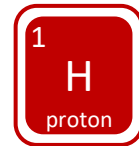
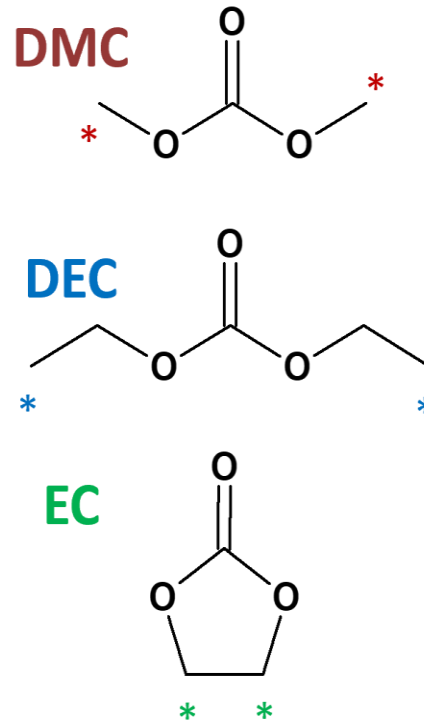
Analytics



Quantification of electrolyte components



60 consecutive measurements



Substance	wt% gravi.	wt% NMR
DMC	33.05	33.07 ± 0.02
DEC	42.91	42.95 ± 0.02
EC	24.04	23.99 ± 0.02



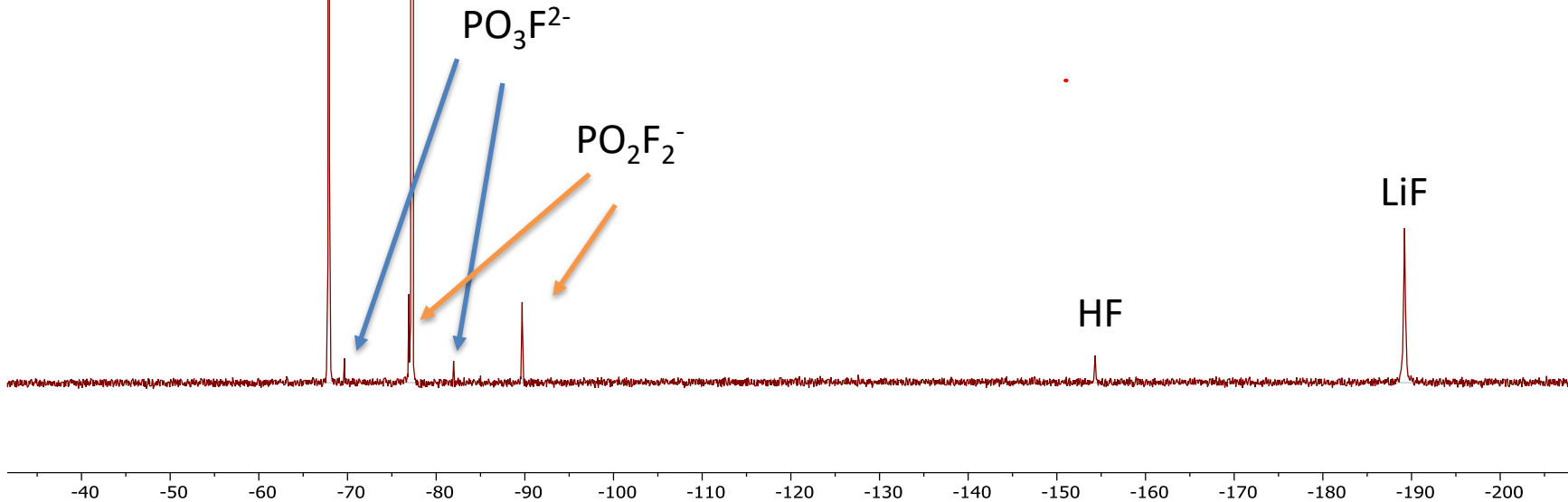
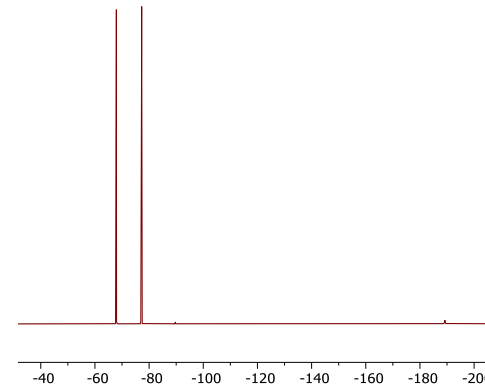
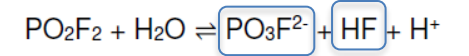
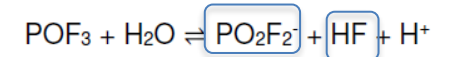
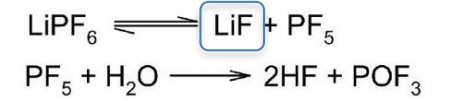
Monitoring the degradation/oxidation of PF_6^-

19
F
fluorine

PF_6^-

Single scan, 5 s

Initial hydrolysis of LiPF_6 :





Monitoring the degradation/oxidation of PF_6^-

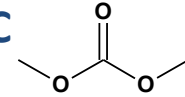
19
F
fluorine

- Addition of a few drops of H_2O to accelerate the degradation of LiPF_6

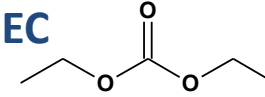
→ Formation of other species following the expected pathway



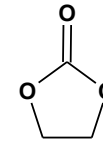
DMC



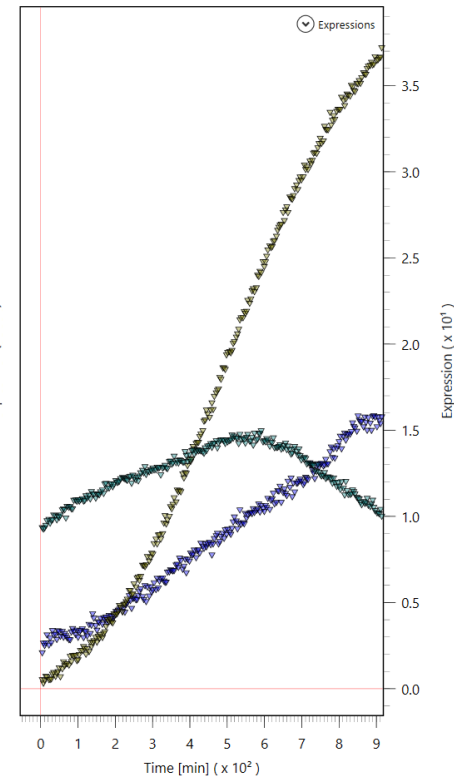
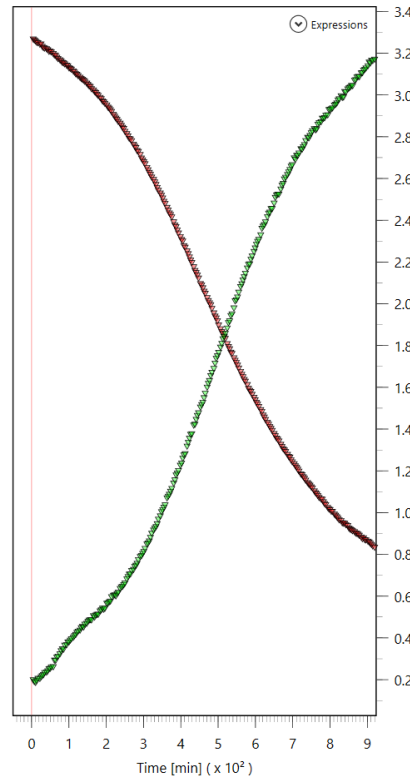
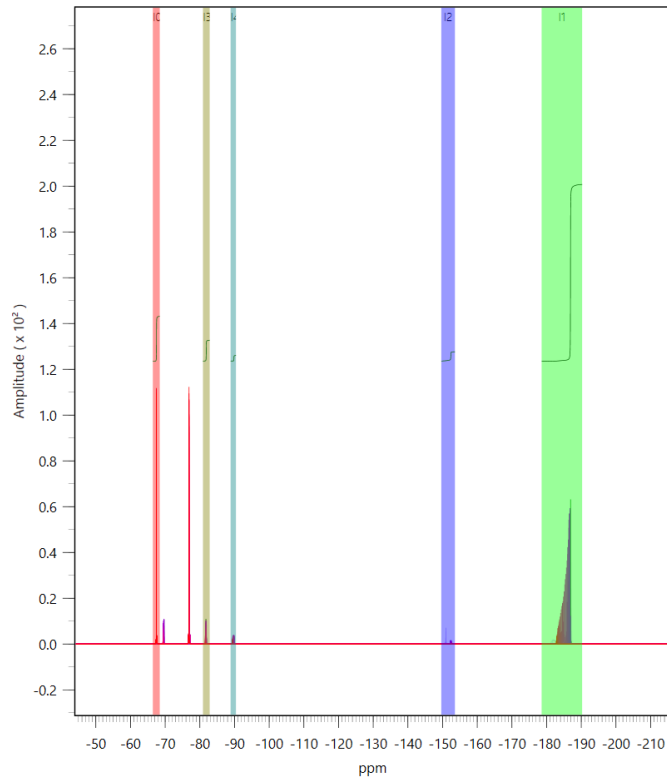
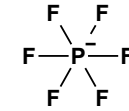
DEC



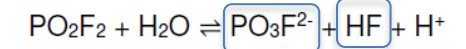
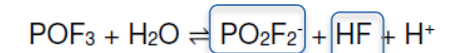
EC



Li^+

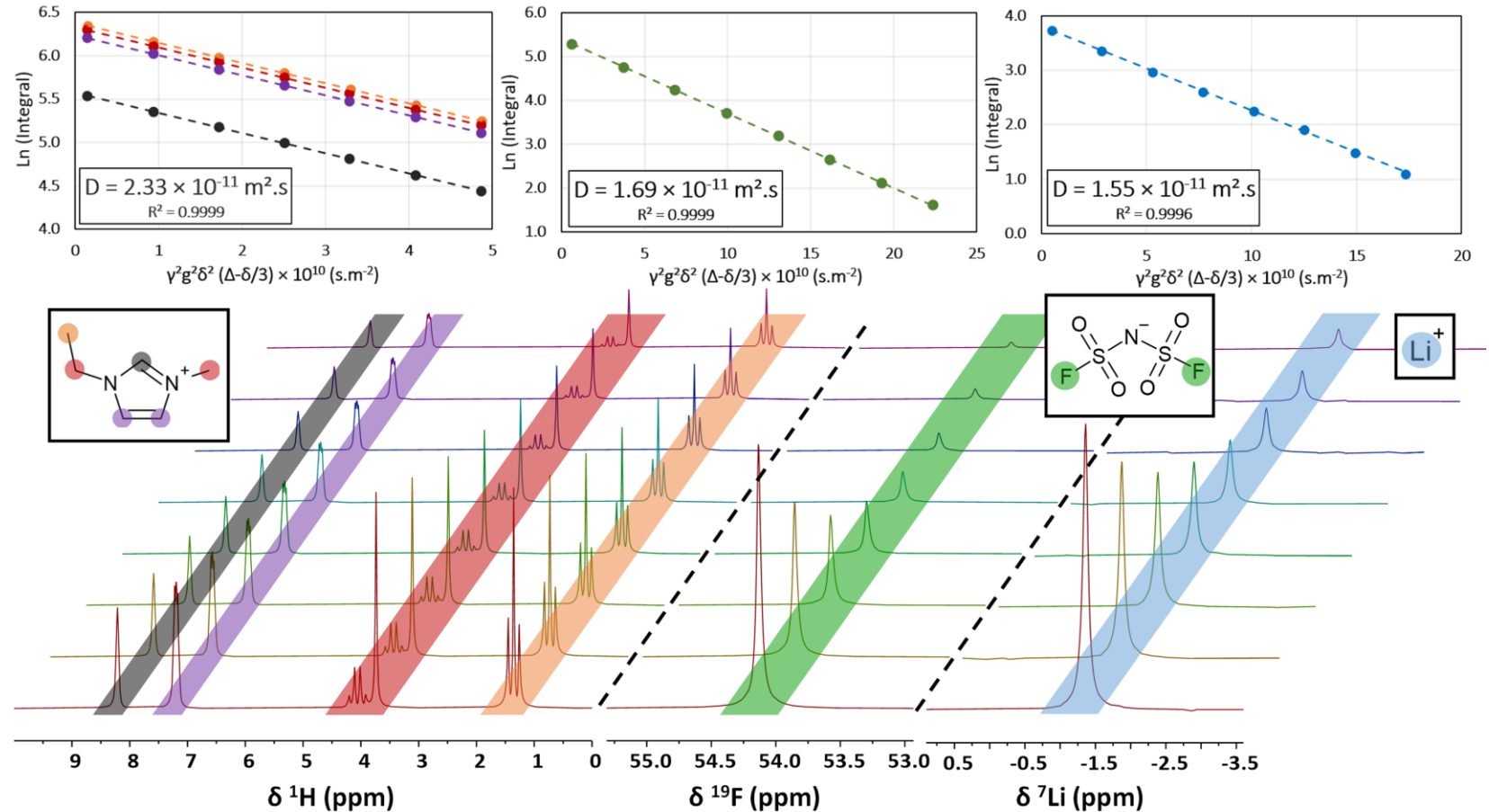
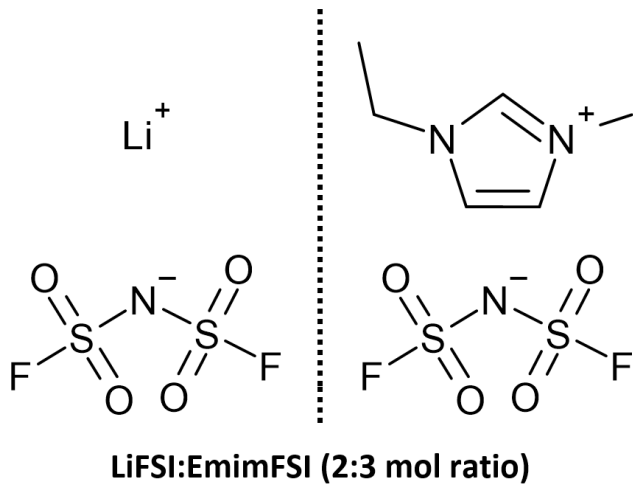


Initial hydrolysis of LiPF_6 :





Ion diffusion in electrolytes

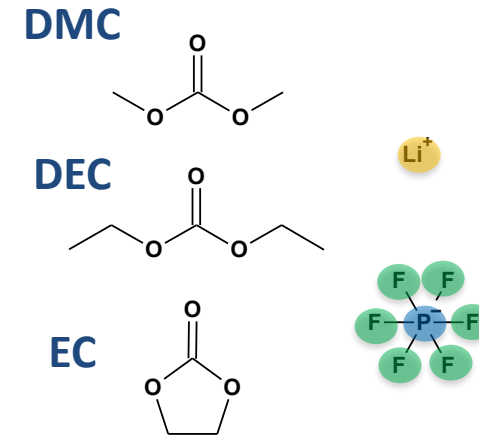
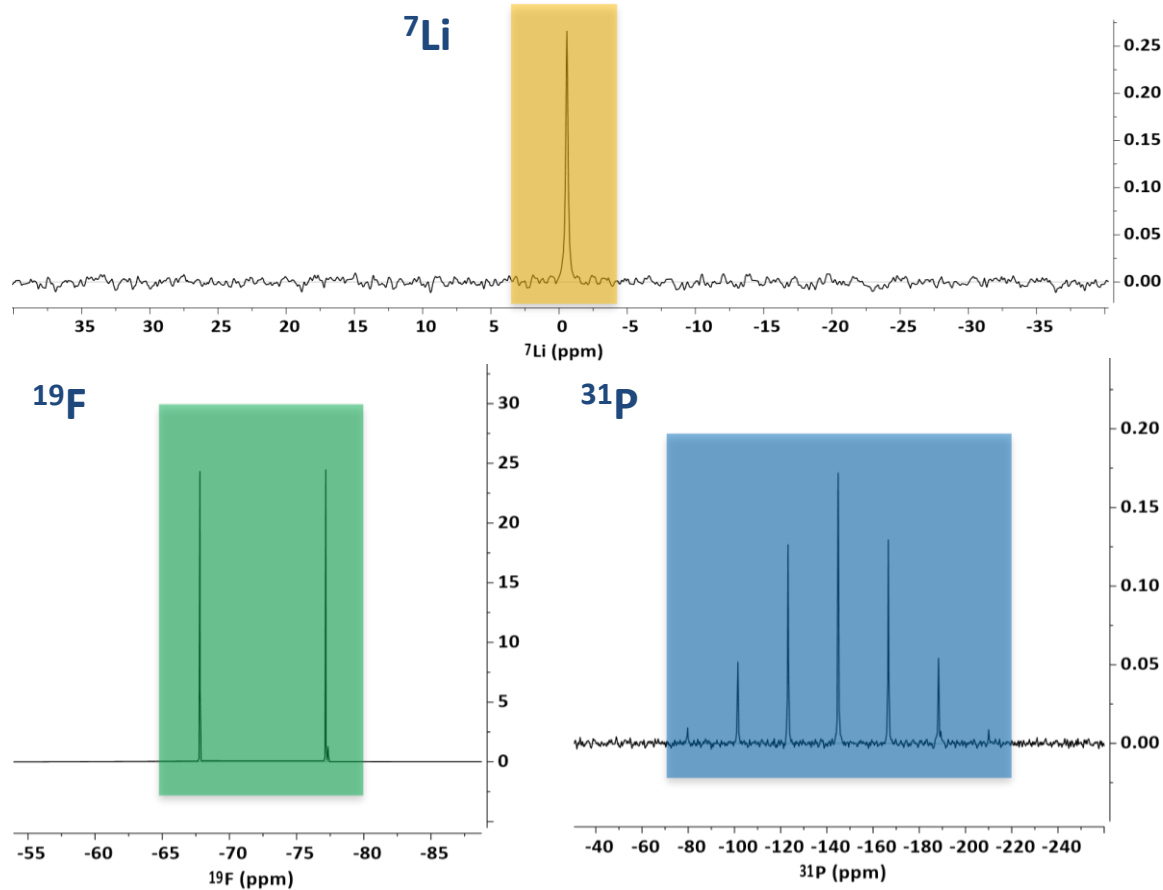


- Li-based Ionic liquid electrolyte
- Transport properties of battery components

Free online resource: <https://magritek.com/2020/09/21/study-of-transport-processes-in-li-ion-electrolytes-by-multi-nuclear-diffusion-nmr/>



Ion properties of electrolytes



• Calculation of Ion properties of electrolytes:

1) Ion conductivity σ

$$\sigma = \frac{F^2 c (D_+ + D_-)}{RT}$$

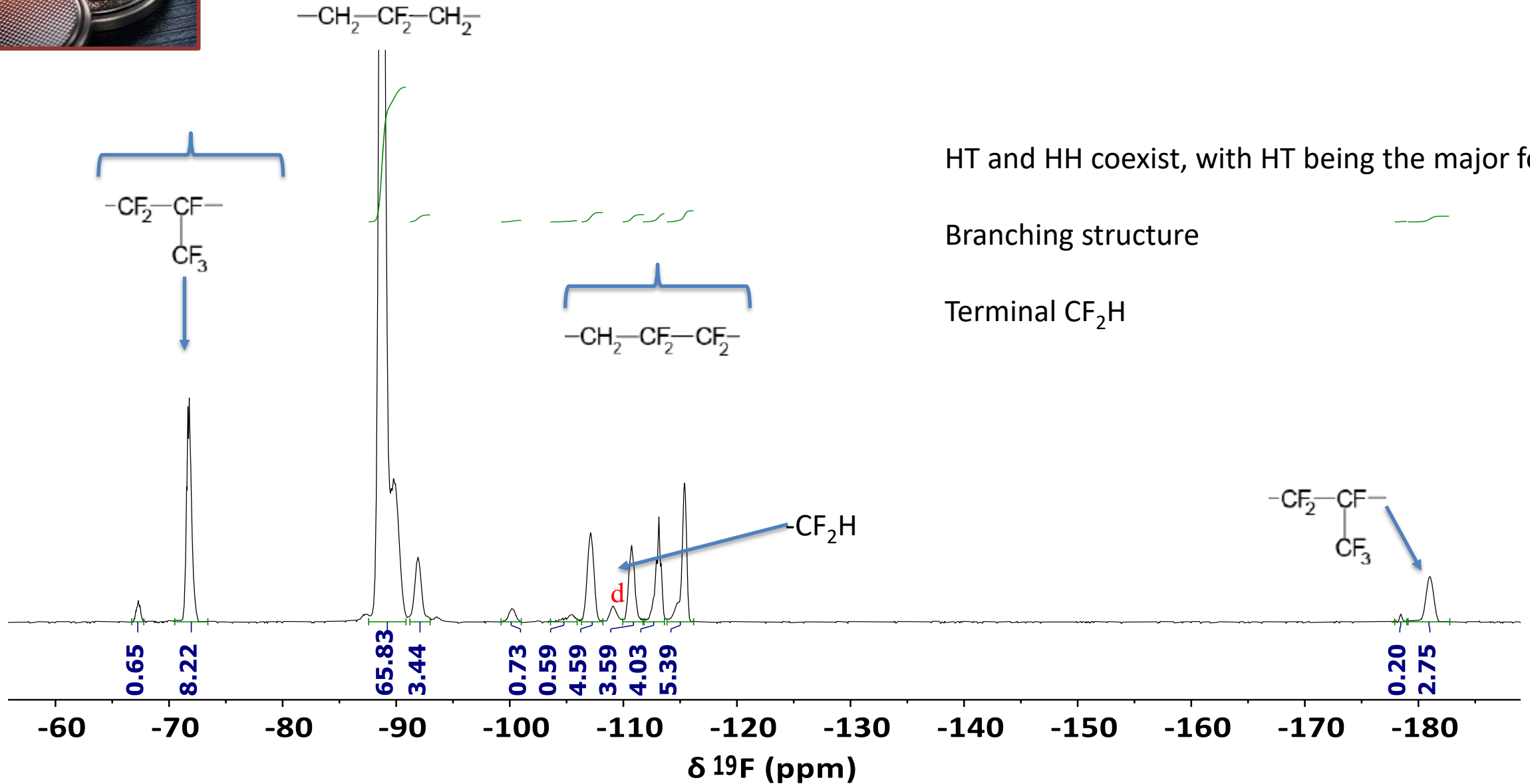
2) Transference/transport number

$$t_+ = \frac{D_+}{D_+ + D_-} \text{ and } t_- = \frac{D_-}{D_+ + D_-}$$

D+ (Li ⁺) [m ² /s]	D- (PF ₆ ⁻) [m ² /s]	Conductivity σ [S/m]	Transference t ₊	Transference t ₋
3.89 10 ⁻¹⁰	5.23 10 ⁻¹⁰	1.261	0.427	0.573



PVDF characterization



HT and HH coexist, with HT being the major form

Branching structure

Terminal CF_2H



NMR spectroscopy

- Negligible-cost consumable
- No maintenance/long lifespan (>10y)
- Low solvent volume (< 0.5 mL)
- Universal liquid detector (No-D)
- Immediate/real-time analysis
- Flow compatible
- Quantitative
- Non-destructive



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- Spinsolve Example Spectra can be found at <http://www.magritek.com/products/spinsolve/nmr-spectra-examples/>

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Standard Laboratory Setup



Reaction Monitoring Setup

